





# Zooming in the plastisphere: the ecological interface for phytoplankton–plastic interactions in aquatic ecosystems

Veronica Nava<sup>1,\*</sup> , Jaffer Y. Dar<sup>2,3</sup>, Vanessa De Santis<sup>4</sup>, Lena Fehlinger<sup>5</sup> , Julia Pasqualini<sup>6</sup> , Oloyede A. Adekolurejo<sup>7,8</sup>, Bryan Burri<sup>9</sup>, Marco J. Cabrerizo<sup>10,11</sup>, Teofana Chonova<sup>12</sup>, Mathilde Cour<sup>13</sup>, Flavia Dory<sup>1</sup>, Annemieke M. Drost<sup>14,15</sup>, Aida Figler<sup>16</sup>, Giulia Gionchetta<sup>17</sup>, Dariusz Halabowski<sup>18</sup>, Daniel R. Harvey<sup>19,20</sup>, Víctor Manzanares-Vázquez<sup>21</sup> , Benjamin Misteli<sup>22</sup>, Laureen Mori-Bazzano<sup>9</sup>, Valentin Moser<sup>23,24</sup>, Federica Rotta<sup>25,26</sup>, Bianca Schmid-Paech<sup>27</sup>, Camille M. Touchet<sup>28</sup> and Julia Gostyńska<sup>29,\*</sup>

<sup>1</sup>Department of Earth and Environmental Sciences, University of Milano-Bicocca, Piazza della Scienza 1, Milan 20126, Italy

<sup>2</sup>ICAR-Central Soil Salinity Research Institute, Karnal 132001, India

<sup>3</sup>Department of Experimental Limnology, Leibniz Institute of Freshwater Ecology and Inland Fisheries, Müggelseedamm 310, Berlin 12587, Germany

<sup>4</sup>Water Research Institute, National Research Council, Corso Tonolli 50, Verbania-Pallanza, Verbania 28922, Italy

<sup>5</sup>GEA Aquatic Ecology Group, University of Vic - Central University of Catalonia, Carrer de la Laura 13, Catalonia 08500 Vic, Spain

<sup>6</sup>Department of River Ecology, Helmholtz Centre for Environmental Research-UFZ, Brückstr. 3a, Magdeburg 39114, Germany

<sup>7</sup>Ecology and Evolution, School of Biology, University of Leeds, Leeds, LS2 9JT, UK

<sup>8</sup>Department of Biology, Adeyemi Federal University of Education, Ondo City, Ondo PMB 520, Nigeria

<sup>9</sup>Department F-A. Forel for Environmental and Aquatic Sciences, University of Geneva, 30 Quai Ernest-Ansermet Sciences II, Genève CH-1205, Switzerland

<sup>10</sup>Department of Ecology & Institute of Water Research, University of Granada, Campus Fuentenueva s/n, Granada 18071, Spain

<sup>11</sup>Estación de Fotobiología Playa Unión, casilla de correos 15, Rawson, Chubut 9103, Argentina

<sup>12</sup>Department Environmental Chemistry, Eawag: Swiss Federal Institute of Aquatic Science and Technology, Überlandstr. 133, Dübendorf CH-8600, Switzerland

<sup>13</sup>Independent Researcher

<sup>14</sup>Department of Aquatic Ecology, Netherlands Institute of Ecology, Droevendaalsesteeg 10, Wageningen 6708 PB, The Netherlands

<sup>15</sup>Department of Freshwater and Marine Ecology, Institute for Biodiversity and Ecosystem Dynamics (IBED), University of Amsterdam, P.O. Box 94240, Amsterdam 1090 GE, The Netherlands

<sup>16</sup>Department of Bioinformatics, Semmelweis University, Tűzoltó utca 7-9, Budapest 1094, Hungary

<sup>17</sup>Department of Environmental Chemistry, Institute of Environmental Assessment and Water Research (IDAEA), Spanish Council of Scientific Research (CSIC), Barcelona 0803, Spain

<sup>18</sup>Department of Ecology and Vertebrate Zoology, Faculty of Biology and Environmental Protection, University of Lodz, Banacha 12/16, Lodz 90-237, Poland

<sup>19</sup>Lake Ecosystems Group, UK Centre for Ecology & Hydrology, Lancaster Environment Centre, Library Avenue, Bailrigg, Lancaster, LA1 4AP, UK

<sup>20</sup>Lancaster Environment Centre, Lancaster University, Lancaster, LA1 4YQ, UK

<sup>21</sup>Department of Research and Development, Cocosphere Environmental Analysis, C/Cruz 39, 29120 Alhaurín el Grande, Málaga, Spain

<sup>22</sup>WasserCluster Lunz - Biologische Station, Dr Carl Kupelwieser Promenade 5, Lunz am See 3293, Austria

\* Authors for correspondence: V. Nava (Tel.: +390264482705; E-mail: [veronica.nava@unimib.it](mailto:veronica.nava@unimib.it)) and J. Gostyńska (Tel.: +48618295761; E-mail: [julia.gostynska@amu.edu.pl](mailto:julia.gostynska@amu.edu.pl)).

<sup>23</sup>Community Ecology, Swiss Federal Institute for Forest, Snow and Landscape Research WSL, Zürcherstrasse 111, Birmensdorf CH-8903, Switzerland

<sup>24</sup>Department of Aquatic Ecology, Eawag: Swiss Federal Institute of Aquatic Science and Technology, Überlandstrasse 133, Dübendorf CH-8600, Switzerland

<sup>25</sup>Department of Earth and Environmental Sciences, University of Pavia, Via Ferrata 1, Pavia 27100, Italy

<sup>26</sup>Institute of Earth Science, University of Applied Science and Arts of Southern Switzerland, Via Flora Ruchat-Roncati 15, Mendrisio CH-6850, Switzerland

<sup>27</sup>University Weihenstephan-Triesdorf of Applied Science, Am Hofgarten 4, Freising 85354, Germany

<sup>28</sup>Université Claude Bernard - Lyon 1, "LEHNA UMR 5023, CNRS, ENTPE, 3-6, rue Raphaël Dubois, Villeurbanne F-69622, France

<sup>29</sup>Department of Hydrobiology, Faculty of Biology, Adam Mickiewicz University, Uniwersytetu Poznańskiego 6, Poznań 61-614, Poland

## ABSTRACT

Phytoplankton is an essential resource in aquatic ecosystems, situated at the base of aquatic food webs. Plastic pollution can impact these organisms, potentially affecting the functioning of aquatic ecosystems. The interaction between plastics and phytoplankton is multifaceted: while microplastics can exert toxic effects on phytoplankton, plastics can also act as a substrate for colonisation. By reviewing the existing literature, this study aims to address pivotal questions concerning the intricate interplay among plastics and phytoplankton/phytobenthos and analyse impacts on fundamental ecosystem processes (e.g. primary production, nutrient cycling). This investigation spans both marine and freshwater ecosystems, examining diverse organisational levels from subcellular processes to entire ecosystems. The diverse chemical composition of plastics, along with their variable properties and role in forming the "plastisphere", underscores the complexity of their influences on aquatic environments. Morphological changes, alterations in metabolic processes, defence and stress responses, including homoaggregation and extracellular polysaccharide biosynthesis, represent adaptive strategies employed by phytoplankton to cope with plastic-induced stress. Plastics also serve as potential habitats for harmful algae and invasive species, thereby influencing biodiversity and environmental conditions. Processes affected by phytoplankton–plastic interaction can have cascading effects throughout the aquatic food web *via* altered bottom-up and top-down processes. This review emphasises that our understanding of how these multiple interactions compare in impact on natural processes is far from complete, and uncertainty persists regarding whether they drive significant alterations in ecological variables. A lack of comprehensive investigation poses a risk of overlooking fundamental aspects in addressing the environmental challenges associated with widespread plastic pollution.

**Key words:** aquatic food webs, autotrophs, epiplastic organisms, harmful algae, macroplastics, microalgae biodiversity, microplastics, primary productivity, metabolic traits.

## CONTENTS

I. Introduction	836
II. Phytoplankton–plastic– interactions: comparison of results from marine and freshwater ecosystems	837
III. The toxicity of plastics on phytoplankton and its ecological relevance	838
(1) Factors affecting the toxicity and effects of microplastics	838
(2) Responses of different taxonomic groups and ecological relevance	839
IV. Plastics as substrates and their influences on ecosystem processes	840
(1) Colonisation process	840
(2) Plastics as a substrate for harmful or invasive taxa	841
(3) The plastisphere and its impact on biodiversity	841
(4) Role of the plastisphere in nutrient provisioning and cycling	842
(5) The impact of the plastisphere on ecosystem processes	843
V. Effects on the aquatic food web	844
VI. Conclusions	845
VII. Acknowledgements	846
VIII. Author contributions	846
IX. References	846
X. Supporting information	854

## I. INTRODUCTION

Plastic pollution is a pervasive global environmental issue that impacts both marine and freshwater ecosystems (Rochman & Hoellein, 2020). Although initial research primarily focused on plastic pollution in marine environments, recent analyses have unveiled comparable or even higher plastic concentrations in freshwater ecosystems (Nava *et al.*, 2023). The ubiquity of plastic waste and its far-reaching consequences extend beyond aquatic environments, and it has been proposed as a geological indicator of the Anthropocene era (Andrady, 2022). Consequently, plastic pollution may represent a threat to biodiversity and the functioning of aquatic ecosystems, especially considering the increasing pace of global plastic production and use (Hu *et al.*, 2019; Borrelle *et al.*, 2020).

Annually, an estimated 4.8–12.7 million metric tons of plastic debris enter the oceans (Jambeck *et al.*, 2015) through numerous pathways. A large portion of plastics enters the ocean from land-based sources, including mismanaged household waste, wastewater discharge, and industrial activities. The remaining fraction is believed to originate from maritime activities, such as those involving fishing vessels and cruise ships (Rochman, 2020). In addition, atmospheric deposition (i.e. wet and dry deposition of plastic particles) is a newly recognized contributor to plastic pollution, where plastic particles are transported over long distances even to remote areas (Allen *et al.*, 2019; Brahney *et al.*, 2020).

The term “microplastic” emerged in the early 2000s and has since garnered significant global attention, primarily due to the abundance and ubiquity of microplastics, and the significant threats they pose to both human health and the environment (Sun & Wang, 2023). Microplastics can be intentionally manufactured in micrometre sizes, as seen in consumer goods such as personal care products, constituting what is known as “primary microplastics” (Frias & Nash, 2019). They can also result from the degradation of larger plastic items due to weathering, ultraviolet (UV) radiation, and mechanical forces, a category referred to as “secondary microplastics” (Cole *et al.*, 2011). Microplastics are typically categorised within a size range between 1 µm and 5 mm, although various studies have adopted different ranges (e.g. Browne *et al.*, 2011; Claessens *et al.*, 2011). This lack of consistency becomes particularly problematic when comparing microplastic data, emphasising the growing importance of establishing a scientific standard (Hartmann *et al.*, 2019). Particles with a diameter in the nanometer range are termed “nanoplastics”. Their minute size raises concerns about their potential interactions with biological processes at the cellular level as they have the capability to infiltrate cellular structures (Larue *et al.*, 2021).

The multifaceted nature of plastics poses threats to aquatic organisms across different organisational levels, from cells to populations (Scherer *et al.*, 2018). However, our current comprehension of the consequences of these pollutants remains limited, particularly regarding their impacts on ecosystem-level dynamics. Plastics do not represent a single compound or material type but rather encompass a diverse

array of chemical compositions, each characterised by specific properties, including variations in heat and chemical resistance (Andrady, 2017). Additionally, the incorporation of numerous additives during resin processing and product fabrication further adds to the complexity of these contaminating materials (Andrady & Rajapakse, 2016). Plastics can also provide substrates for various organisms since their surface can be easily biofouled when introduced into aquatic environments. This unique ecological niche is known as the “plastisphere” (Zettler, Mincer & Amaral-Zettler, 2013). This term is widely used to describe various taxa, including bacteria, microalgae, and larger organisms like molluscs, among others, associated with plastics in numerous aquatic ecosystems (Amaral-Zettler, Zettler & Mincer, 2020). The plastisphere is sometimes referred to as a human-made “eighth continent” not only offering a stable, durable, and buoyant habitat for organisms (Barros & Seena, 2021; Gao *et al.*, 2021) but also playing a central role in several biogeochemical processes. For instance, epiplastic biofilms can harbour species resistant to metals or antibiotics, facilitate horizontal gene transfer, drive species evolution for plastic biodegradation, and serve as vectors for the transport of alien species (Dąbrowska, 2021; Du *et al.*, 2022; Rani-Borges, Moschini-Carlos & Pompêo, 2021; Leite *et al.*, 2022).

The initial observations of bacterial and diatom colonisation on plastic debris date back to the 1970s (Carpenter & Smith, 1972). Early studies of the plastisphere focused on microscopically identifying the microorganisms inhabiting such “environment” (Amaral-Zettler *et al.*, 2020). More recently, studies have investigated not only the biodiversity of this habitat through next-generation sequencing-based methods (Bakal *et al.*, 2019) but also delved into the evolution and succession of their communities, trophic interactions, metabolism, and the influence of the plastisphere taxa on their surrounding environment (Bryant *et al.*, 2016; Casabianca *et al.*, 2021; Cheng *et al.*, 2021; Hope *et al.*, 2021). Prior research has predominantly focused on bacteria, often overlooking eukaryotes, particularly microalgae, despite their being significant components of the biofilm community within the plastisphere (Barros & Seena, 2021; Nava & Leoni, 2021; Xianbiao *et al.*, 2023). Consequently, despite the increasing attention given to this subject, an in-depth comprehension of the ecological impact of plastics on marine and freshwater phytoplankton/phytobenthos is still lacking.

In this review, we explore the intricate interplay between plastics (both micro- and macroplastics) and phytoplankton/phytobenthos, addressing pivotal questions about their influence on fundamental aquatic ecosystem processes, such as productivity and nutrient cycling. We analyse and compare results from marine and freshwater ecosystems across different organisational levels, from subcellular processes to entire ecosystems. Our investigation encompasses the effects of plastics of different sizes, including both the potential direct toxic effects of smaller plastics on phytoplankton and the role of larger plastics as surfaces for colonisation (i.e. plastisphere research). We aim to provide insights into the following research questions and knowledge gaps: (i)

how do phytoplankton–plastics interactions occur, and what are their implications for the broader ecological community, particularly regarding biodiversity and impacted functions; (ii) to what extent are ecosystem processes and metabolic functions affected by the interaction between (micro)plastics and phytoplankton; and (iii) how do the effects mediated by phytoplankton and plastics affect the food web *via* bottom-up and top-down processes?

## II. PHYTOPLANKTON–PLASTIC–INTERACTIONS: COMPARISON OF RESULTS FROM MARINE AND FRESHWATER ECOSYSTEMS

A literature review of *Web of Science* publications from 1992 to 2022 (see online Supporting Information, Appendix S1, for search methodology, and Table S1 for list of included studies) showed that research on the interaction between plastics and phytoplankton has predominantly concentrated on marine ecosystems, representing about 60% of the identified publications (Fig. 1A).

Most studies examined the toxic effects of plastics on phytoplankton, primarily focusing on individual species (approximately 70% of all publications retrieved; Fig. 1B). In marine studies, diatoms were the most frequently investigated group (e.g. species of *Chaetoceros*, *Thalassiosira*, *Phaeodactylum*

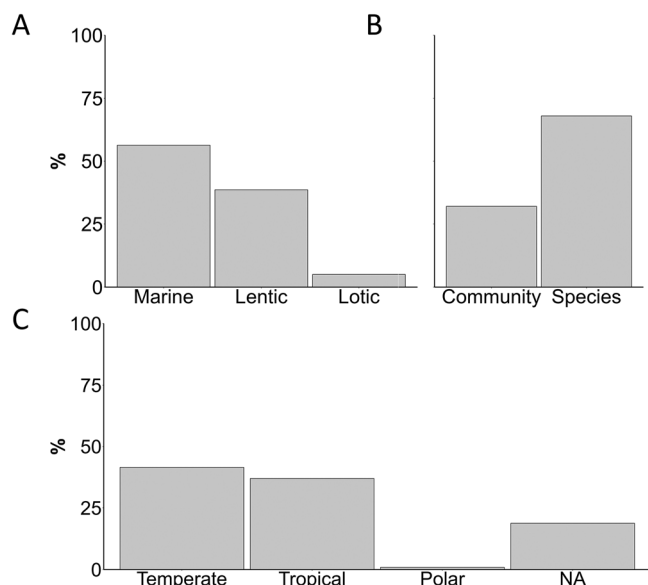
*tricornutum*), followed by green algae (e.g. species of *Dunaliella* or *Tetraselmis*), haptophytes (e.g. *Ochromonas* spp.), dinoflagellates (e.g. *Alexandrium* spp.), and cyanobacteria (e.g. *Prochlorococcus* spp.). In freshwater studies, the species investigated most frequently were cyanobacteria, with *Microcystis aeruginosa* being a prominent example, followed by various species of green algae (e.g. species of *Chlorella*, *Chlamydomonas*, and *Scenedesmus*).

Of all the studies considered, approximately 80% in total focused on species or communities from temperate (42%) and tropical (39%) environments (Fig. 1C). Surprisingly, in around 20% of studies, no explicit reference was made to the environment from which the taxon was isolated or in which the research was conducted. To date, only two studies have explored the effects of plastics in polar environments or on polar organisms: Antarctic microbial biofilms (Caroppo *et al.*, 2022) and arctic cyanobacteria (Xin *et al.*, 2022).

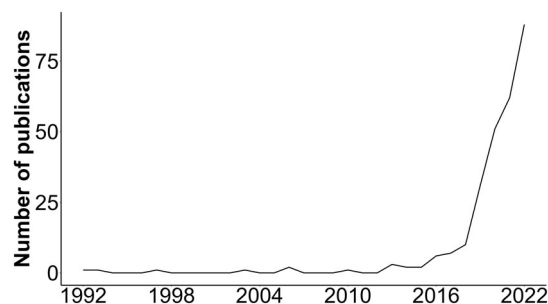
In addition to the general patterns described above, over the last decade there has been a notable increase in the number of publications (Fig. 2). The proportion of studies focusing on freshwater ecosystems has risen from none in 2013 to 43% of the total in 2022, and the proportion on tropical ecosystems has also increased (from none in 2013 to 39% of the total in 2022). Additionally, there has been an increase in studies on individual species rather than communities (from 50% in 2013 to 72% of the total in 2022).

Different toxic effects have been reported for (micro)plastics in studies of freshwater and marine phytoplankton. For example, the presence of microplastics inhibited photosynthesis in the freshwater microalga *Chlamydomonas reinhardtii* while promoting it in the marine microalga *P. tricornutum* (Li *et al.*, 2020; Chen *et al.*, 2022). Similarly, growth of freshwater *Chlorella* sp. was reduced, whereas the marine *P. tricornutum* showed adaptive capacity, showing a positive response when exposed to the same type of microplastics (Song *et al.*, 2020). It remains challenging to determine whether these differences are associated with variations in environmental conditions or species-specific characteristics.

When examining the colonisation of phytoplankton on plastic surfaces, distinct patterns emerge between freshwater and marine environments, due to differences in nutrient availability, water temperature, dissolved oxygen, salinity



**Fig. 1.** Percentage of studies ( $N = 244$ ) retrieved from *Web of Science* covering the period 1992–2022 that examine the relationship between (micro) plastics and phytoplankton (A) in freshwater (i.e. lentic and lotic) and marine ecosystems, (B) at the community or species level, and (C) in different biomes. NA denotes data not available. For the query string used, see Appendix S1. The included studies are provided in Table S1.



**Fig. 2.** Number of publications examining the effects of (micro) plastics on phytoplankton per year identified by our literature research.

levels, and microbial communities (Harrison, 2018; Barros & Seena, 2021; Miao *et al.*, 2021) influencing the establishment and development of phytobenthos communities on plastics. Consequently, distinct processes are observed, and overall patterns may differ across environments. Compared to the generally more stable temperature conditions in marine habitats, freshwater ecosystems, particularly in smaller water bodies like ponds, experience rapid temperature fluctuations (Woodward, Perkins & Brown, 2010) that can influence the development of biofilms. In water bodies with higher temperatures, biofilms can be thicker, with higher levels of chlorophyll *a*, extracellular polymeric substances (EPS), and total phosphorus content, while lower temperatures have the opposite effect (Zhao *et al.*, 2018). Salinity level also determines the types of organisms that can grow on the surface of plastic debris: in marine environments, communities tend to be dominated by species adapted to higher salinities, whilst in freshwater, less-halotolerant species are more common (Lauritano *et al.*, 2020). Furthermore, flow dynamics play a crucial role, impacting both the quantity and composition of the epiplastic community. Lentic environments, characterised by still or slowly moving water (e.g. lakes and ponds), permit nutrient accumulation in the water (Buffagni, 2021). These conditions favour the rapid proliferation of phytoplankton, often resulting in algal blooms. Conversely, lotic ecosystems (e.g. rivers and streams) are characterised by continuous water flow, which ensures variations of oxygen and temperature. This flow dynamic creates a less-favourable environment for extensive growth of phytobenthic organisms (Schneider & Petrin, 2017).

### III. THE TOXICITY OF PLASTICS ON PHYTOPLANKTON AND ITS ECOLOGICAL RELEVANCE

#### (1) Factors affecting the toxicity and effects of microplastics

Numerous studies have documented the interactions between plastics and phytoplankton in aquatic environments, hinting at potential toxicity effects (e.g. Khoironi, Anggoro & Sudarno, 2019; Nava & Leoni, 2021). These interactions are influenced by various characteristics of the plastics, including their shape, size, polymer density, and chemical composition (Chen *et al.*, 2020b; Liu *et al.*, 2021b). Several attributes of microplastics have the capacity to affect their buoyancy, transport, and distribution, consequently modifying vertical fluxes and impacting the exposure of phytoplankton to these particles (Kooi *et al.*, 2017; Eich, Weber & Lott, 2021).

Particle size has a pivotal role in determining toxic effects of microplastics, and the literature documents various size-dependent consequences. As detailed by Chen *et al.* (2020b), small plastic particles (1–2  $\mu\text{m}$ ) can be readily adsorbed to or internalised by cell walls, reducing phytoplankton growth through the mechanical inhibition of

nutrient uptake and gas exchange (Liu *et al.*, 2020; Prata *et al.*, 2019). Nanosized plastics adsorbed to the surface of phytoplanktonic organisms may have a shading effect that reduces photosynthetic efficiency and growth (Zhang *et al.*, 2022a). Liu *et al.* (2020) found that the presence of 2  $\mu\text{m}$  microplastics, in addition to causing cell membrane damage, is linked to increased cellular stress (i.e. reactive oxygen species, ROS).

Research has predominantly focused on specific polymers, such as polyethylene terephthalate (PET) and polypropylene (PP), and has revealed a negative correlation between increasing microplastic concentrations and phytoplankton growth (Khoironi *et al.*, 2019). Dose-dependent inhibition of phytoplankton growth has been also observed in the presence of nano- and micro- polystyrene (PS) beads, due to direct damage of the cell membrane and increased levels of ROS (Xiao *et al.*, 2020). In addition, PS has been found to alter the expression of genes involved in various physiological functions, including ATP synthesis, consequently influencing cell metabolic activity (Zhou *et al.*, 2021). Polyvinyl chloride (PVC) and polyethylene (PE) act by significantly reducing chlorophyll content and photochemical efficiency ( $F_v/F_m$ ) of photosystem II (Wang *et al.*, 2020b; Senousy *et al.*, 2023).

The toxic impacts of plastics on phytoplankton are not solely connected to the polymers themselves but also to the leachates they produce. It has been reported that some leachates (e.g. acetophenone, fluoranthene, dioctyl phthalate, or zinc) can inhibit the growth of certain species of cyanobacteria (Capolupo *et al.*, 2020). The extent of inhibition depends on the cell size (surface/volume ratio), morphology, physiology, and the quantity of leachate (Fernández-Juárez *et al.*, 2021; Fu *et al.*, 2019; Larue *et al.*, 2021). Tetu *et al.* (2019) investigated the effect of exposure to high-density PE and PVC leachate on the marine cyanobacterial genus *Prochlorococcus* and identified effects on growth and photosynthetic capacity, resulting in genome-wide transcriptional changes. Capolupo *et al.* (2020) investigated the effects of plastic leachate on freshwater (*Raphidocelis subcapitata*) and marine (*Skeletonema costatum*) microalgae. Almost all components of the leachate (e.g. benzothiazole, acetophenone, lead) inhibited the growth of algae. Biofilm formation on plastic surfaces (see Section IV) may influence the leaching of additives, although how the dynamics of leaching are affected by biofilms is still unclear. Research has identified a “cover effect”, where the biofilm forms a barrier on the surface of the plastics, potentially altering the kinetics of chemical exchange (Peng *et al.*, 2023; Binda *et al.*, 2024).

In addition, microplastics have the capacity to adsorb organic and inorganic harmful substances from the environment. By assimilating them, microplastics effectively reduce the environmental availability of such substances, thereby mitigating damage to sensitive species (Fernández-Juárez *et al.*, 2021). For instance, Fu *et al.* (2019) showed that PVC particles can adsorb copper ions, thereby limiting their toxic effects on microalgal cells. However, it is important to note that the effectiveness of this protective role is contingent upon whether or not these microplastics are subsequently absorbed



by microorganisms in the ecosystem. If microorganisms do take up these plastic particles, the absorbed harmful substances may impact the organism and/or be transferred through the food web, posing potential risks to higher trophic levels (Fu *et al.*, 2019).

Morphological changes have been extensively observed when micro- and nanoplastics adhere to cell surfaces. This is particularly evident for positively charged microplastics, which adhere to the typically negatively charged cytomembrane of microalgae. As a result, these microplastics become embedded in the cell membrane (Larue *et al.*, 2021). Researchers have observed that the presence of plastic fragments can lead to deformation of chloroplasts and thylakoid membranes, leading to a reduction in cell chlorophyll content and hence diminished photosynthetic efficiency (Mao *et al.*, 2018; Prata *et al.*, 2018; Fu *et al.*, 2019). For instance, Besseling *et al.* (2014) reported a reduction in chlorophyll content in cells of *Chlorella* and *Scenedesmus* spp. when exposed to PS nanoplastics. Sjollem *et al.* (2016) revealed a 45% decrease in photosynthetic rate of *Dunaliella tertiolecta* in the presence of microplastics, whereas Yan *et al.* (2021) demonstrated the deformation and disruption of thylakoid structures in *C. reinhardtii* when exposed to plastic-induced stress. The likely cause of these effects is the surface adsorption of plastics, resulting in decreased expression of photosynthesis-related genes and metabolic disruption. Damage to photosynthesis increases the production of ROS, culminating in oxidative damage and lipid peroxidation (Prata *et al.*, 2019; Nava & Leoni, 2021). ROS hinders the synthesis of chlorophyll *a* and *b* and disrupts electron transport between primary and secondary acceptor plastoquinones, thereby diminishing the efficiency of photosystem II (Yan *et al.*, 2021). Lipid peroxidation elevates the level of malondialdehyde (MDA) within the cells, raising the permeability of the cell membrane and facilitating the uptake of plastic particles. When exposed to micro- and nanoplastics, Yan *et al.* (2021) observed elevated levels of ROS and MDA in *C. reinhardtii* cells. This increase in oxidative stress led to greater membrane permeability, which, in turn, resulted in increased plastic uptake by the cells. To defend against oxidative stress, the production of superoxide dismutase (SOD), catalase (CAT), and carotenoids increases. However, if the ROS content surpasses the cell's self-repair capacity, it can trigger apoptosis, necrosis and cell death (Prata *et al.*, 2019; Nava & Leoni, 2021; Jiazhu *et al.*, 2022). When exposed to environmentally relevant concentrations of microplastics (range 10–1000 mg/L), Yuanyuan *et al.* (2022) noted augmented production of pigments in the microalga *Chlorella vulgaris* as a defence against oxidative stress. Jiazhu *et al.* (2022) observed increased production of SOD in *Prorocentrum donghaiense* exposed to microplastics. However, this response proved insufficient to counteract the oxidative stress, as the cell membrane sustained damage due to lipid peroxidation induced by accumulating ROS. Oxidative stress can also promote the release of harmful metabolites, such as microcystin by *M. aeruginosa* (Zheng

*et al.*, 2021), as a mechanism to mitigate microplastic-induced oxidative stress (Amaneeh *et al.*, 2023).

Other defence mechanisms have been documented in microalgae in response to exposure to plastics. One common response to plastic exposure involves the homoaggregation of cells, which serves to decrease the surface area exposed to plastic particles (Yan *et al.*, 2021). Researchers have reported elevated expression of genes responsible for the biosynthesis of EPS [e.g. Yan *et al.* (2021) in *C. reinhardtii*]. EPS serve to thicken the cell wall, providing a shield against physical harm. However, microplastics can accumulate within the EPS, obstructing nutrient uptake, and limiting light availability (Prata *et al.*, 2019; Liu *et al.*, 2021b; Yan *et al.*, 2021).

Effects of microplastics can also manifest through the modification of metabolic mechanisms. Amaneeh *et al.* (2023) observed a decrease in phosphorus uptake in the cyanobacterium *Halotache* sp. when exposed to microplastics. While this decrease was likely a result of an indirect mechanism, specifically phosphate ion adsorption to the plastic particles, it highlights the far-reaching implications of microplastic exposure. Other studies have shown that the presence of microplastics and their organic additives can change phosphate homeostasis, leading to increased alkaline phosphatase activity, while reducing the rate of  $\text{PO}_4^{3-}$  uptake (Fernández-Juárez *et al.*, 2021). In addition, reduction in the quantity of oil bodies, which function as an emergency energy source for the maintenance of normal growth, has been observed in the marine diatom *Chaetoceros neogracile* under microplastic stress-induced conditions (Seoane *et al.*, 2019). A shift in lipid composition (e.g. chloroplast galactolipids) and alterations in fatty acid profile, coupled with reductions in neutral lipid content and an increase in esterase activity, were reported in *Chlorella sorokiniana* exposed to PS (Amaneeh *et al.*, 2023). Such alterations in fatty acids can have cascading effects throughout the food web (see Section V).

## (2) Responses of different taxonomic groups and ecological relevance

Plastics exert distinct effects on different groups of phytoplankton, with some beneficial and others detrimental (Rani-Borges *et al.*, 2021). Most studies show a negative effect of microplastics on both marine and freshwater phytoplankton (Gao *et al.*, 2021). The specific impact of microplastics often depends on group- or species-specific traits. For instance, some cyanobacteria can tolerate the presence of plastics better than other organisms (Fernández-Juárez *et al.*, 2021; Hitchcock, 2022). They can form microfilms or colonies around microplastics (Dussud *et al.*, 2018b; Hitchcock, 2022) and exhibit a greater capacity to cope with light limitation due to shading caused by microplastics (Sjollem *et al.*, 2016; Zhu *et al.*, 2021). Similarly, the capacity to form hetero-aggregates appears to enable certain marine diatoms to manage microplastic toxicity, as this may improve their photosynthetic efficiency and restore their growth (Long *et al.*, 2017; Wang *et al.*, 2020a). The formation of

hetero-aggregates depends on various factors, including the size of the particles (Wang & Chen, 2023) and the type of polymer. Lagarde *et al.* (2016) noted the formation of hetero-aggregates for PP but not for high-density polyethylene (HDPE) microplastics. Trait-specific effects of microplastics may also be influenced by the thickness of the cell wall, which acts as a barrier against microplastic penetration (Sjollema *et al.*, 2016; Nolte *et al.*, 2017; Parsai *et al.*, 2022). Furthermore, cell dimensions play a role, with smaller cells being more susceptible to the impact of microplastics due to their relatively higher cell surface area to volume ratio (Sjollema *et al.*, 2016; Chen *et al.*, 2020b; Ge *et al.*, 2022).

The majority of studies examining the effects of microplastics on microalgae focus on green algae and diatoms (Larue *et al.*, 2021). One important consideration is the environmental relevance of the microplastics used. Many investigations have assessed plastic toxicity using pristine spherical particles, such as spherical PS particles (Larue *et al.*, 2021). However, real-world environmental plastics exhibit considerable variation in terms of shape, size, polymer type, and surface characteristics. Furthermore, these characteristics can change as plastics age (Larue *et al.*, 2021), although it remains challenging to determine the associated implications. Further research will be essential to unravel ecologically relevant impacts.

#### IV. PLASTICS AS SUBSTRATES AND THEIR INFLUENCES ON ECOSYSTEM PROCESSES

##### (1) Colonisation process

In both freshwater and marine ecosystems, plastics are rapidly colonised by a variety of organisms, including bacteria, single-celled eukaryotic organisms, larvae, and spores (Amaral-Zettler *et al.*, 2020; Yu *et al.*, 2023). Upon coming into contact with plastic, free-living microorganisms can form biofilms to transition from a planktonic mode to a sessile mode. Diatoms and various bacteria, such as Gammaproteobacteria, Cyanobacteria, and Alphaproteobacteria, are well recognised as early colonisers (Amaral-Zettler *et al.*, 2020; Odobel *et al.*, 2021; Yu *et al.*, 2023).

During the initial phase of colonisation, pioneer bacteria produce EPS through the regulation of lipid/fatty acids and c-di-GMP (bis(3'-5')-cyclic dimeric guanosine monophosphate) signals (Su *et al.*, 2022a). The secretion of EPS enhances adhesion and facilitates further colonisation by various microorganisms, for example by members of Bacteroides (Fig. 3). These organisms are highly adaptable and promote the irreversible attachment of microflora by forming pili, adhesive proteins, and additional extracellular polymeric substances (Du *et al.*, 2022). As the process continues, different organisms upregulate gene expression related to communication, adhesion, substance transport, and chemotaxis (Solano, Echeverez & Lasa, 2014). This gives rise to a tissue-like matrix (Yu *et al.*, 2023) characterised by channels that aid in distributing nutrients between cells

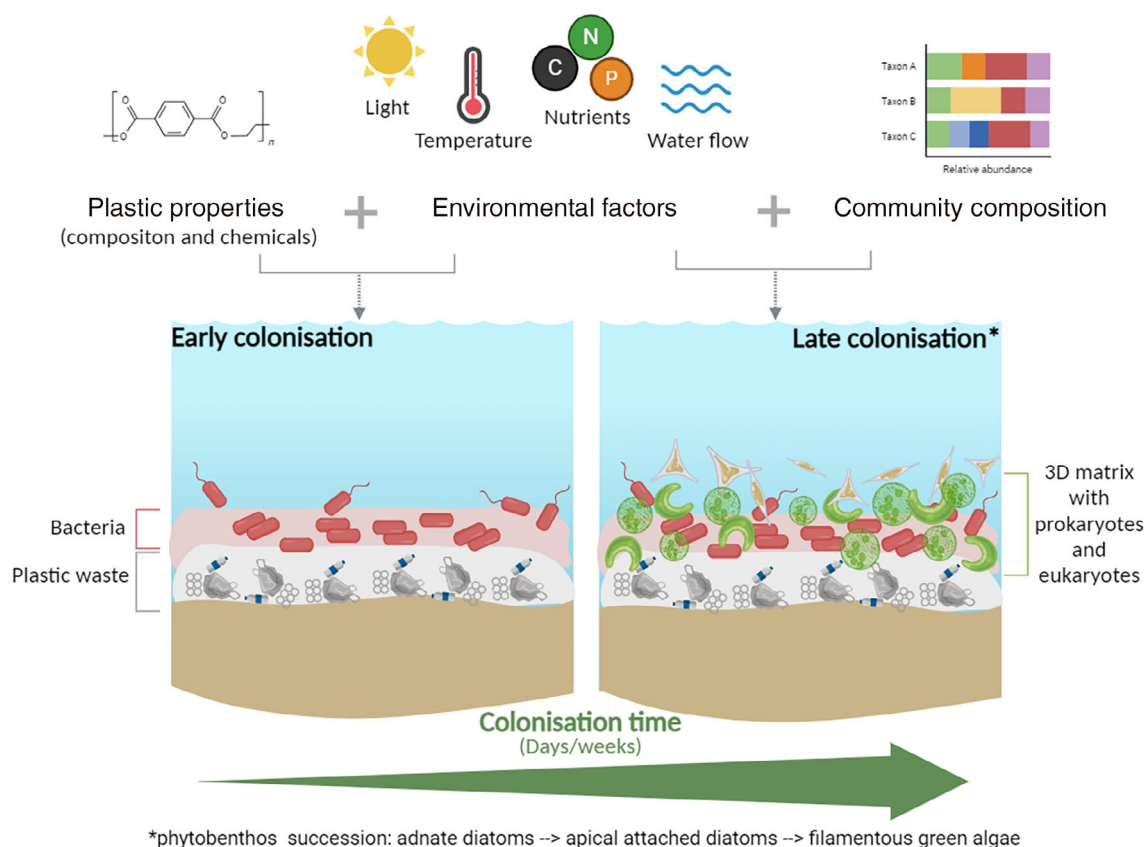
(Amaral-Zettler *et al.*, 2020). Other groups, including fungi and archaea, also become part of the biofilm, although their precise role in the colonisation process remains poorly understood (Yu *et al.*, 2023).

Over time, the biofilm structure undergoes transformation into a more complex three-dimensional (3D) structure, accommodating both heterotrophic bacteria and prokaryotic and eukaryotic autotrophs (Yokota *et al.*, 2017). Several studies have reported that the community composition on plastic surfaces differs from that of the surrounding free-living organisms, both in freshwater and marine systems (Bryant *et al.*, 2016; Odobel *et al.*, 2021; Wang *et al.*, 2022b; Sosa & Chen, 2022). However, no group of microorganisms has been exclusively observed on plastic (Amaral-Zettler *et al.*, 2020). The diversity of the biofilm community tends to increase over time, with filamentous cyanobacteria and associated heterotrophic bacteria becoming dominant while early-colonising diatoms decrease in abundance. Nevertheless, most studies report lower species richness on plastic and greater evenness than in the community in the surrounding environment (Amaral-Zettler *et al.*, 2020).

The characteristics and properties of plastic surfaces play a crucial role in determining colonisation processes. This concept broadly applies to biofilm formation across various materials, with existing literature highlighting how substrate properties influence colonisation (Vadeboncoeur *et al.*, 2006). In plastisphere research, there is substantial debate over whether substrate properties or environmental conditions are more dominant in shaping colonising communities. Many studies suggest that environmental factors are more influential in determining community composition, particularly in mature biofilms. This is likely because only the initial layers of the biofilm come into direct contact with the substrate (Rummel *et al.*, 2017; Nava *et al.*, 2024). Additionally, recent research indicates that as biofilms mature, their community structure diverges from patterns observed after shorter incubation periods, underscoring the need for studies that investigate longer-term processes in plastisphere research. Kirstein *et al.* (2019) compared communities after short-term (6 weeks) and long-term (5 months) incubation, revealing shifts towards communities with lower richness over time for all plastic types. This suggests selection for microbes specialised to low-nutrient conditions or to specific plastic types. However, few studies have evaluated longer-term colonisation, particularly regarding the autotrophic component of the community, making it difficult to determine the time required to reach peak biomass for plastic substrates and whether this differs from naturally occurring substrates.

Ageing processes can alter the properties of plastic substrates, which in turn can affect the environmental fate of plastics, their sorption characteristics, and ultimately their impact on biological communities, including planktonic organisms (Binda *et al.*, 2024).

While the available literature has provided insights into many mechanisms related to biofilm colonisation on plastic substrates in marine and freshwater environments, many aspects remain



**Fig. 3.** Schematic representation of the colonisation process of plastic surfaces over time, from initial colonisation by pioneer species to later stages. Early colonisation involves microorganisms embedded in an exopolysaccharides (EPS) matrix. The process is influenced by various properties of the plastic substrates, such as texture, polymer composition, and colour. As the community evolves over time, the matrix becomes more complex, developing a three-dimensional (3D) structure that supports a variety of organisms. This includes the early appearance of adnate diatoms (Bacillariophyceae), followed by the growth of filamentous organisms such as cyanobacteria and Chlorophyceae.

unresolved. Further investigation is required, for example, on how light, nutrient availability or temperature modulate the colonisation by microorganisms or how the timing and rate of different stages of biofilm formation occur.

## (2) Plastics as a substrate for harmful or invasive taxa

Plastics can also serve as a suitable substrate for potentially harmful microorganisms (Wang *et al.*, 2019). In a recent review, Audrézet *et al.* (2021) emphasised the association of phyto-benthic organisms with plastic debris. For example, the diatom *Ceratoneis closterium* has been consistently identified on marine plastics. This organism, which is known to be associated with mucilage events (the formation of large aggregates in the water column), can pose a risk to aquatic organisms, particularly fishes, by reducing oxygen availability (Masó *et al.*, 2016). Casabianca *et al.* (2019) reported the presence of toxin-producing cyanobacteria on floating plastics in the sea, including species belonging to the toxic bloom-forming genus *Pseudo-nitzschia*. Floating plastic debris collected from various locations along

the Catalan coast in the northwestern Mediterranean has been found to host potentially harmful dinoflagellates (*Ostreopsis* spp. and *Coelia* spp.), and both temporary cysts and vegetative cells of the harmful algal bloom species *Alexandrium taylori* (Masó *et al.*, 2003).

On a broader scale, plastic debris plays a recognized role in facilitating the spread of non-native invasive species (Audrézet *et al.*, 2021). This has been attributed to the ability of plastic fragments to be transported over significant distances, thereby promoting the dispersal of non-native and potentially invasive epiplastic organisms. Examples include the brown alga *Undaria pinnatifida* and the green alga *Codium fragile*, with the latter being particularly invasive in Europe (Audrézet *et al.*, 2021). This understudied phenomenon has the potential to pose a significant threat to biodiversity and ecosystem health (Casabianca *et al.*, 2019).

## (3) The plastisphere and its impact on biodiversity

Plastic pollution has the potential to alter the biodiversity of the recipient environment (Du *et al.*, 2022). Plastics serve as



substrates for colonisation by microorganisms (see Section-IV.1), and different taxa exhibit varying colonisation capabilities. Consequently, the plastisphere is implicated in potential changes to phytoplankton/phytobenthos community composition, influencing their biodiversity. However, current studies lack a connection between the plastisphere and descriptors of phytoplankton/phytobenthos biodiversity, such as richness, evenness, and trait- or taxonomy-based assessments. Notably, most research on the impact of plastic pollution on microorganism diversity has predominantly focused on prokaryotes (Odobel *et al.*, 2021; Du *et al.*, 2022), overlooking the broader spectrum of biodiversity, which includes eukaryotic organisms (Barros & Seena, 2021).

Several recent studies have demonstrated variations in the composition and diversity of the epiplastic community depending on different factors. For example, using 16S ribosomal RNA (rRNA) sequencing, Wen *et al.* (2020) showed that plastic colour can have impacts on the community structure and functional diversity of the plastisphere. Biofilms colonising blue plastics appeared to have a higher functional diversity than those on transparent or yellow plastics, suggesting that plastic colours/additives may drive selective pressures on microorganisms colonising plastics. Furthermore, studies have revealed differences in community composition between plastics and other non-synthetic materials, as well as differences compared with the microalgal planktonic community (Shen *et al.*, 2021). This underscores the potential of the epiplastic community to occupy a new and distinct ecological niche that favours specific organisms. For instance, Kettner *et al.* (2019) investigated the eukaryotic community composition on PE and PS, and demonstrated that the epiplastic community was significantly distinct from communities on wood and in the surrounding water, with overall lower diversity (based on richness, Shannon diversity and Simpson diversity). The authors concluded that PE and PS excluded certain organisms rather than attracting a specialised epiplastic community. In addition, no significant differences were detected between the eukaryotic communities on PE and PS, in accordance with other studies comparing microbial communities among plastic polymers (e.g. Hoellein *et al.*, 2014; Oberbeckmann, Osborn & Duhaime, 2016; Nava *et al.*, 2022). Oberbeckmann *et al.* (2014) did not find any difference in the relative diversities of communities present on different plastic types.

The effects of plastics can propagate across the planktonic community. Laboratory experiments (e.g. Hitchcock, 2022; Kettner *et al.*, 2019) have shown that phytoplankton diversity is reduced in environments with high concentrations of microplastics compared to less-polluted or control samples. This decline in diversity may be attributed to microplastics promoting the dominance of a few tolerant species, thereby reducing overall community diversity (Amaneeh *et al.*, 2023). Moreover, plastic biofilms can act as potential vectors for exogenous species that may detach from the biofilm and compete for resources within the pelagic community (Zettler *et al.*, 2013).

Alterations in phytoplankton diversity can influence secondary consumers, particularly if cyanobacteria increase in abundance while other more palatable components decrease. While there have been initial attempts to model the potential food-web implications of microplastics (e.g. Kong & Koelmans, 2019), there is a substantial need for comprehensive research to assess their impact across multiple trophic levels, in particular their effects on community structure.

#### (4) Role of the plastisphere in nutrient provisioning and cycling

While carbon is typically abundant in aquatic ecosystems, nitrogen, phosphorus, and/or iron are limiting in most ecosystems on Earth. Plastic debris provides a surface that enhances the accessibility of these limiting nutrients compared to the more diluted surrounding water system (Amaral-Zettler *et al.*, 2020; Wright *et al.*, 2020). In general, autotrophic biofilms play a pivotal role in nutrient cycling. Although experimental studies have demonstrated lower nutrient uptake rates from the water column of biofilms compared to free-living phytoplankton, biofilms appear to be more effective at nutrient retention (Vadeboncoeur & Steinman, 2002). Microorganisms within the biofilm have the capacity to derive nutrients not only from the water column but also through internal cycling and from the substrate to which they are attached (Vadeboncoeur & Steinman, 2002). Therefore, the plastisphere can provide a competitive advantage in terms of nutrient acquisition. Barros & Seena (2021) demonstrated that, in environments with high nutrient levels, microplastics efficiently adsorb these nutrients, transforming plastics into nutrient-rich surfaces for early-colonising microorganisms. This nutrient adsorption fosters microbial growth and enzymatic activities, resulting in more cohesive microbial communities. Moreover, nutrient limitation can be less severe in the plastisphere due to various mechanisms. For example, cyanobacteria with more efficient light-harvesting systems (phycobilisome antenna protein-encoding genes as opposed to Chl *a/b*-binding light-harvesting protein-encoding genes) have been observed on the surface of plastics (Bryant *et al.*, 2016). These more efficient systems require lower protein (i.e. nitrogen) investment per tetrapyrrole (e.g. in chlorophyll and phycobilin), thus providing an advantage in low-nutrient conditions (Bryant *et al.*, 2016). Another example is the increased presence of nitrogenase genes (i.e. *nifH*, *nifD*, and *nifK*) in metagenomes associated with plastic particles, indicating that nitrogen fixation might represent a strategy to overcome nitrogen limitation (Bryant *et al.*, 2016; Du *et al.*, 2022).

Autotrophic organisms can release nutrients into the surrounding environment, and these can potentially affect the local planktonic community (Priya *et al.*, 2022). Moreover, leaching of additives by microplastics can even promote microbial growth by serving as an auxiliary nutrient source (Rummel *et al.*, 2017). Plastic leachate has been reported to influence microbial communities, increasing bacterial

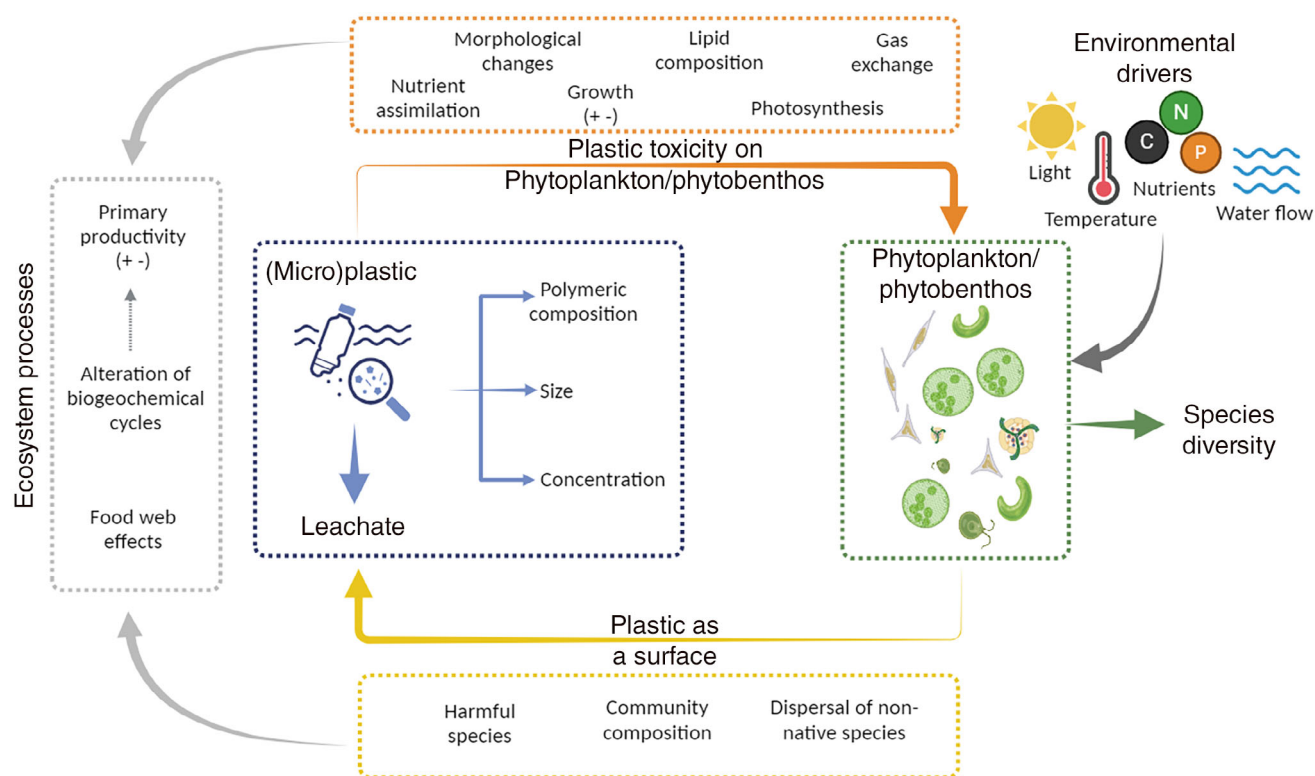
biomass at lake surfaces. Bacterial growth was found to be 1.72 times more efficient in the presence of plastic leachate compared to natural organic matter, primarily due to the greater accessibility of carbon (Sheridan *et al.*, 2022).

### (5) The impact of the plastisphere on ecosystem processes

Primary productivity is a crucial ecosystem process and supports the overall functioning of ecosystems (Larue *et al.*, 2021). Several studies have investigated the potential effects of the plastisphere on primary productivity in different ecosystems (Larue *et al.*, 2021; Miao *et al.*, 2021; Conan *et al.*, 2022). The impact can vary depending on the ecosystem, specific organisms involved, and factors such as water velocity, nutrient and light availability, and the chemistry of the substrate surface (Fig. 4) (Battin *et al.*, 2016; Bridier *et al.*, 2017; Castro-Castellon *et al.*, 2022; Chaudhary *et al.*, 2022; Vincent *et al.*, 2022). While some studies suggest that the plastisphere may enhance primary productivity and microbial diversity,

others indicate potential negative effects through changes in species diversity and altered community composition (Zettler *et al.*, 2013; Oberbeckmann, Kreikemeyer & Labrenz, 2018; Amaral-Zettler *et al.*, 2020). Plastics can provide a substrate for the attachment and growth of photosynthetic organisms, potentially enhancing local primary productivity. However, the overall effect on ecosystem-wide productivity is often negative (Amaneeh *et al.*, 2023). Plastics can also harbour species that outcompete photosynthetic organisms, leading to rapid decreases in primary productivity (Zettler *et al.*, 2013). Moreover, impacts on primary productivity are not due solely to the plastics themselves but also to the leaching of additives/plasticizers. Compounds such as phthalates and bisphenol A, for example, can reduce photosynthetic capacity (Wright *et al.*, 2020; Chaudhary *et al.*, 2022).

Microbial communities are sensitive to environmental changes (Fig. 4) and these can influence the structure and functioning of biofilms, thereby affecting primary productivity (Cross, Wallace & Rosemond, 2007; Battin *et al.*, 2016).



**Fig. 4.** Representation of the interaction between phytoplankton/phytobenthos, (micro)plastics and environmental factors (e.g. light, nutrients, temperature), and their subsequent effects on wider ecosystem processes. The effects of microplastics depend on their size, concentration, and polymeric composition, and by their leachates. These interactions may have toxic effects on phytoplankton/phytobenthos, that is morphological changes, alterations in nutrient assimilation, shifts in gas exchange, increased production of fatty acids, and impacts on growth and photosynthetic activity. The phytoplankton/phytobenthos–plastic interaction also involves effects linked to plastics serving as a surface for algal colonisation. (Micro) plastics can potentially influence community composition, provide a habitat for harmful species, and act as a vector for non-native species. Together, both toxicity and provision of a substrate for growth by (micro) plastics can impact crucial ecological processes, including primary productivity and nutrient cycling, with possible repercussions throughout the entire aquatic food web.  $\pm$  denotes instances where the effects may be positiv or negative.

The consequences of plastic pollution, therefore, are not simply due to the physical modification of habitats but may also impact the biogeochemical processes of aquatic ecosystems. Biofilms that develop on plastic substrates support robust microbial growth that affects similar ecosystem processes as natural substrates, thus affecting the base of aquatic food webs (Hoellein *et al.*, 2019; Amaral-Zettler *et al.*, 2020; Wright *et al.*, 2020).

The plastisphere can exert an influence on the metabolic traits of primary producers. Enzymes involved in nitrogen and phosphorus acquisition may be upregulated in plastisphere-associated communities in response to the altered nutrient environment around plastic debris. Consequently, this can alter the stoichiometry of nutrient cycling, potentially affecting the growth dynamics and community composition of phytoplankton populations (Fig. 4; Hutchins & Fu, 2017; Jacquin *et al.*, 2019).

The plastisphere may also affect carbon sequestration, perturbing carbon cycling, and potentially triggering cascading effects on ecosystem functioning (Arias-Andres, Rojas-Jimenez & Grossart, 2019; Amaneesh *et al.*, 2023). For example, the presence of microplastics in aquatic environments can affect the cycling of organic matter. The activity of enzymes crucial for the decomposition of organic substances can be altered by microbial consortia that colonise plastic debris by increasing the intensity of denitrification and N<sub>2</sub>O production, thereby influencing the overall rate of organic matter turnover (Su *et al.*, 2022a). For instance, biofilms on plastic can exhibit a high abundance of saprotrophic Chytridiomycota, which are known to play a key role in the decomposition of organic matter within biofilms (Oberbeckmann *et al.*, 2016; Kettner *et al.*, 2017).

To summarise, (micro)plastics result in changes to the community composition of the epiplastic community and are likely to shape the development and evolution of phytoplankton/phytobenthos. Consequently, these alterations have far-reaching implications for the dynamics of the entire ecosystem (Amaral-Zettler *et al.*, 2020).

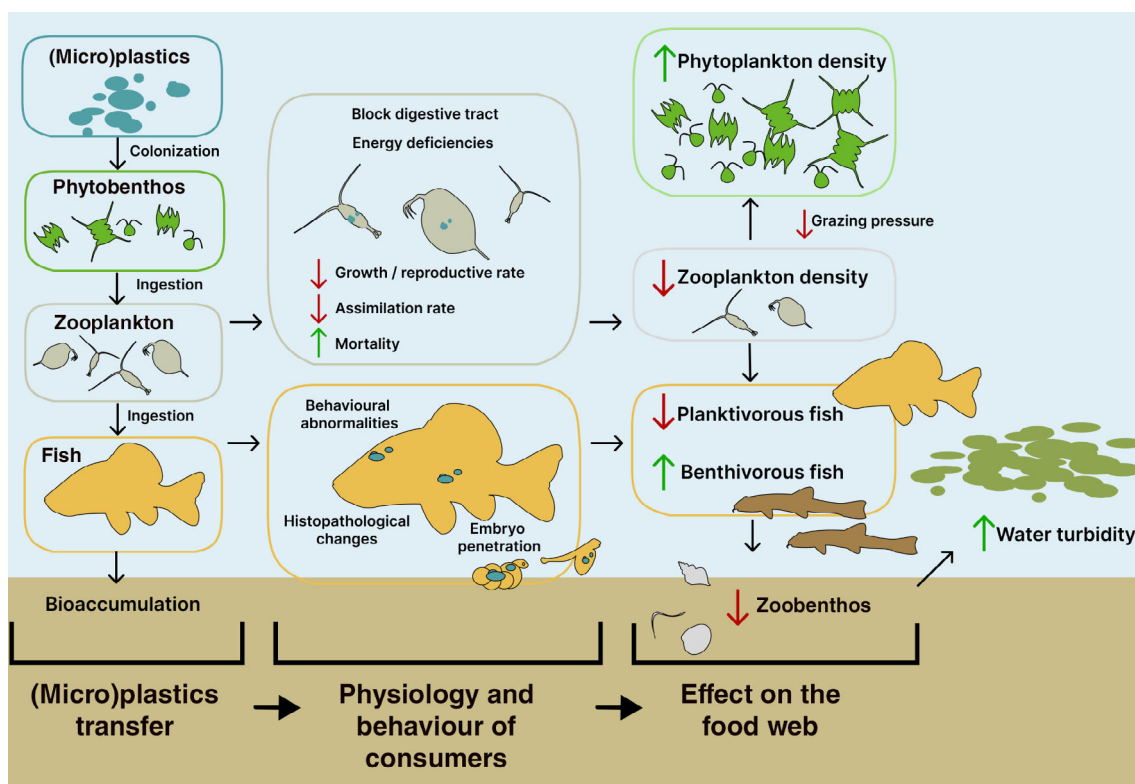
## V. EFFECTS ON THE AQUATIC FOOD WEB

Due to their small size, microplastics are readily accessible to organisms throughout the food web, as highlighted by their presence in a variety of species (Li *et al.*, 2020). Microplastics can affect food webs (Fig. 5) by alteration of (i) trophic relationships between producers and consumers; (ii) the flux of energy across different trophic levels; and (iii) the behaviour and physiology of consumers (Foley *et al.*, 2018; Gerdes *et al.*, 2019; Malinowski *et al.*, 2023).

The interaction of phytoplankton with plastics plays a crucial role in shaping food web processes. Plastic particles adsorbed by phytoplankton at the base of the trophic chain can thus reach subsequent trophic levels. Chae *et al.* (2018) observed the uptake of microplastics by fish species including *Orizias sinensis* and *Zacco temminckii*. The plastic particles

entered the fish through the consumption of the crustacean *Daphnia magna*, which fed on the green alga, *C. reinhardtii*. Ingestion of microplastics by this route led to histopathological changes in the liver of the affected fish. Of concern, microplastic ingestion extended into the next generation through penetration of embryo walls, establishing a direct link between plastic ingestion and inter-generational physiological consequences (Chae *et al.*, 2018). Similarly, Mattsson *et al.* (2017) attributed behavioural abnormalities in goldfish *Carassius carassius* to the accumulation of plastic particles in the brain. In their study, positively charged PS nanoparticles ingested by the zooplankton *D. magna* through its diet of *Scenedesmus* spp. algae were subsequently consumed by the goldfish. In a similar experiment, Cedervall *et al.* (2012) documented altered fish feeding behaviour and reduced fat reserve metabolism as a result of food chain exposure to microplastics. Biofouling of plastics (see Section IV.1) can increase ingestion rates at higher trophic levels, as plastics become more appealing to organisms like zooplankton due to the accumulation of biofilms with microalgae on their surfaces (Polhill *et al.*, 2022).

The processes underlying transfer of microplastics through the food web are complex and involve various organisms at different trophic levels. Microplastics can impact the functioning of aquatic ecosystems by affecting both bottom-up and top-down regulatory mechanisms. For instance, the consumption of hetero-aggregates formed by phytoplanktonic organisms and microplastic particles has potential negative impacts on zooplankton, including a reduction in the neutral lipid content and quality of the diet, in addition to dilution of food (i.e. replacement part of the diet with non-nutritive particles) (Bucci *et al.*, 2024). The resulting reduced assimilation rate of nutrients ultimately can lead to a decline in zooplankton population density (Casabianca *et al.*, 2020; Nava & Leoni, 2021). Microplastics also can block the digestive tracts of zooplankton (Rosenkranz *et al.*, 2009; Nasser & Lynch, 2016), reduce their feeding rate (Nasser & Lynch, 2016), or directly interfere with their feeding processes (Au *et al.*, 2015; Blarer & Burkhardt-Holm, 2016). The consequences include energy deficiency, decreased growth, reduced activity, impaired reproductive capacity, and even mortality [Besseling *et al.*, 2014; Wang *et al.* (2019) and references therein]. Thus, increased microplastic concentrations can directly lead to a decline in zooplankton populations, and consequently impact food web dynamics. Malinowski *et al.* (2023) investigated the impact of increasing microplastic concentrations on filter-feeding crustaceans (*Daphnia dentifera* and *Arctodiaptomus dorsalis*) concurrent with phytoplankton exposure. The study found that higher microplastic levels reduced zooplankton grazing pressure on phytoplankton, leading to increased algal populations. Similar results were observed in a trophic cascade experiment conducted by Pan *et al.* (2022) which found that increased microplastic load reduced the grazing activity of *D. magna* as well as its population density. The diminished density of zooplankton can subsequently restrict the density or abundance of planktivorous fishes and, ultimately, of piscivorous fishes that



**Fig. 5.** How microplastics may shape aquatic food webs. Microplastics, due to their small size, are available to organisms throughout the aquatic food web, influencing energy flow, physiology and consumer behaviour. Microplastics can form hetero-aggregates with phytoplankton, affecting zooplankton and limiting nutrient uptake. Increased microplastic concentrations can lead to increased mortality of zooplankton, reducing predation pressure on phytoplankton, and thereby leading to an increase in phytoplankton populations. Declines in the abundance of planktivorous fish can change the composition of fish communities, positively affecting benthivorous fish which might result in increased sediment disturbance. Additionally, the egestion of microplastics by zooplankton may negatively impact the zoobenthos. ↑/↓ indicate positive or negative effects of microplastics on the functioning of a population in the aquatic ecosystem.

depend on them, as modelled by Kong & Koelmans (2019). Consequently, benthivorous fishes can become more prevalent, reducing the abundance of zoobenthos and increasing sediment disturbance and water turbidity (Casabianca *et al.*, 2020; Nava & Leoni, 2021). Zooplankton can egest microplastics in the form of faecal particles and plastic residuals therein can adversely affect benthic fauna communities feeding in the sediment by causing physical damage or exerting toxic effects (Green, 2016).

In addition to direct adverse effects, both micro- and macroplastics, when ingested, can introduce a variety of substances into the food web. These substances include additives, pollutants, and potentially harmful microorganisms [Wang *et al.* (2019) and references therein]. For instance, plastic debris can act as a vector in the transport of toxic benthic algae, and therefore of the toxins they produce, into the marine food web (Leite *et al.*, 2022). Recent studies have shown that toxic chemicals, heavy metals, pathogenic organisms, and degraded plastic particles can accumulate in biofilms associated with plastics, ultimately entering higher trophic levels (Okeke *et al.*, 2022). By contrast, evidence from some studies does not support the suggestion that plastics contribute significantly to the

biomagnification of contaminants. Koelmans *et al.* (2016) found that the flux of hydrophobic organic compounds (HOCs) accumulated from natural prey typically exceeds the flux from ingested microplastics in most environments. This suggests that microplastic ingestion is unlikely to elevate significantly exposure to HOCs in aquatic ecosystems. Although microplastics could potentially accumulate and biomagnify through the food web at higher trophic levels, there is a lack of empirical multi-trophic studies (Bhatt & Chauhan, 2023). A few modelling approaches have found weak evidence of biomagnification at higher trophic levels, for example in cetaceans (Alava, 2020) and otters (O'Connor *et al.*, 2022).

## VI. CONCLUSIONS

(1) Deepening our understanding of the ecological implications linked to plastic pollution is essential to comprehend its impacts fully. This begins with understanding how plastics interact with the organisms at the base of aquatic food webs, such as phytoplankton/phytobenthos. (Micro)plastics exhibit



both toxicity for phytoplanktonic organisms and act as a substrate for the growth of phyto-benthos, thereby expanding the potential impacts of these contaminants. However, determining the direction of these impacts, whether positive or negative, remains challenging, and further research is needed.

(2) While existing studies on microplastics have predominantly focused on their toxicity to phytoplankton, often considering single species, a more comprehensive understanding of the functioning of the plastisphere and of microplastic–phytoplankton interactions requires research focused on complex natural communities. Expanding our comprehension of toxic effects beyond well-known polymers (e.g. PE, PET, PP) to other types of plastics and their additives with different chemical compositions is crucial. This research should extend beyond marine ecosystems to encompass freshwater environments, especially lotic ecosystems, and consider underrepresented biomes, particularly boreal and polar ecosystems, as well as other areas vulnerable to the impacts of plastic pollution.

(3) While we are beginning to understand the process of biofilm colonisation on different plastic surfaces, the precise role of environmental factors such as light, nutrients, hydrodynamic conditions and temperature in biofilm development remains less understood. It is unclear whether these environmental parameters modulate biofilm development on plastic as they do for biofilms on natural substrates (i.e. benthic sediment). Many studies have reported differences between biofilm communities on plastic and those in the surrounding environments, suggesting that responses observed in biofilms growing on natural substrates may not be applicable to biofilms growing on plastics.

(4) Plastic is known to be colonised by diverse phyto-benthic communities that become more complex over time. Within these communities, harmful and invasive microorganisms can thrive on plastic, posing a threat to biodiversity. Plastic substrates induce changes in the structure and functioning of photoautotrophic communities, potentially leading to local alterations in biogeochemical fluxes, particularly in the nutrient cycles. This, in turn, could impact ecosystem functioning and productivity and the carbon cycle. Given the fundamental role of biofilms in various ecosystem processes and the widespread nature of plastic pollution, we stress that more information is needed on the role of biofilms growing on plastic substrates in these processes.

(5) Microplastics, due to their small size, can enter the food web, affecting organisms through the introduction of toxins and from structural features of the plastic itself, leading to impaired body functions or behaviours. These effects have been observed across ecosystems and regions, however, there is still limited information on trophic transfer dynamics and its consequences. Only a few studies have attempted to address the bioaccumulation and biomagnification of microplastics at higher trophic levels, particularly at an empirical level. Furthermore, recorded responses to microplastic exposure at the assemblage level in natural aquatic populations at different trophic levels are still insufficient for a

comprehensive understanding of the ecological impacts of microplastics resulting from trophic interactions.

## VII. ACKNOWLEDGEMENTS

The contributing authors of this review are part of the 4th Collaborative European Freshwater Science Project for Young Researchers: "Life in plastic, it's fantastic: unravelling the microalgal community of the plastisphere across European lentic systems (PhytoPlastic)". This project is supported by a joint effort of the European Federation of Freshwater Sciences (EFFS) board, the EFFS-Federated Societies, and the European Fresh and Young Researchers (EFYR). V.N. was supported by the Department of Earth and Environmental Sciences, University of Milano-Bicocca. J.Y.D. was supported by Netaji Subhas ICAR International Fellowship, 2019. L.F. received the Excellence Stipend for Studies from the Gesellschaft für Forschungsförderung Niederösterreich. A.M.D. was supported by funding from the European Union (ERC: BLOOMTOX, 101044452). M.J.C. was supported by Captación, Incorporación y Movilidad de Capital Humano de I+D+i postdoctoral contract from Junta de Andalucía (POSTDOC-21-00044), and by programa de proyectos de investigación para la incorporación de jóvenes doctores a nuevas líneas de investigación - Universidad de Granada. The authors declare no competing interests.

## VIII. AUTHOR CONTRIBUTIONS

Conceptualization: V.N., J.Y.D., V.D.S., L.F., J.P., O.A.A., B.B., M.J.C., T.CH., M.C., F.D., A.M.D., A.F., G.G., D.H., D.R.H., V.M-V., B.M., L.M-B., V.M., F.R., B.S-P., C.M.T., J.G.; Writing – original draft preparation: V.N., J.Y.D., V.D.S., L.F., J.P., O.A.A., B.B., M.J.C., T.CH., M.C., F.D., A.M.D., A.F., G.G., D.H., D.R.H., V.M-V., B.M., L.M-B., V.M., F.R., B.S-P., C.M.T., J.G.; Writing – review and editing: V.N., J.Y.D., V.D.S., L.F., J.P., O.A.A., B.B., M.J.C., T.CH., M.C., F.D., A.M.D., A.F., G.G., D.H., D.R.H., V.M-V., B.M., L.M-B., V.M., F.R., B.S-P., C.M.T., J.G.; Supervision: V.N., J.Y.D., V.D.S., L.F., J.P., J.G.; Visualization: V.N., J.P., G.G.; Project administration: V.N., J.G.; Funding acquisition: V.V., J.G.

## IX. REFERENCES

- \*ABOMOHR, A. & HANELT, D. (2022). Recent advances in micro-/nanoplastic (MNPs) removal by microalgae and possible integrated routes of energy recovery. *Microorganisms* **10**, 2400.
- \*AFIANTI, N. F., RACHMAN, A., HATMANTI, A., YOGASWARA, D., ANGGIANI, M., FITRIYA, N. & DARMAHYATI, Y. (2022). Microbial biofilm of plastic in tropical marine environment and their potential for bioremediation of plastic waste. *Journal of Ecological Engineering* **23**, 261–275.
- \*AHMED, A. S. S., BILLAH, M. M., ALI, M. M., BHUIYAN, M. K. A., GUO, L., MOHINUZZAMAN, M., HOSSAIN, M. B., RAHMAN, M. S., ISLAM, M. S.,

- YAN, M. & CAI, W. (2023). Microplastics in aquatic environments: a comprehensive review of toxicity, removal, and remediation strategies. *Science of the Total Environment* **876**, 162414.
- ALAYA, J. J. (2020). Modeling the bioaccumulation and biomagnification potential of microplastics in a cetacean foodweb of the northeastern pacific: a prospective tool to assess the risk exposure to plastic particles. *Frontiers in Marine Science* **7**, 566101.
- ALLEN, S., ALLEN, D., PHOENIX, V. R., LE ROUX, G., DURÁNTEZ JIMÉNEZ, P., SIMONNEAU, A., BINET, S. & GALOP, D. (2019). Atmospheric transport and deposition of microplastics in a remote mountain catchment. *Nature Geoscience* **12**, 339–344.
- AMANEESEH, C., ANNA BALAN, S., SILPA, P. S., KIM, J. W., GREESHMA, K., ASWATHI MOHAN, A., ROBERT ANTONY, A., GROSSART, H.-P., KIM, H.-S. & RAMANAN, R. (2023). Gross negligence: impacts of microplastics and plastic leachates on phytoplankton community and ecosystem dynamics. *Environmental Science & Technology* **57**, 5–24.
- AMARAL-ZETTLER, L. A., ZETTLER, E. R. & MINCER, T. J. (2020). Ecology of the plastisphere. *Nature Reviews Microbiology* **18**, 139–151.
- \*AMARAL-ZETTLER, L. A., ZETTLER, E. R., SLIKAS, B., BOYD, G. D., MELVIN, D. W., MORRALL, C. E., PROSKUROWSKI, G. & MINCER, T. J. (2015). The biogeography of the plastisphere: implications for policy. *Frontiers in Ecology and the Environment* **13**, 541–546.
- ANDRADY, A. L. (2017). The plastic in microplastics: a review. *Marine Pollution Bulletin* **119**, 12–22.
- ANDRADY, A. L. (2022). Plastics in the Anthropocene. In *Plastics and the Ocean*, 1st Edition (ed. A. L. ANDRADY), pp. 1–42. John Wiley & Sons, Inc., Hoboken, NJ.
- ANDRADY, A. L. & RAJAPAKSE, N. (2016). Additives and Chemicals in Plastics. In *Hazardous Chemicals Associated with Plastics in the Marine Environment* (eds H. TAKADA and H. K. KARAPANAGIOTI), pp. 1–17. Springer International Publishing, Cham.
- \*ANSARI, F. A., RATHA, S. K., RENUKA, N., RAMANNA, L., GUPTA, S. K., RAWAT, I. & BUX, F. (2021). Effect of microplastics on growth and biochemical composition of microalga *Acutodesmus obliquus*. *Algal Research* **56**, 102296.
- ARIAS-ANDRES, M., ROJAS-JIMENEZ, K. & GROSSART, H.-P. (2019). Collateral effects of microplastic pollution on aquatic microorganisms: an ecological perspective. *TrAC Trends in Analytical Chemistry* **112**, 234–240.
- AU, S. Y., BRUCE, T. F., BRIDGES, W. C. & KLAINE, S. J. (2015). Responses of *Hyalella azteca* to acute and chronic microplastic exposures. *Environmental Toxicology and Chemistry* **34**, 2564–2572.
- \*AUDRÉZET, F., ZAIKO, A., CAHILL, P., CHAMPEAU, O., TREMBLAY, L. A., SMITH, D., WOOD, S. A., LEAR, G. & POCHON, X. (2022). Does plastic type matter? Insights into non-indigenous marine larvae recruitment under controlled conditions. *PeerJ* **10**, e14549.
- AUDRÉZET, F., ZAIKO, A., LEAR, G., WOOD, S. A., TREMBLAY, L. A. & POCHON, X. (2021). Biosecurity implications of drifting marine plastic debris: current knowledge and future research. *Marine Pollution Bulletin* **162**, 111835.
- \*BAI, Z., WANG, N. & WANG, M. (2021). Effects of microplastics on marine copepods. *Ecotoxicology and Environmental Safety* **217**, 112243.
- BAKAL, T., JANATA, J., SABOVA, L., GRABIC, R., ZLÁBEK, V. & NAJMANOVA, L. (2019). Suitability and setup of next-generation sequencing-based method for taxonomic characterization of aquatic microbial biofilm. *Folia Microbiologica* **64**, 9–17.
- \*BANDH, S. A., MALLA, F. A., QAYOOM, I., MOHI-UD-DIN, H., BUTT, A. K., ALTAFA, A., WANI, S. A., BETTS, R., TRUONG, T. H., PHAM, N. D. K., CAO, D. N. & AHMED, S. F. (2023). Importance of blue carbon in mitigating climate change and plastic/microplastic pollution and promoting circular economy. *Sustainability* **15**, 2682.
- \*BARONE, G. D., FERIZOVIĆ, D., BIUNDO, A. & LINDBLAD, P. (2020). Hints at the applicability of microalgae and cyanobacteria for the biodegradation of plastics. *Sustainability* **12**, 10449.
- BARROS, J. & SEENA, S. (2021). Plastisphere in freshwaters: an emerging concern. *Environmental Pollution* **290**, 118123.
- BATTIN, T. J., BESEMER, K., BENGTSSON, M. M., ROMANI, A. M. & PACKMANN, A. I. (2016). The ecology and biogeochemistry of stream biofilms. *Nature Reviews Microbiology* **14**, 251–263.
- \*BATTULGA, B., ATARASHI-ANDOH, M., NAKANISHI, T. & KOARASHI, J. (2022). A new approach to extracting biofilm from environmental plastics using ultrasound-assisted syringe treatment for isotopic analyses. *Science of the Total Environment* **849**, 157758.
- \*BAUDRIMONT, M., ARINI, A., GUÉGAN, C., VENEL, Z., GIGAULT, J., PEDRONO, B., PRUNIER, J., MAURICE, L., TER HALLE, A. & FEURTET-MAZEL, A. (2020). Ecotoxicity of polyethylene nanoplastics from the North Atlantic oceanic gyre on freshwater and marine organisms (microalgae and filter-feeding bivalves). *Environmental Science and Pollution Research* **27**, 3746–3755.
- \*BEIRAS, R. & TATO, T. (2019). Microplastics do not increase toxicity of a hydrophobic organic chemical to marine plankton. *Marine Pollution Bulletin* **138**, 58–62.
- \*BEIRAS, R., VERDEJO, E., CAMPOY-LÓPEZ, P. & VIDAL-LIÑÁN, L. (2021). Aquatic toxicity of chemically defined microplastics can be explained by functional additives. *Journal of Hazardous Materials* **406**, 124338.
- \*BELLINGERI, A., BERGAMI, E., GRASSI, G., FALERI, C., REDONDO-HASSELLERHARM, P., KOELMANS, A. A. & CORSI, I. (2019). Combined effects of nanoplastics and copper on the freshwater alga *Raphidocelis subcapitata*. *Aquatic Toxicology* **210**, 179–187.
- \*BEN ALI, R., BEN OUADA, S., LEBOULANGER, C., JEBALI, A., SAYADI, S. & BEN OUADA, H. (2022). Emerging contaminants and nutrients recovery by *Picozystis* sp. under continuous culture in contaminated secondary municipal wastewater effluent. *Algal Research* **66**, 102804.
- \*BERGAMI, E., PUGNALINI, S., VANNUCCINI, M. L., MANFRA, L., FALERI, C., SAVORELLI, F., DAWSON, K. A. & CORSI, I. (2017). Long-term toxicity of surface-charged polystyrene nanoplastics to marine planktonic species *Dunaliella tertiolecta* and *Artemia franciscana*. *Aquatic Toxicology* **189**, 159–169.
- BESSELING, E., WANG, B., LÜRLING, M. & KOELMANS, A. A. (2014). Nanoplastic affects growth of *S. Obliquus* and reproduction of *D. Magna*. *Environmental Science & Technology* **48**, 12336–12343.
- BHATT, V. & CHAUHAN, J. S. (2023). Microplastic in freshwater ecosystem: bioaccumulation, trophic transfer, and biomagnification. *Environmental Science and Pollution Research* **30**, 9389–9400.
- BINDA, G., KALČÍKOVÁ, G., ALLAN, I. J., HURLEY, R., RÖDLDAND, E., SPANU, D. & NIZZETTO, L. (2024). Microplastic aging processes: environmental relevance and analytical implications. *TrAC Trends in Analytical Chemistry* **172**, 117566.
- BLARER, P. & BURKHARDT-HOLM, P. (2016). Microplastics affect assimilation efficiency in the freshwater amphipod *Gammarus fossarum*. *Environmental Science and Pollution Research* **23**, 23522–23532.
- BORRELLE, S. B., RINGMA, J., LAW, K. L., MONNAHAN, C. C., LEBRETON, L., MCGIVERN, A., MURPHY, E., JAMBECK, J., LEONARD, G. H., HILLEARY, M. A., ERIKSEN, M., POSSINGHAM, H. P., DE FROND, H., GERBER, L. R., POLIDORO, B., ET AL. (2020). Predicted growth in plastic waste exceeds efforts to mitigate plastic pollution. *Science* **369**, 1515–1518.
- BRAHNEY, J., HALLERUD, M., HEIM, E., HAHNENBERGER, M. & SUKUMARAN, S. (2020). Plastic rain in protected areas of the United States. *Science* **368**, 1257–1260.
- BRIDIER, A., PIARD, J.-C., PANDIN, C., LABARTHE, S., DUBOIS-BRISSENET, F. & BRIANDET, R. (2017). Spatial organization plasticity as an adaptive driver of surface microbial communities. *Frontiers in Microbiology* **8**, 1364.
- BROWNE, M. A., CRUMP, P., NIVEN, S. J., TEUTEN, E., TONKIN, A., GALLOWAY, T. & THOMPSON, R. (2011). Accumulation of microplastic on shorelines worldwide: sources and sinks. *Environmental Science & Technology* **45**, 9175–9179.
- BRYANT, J. A., CLEMENTE, T. M., VIVIANI, D. A., FONG, A. A., THOMAS, K. A., KEMP, P., KARL, D. M., WHITE, A. E. & DELONG, E. F. (2016). Diversity and activity of communities inhabiting plastic debris in the north pacific gyre. *mSystems* **1**, e00024.
- BUCCI, K., BAYOUMI, M., STEVACK, K., WATSON-LEUNG, T. & ROCHMAN, C. M. (2024). Microplastics may induce food dilution and endocrine disrupting effects in fathead minnows (*Pimephales promelas*), and decrease offspring quality. *Environmental Pollution* **345**, 123551.
- BUFFAGNI, A. (2021). The lentic and lotic characteristics of habitats determine the distribution of benthic macroinvertebrates in Mediterranean rivers. *Freshwater Biology* **66**, 13–34.
- \*CAO, J., LIAO, Y., YANG, W., JIANG, X. & LI, M. (2022a). Enhanced microalgal toxicity due to polystyrene nanoplastics and cadmium co-exposure: from the perspective of physiological and metabolomic profiles. *Journal of Hazardous Materials* **427**, 127937.
- \*CAO, Q., SUN, W., YANG, T., ZHU, Z., JIANG, Y., HU, W., WEI, W., ZHANG, Y. & YANG, H. (2022b). The toxic effects of polystyrene microplastics on freshwater algae *Chlorella pyrenoidosa* depends on the different size of polystyrene microplastics. *Chemosphere* **308**, 136135.
- CAPOLUPO, M., SØRENSEN, L., JAYASENA, K. D. R., BOOTH, A. M. & FABBRI, E. (2020). Chemical composition and ecotoxicity of plastic and car tire rubber leachates to aquatic organisms. *Water Research* **169**, 115270.
- CAROPPO, C., AZZARO, M., DELL'ACQUA, O., AZZARO, F., MAIMONE, G., RAPPAZZO, A. C., RAFFA, F. & CARUSO, G. (2022). Microbial biofilms colonizing plastic substrates in the Ross Sea (Antarctica). *Journal of Marine Science and Engineering* **10**, 1714.
- CARPENTER, E. J. & SMITH, K. L. JR. (1972). Plastic on the Sargasso sea surface. *Science* **175**, 1240–1241.
- CASABIANCA, S., BELLINGERI, A., CAPELLACCI, S., SBRANA, A., RUSSO, T., CORSI, I. & PENNA, A. (2021). Ecological implications beyond the ecotoxicity of plastic debris on marine phytoplankton assemblage structure and functioning. *Environmental Pollution* **290**, 118101.
- CASABIANCA, S., CAPELLACCI, S., GIACOBBE, M. G., DELL'AVERSANO, C., TARTAGLIONE, L., VARRIALE, F., NARIZZANO, R., RISSO, F., MORETTO, P., DAGNINO, A., BERTOLOTTO, R., BARBONE, E., UNGARO, N. & PENNA, A.

- (2019). Plastic-associated harmful microalgal assemblages in marine environment. *Environmental Pollution* **244**, 617–626.
- CASABIANCA, S., CAPELLACCI, S., PENNA, A., CANGIOTTI, M., FATTORI, A., CORSI, I., OTTAVIANI, M. F. & CARLONI, R. (2020). Physical interactions between marine phytoplankton and PET plastics in seawater. *Chemosphere* **238**, 124560.
- CASTRO-CASTELLON, A. T., HORTON, A. A., HUGHES, J. M. R., RAMPLEY, C., JEFFERS, E. S., BUSSI, G. & WHITEHEAD, P. (2022). Ecotoxicity of microplastics to freshwater biota: considering exposure and hazard across trophic levels. *Science of the Total Environment* **816**, 151638.
- CEDERVALL, T., HANSSON, L.-A., LARD, M., FROHM, B. & LINSE, S. (2012). Food chain transport of nanoparticles affects behaviour and fat metabolism in fish. *PLoS One* **7**, e32254.
- \*CERA, A. & SCALICI, M. (2021). Freshwater wild biota exposure to microplastics: a global perspective. *Ecology and Evolution* **11**, 9904–9916.
- \*CESCHIN, S., BELLINI, A. & SCALICI, M. (2021). Aquatic plants and ecotoxicological assessment in freshwater ecosystems: a review. *Environmental Science and Pollution Research* **28**, 4975–4988.
- \*CHAE, Y., KIM, D. & AN, Y.-J. (2019). Effects of micro-sized polyethylene spheres on the marine microalga *Dunaliella salina*: focusing on the algal cell to plastic particle size ratio. *Aquatic Toxicology* **216**, 105296.
- CHAE, Y., KIM, D., KIM, S. W. & AN, Y.-J. (2018). Trophic transfer and individual impact of nano-sized polystyrene in a four-species freshwater food chain. *Scientific Reports* **8**, 284.
- CHAUDHARY, A., DUNN, S. T., KELLY, J. & HOELLEIN, T. J. (2022). Plastic microbiome development in a freshwater ecosystem. *Science of the Total Environment* **848**, 157697.
- \*CHEN, Q., LI, Y. & LI, B. (2020a). Is color a matter of concern during microplastic exposure to *Senedesmus obliquus* and *Daphnia magna*? *Journal of Hazardous Materials* **383**, 121224.
- \*CHEN, X., WANG, Y., CHEN, S., SUN, Y., TAN, Q., DING, Z., LU, Y. & YU, Y. (2021). Microplastics as carbon-nutrient sources and shaper for microbial communities in stagnant water. *Journal of Hazardous Materials* **420**, 126662.
- CHEN, Y., LING, Y., LI, X., HU, J., CAO, C. & HE, D. (2020b). Size-dependent cellular internalization and effects of polystyrene microplastics in microalgae *P. helgolandica* var. *singtaensis* and *S. quadricauda*. *Journal of Hazardous Materials* **399**, 123092.
- CHEN, Z., LI, L., HAO, L., HONG, Y. & WANG, W. (2022). Hormesis-like growth and photosynthetic physiology of marine diatom *Phaeodactylum tricornutum* Bohlin exposed to polystyrene microplastics. *Frontiers of Environmental Science & Engineering* **16**, 2.
- CHENG, J., JACQUIN, J., CONAN, P. & PUJO-PAY, M. (2021). Relative influence of plastic debris size and shape, chemical composition and phytoplankton-bacteria interactions in driving seawater plastisphere abundance, diversity and activity. *Frontiers in Microbiology* **11**, 610231.
- \*CHIA, W. Y., YING TANG, D. Y., KHOO, K. S., KAY LUP, A. N. & CHEW, K. W. (2020). Nature's fight against plastic pollution: algae for plastic biodegradation and bioplastics production. *Environmental Science and Ecotechnology* **4**, 100065.
- CLAESSENS, M., MEESTER, S. D., LANDUYT, L. V., CLERCK, K. D. & JANSSEN, C. R. (2011). Occurrence and distribution of microplastics in marine sediments along the Belgian coast. *Marine Pollution Bulletin* **62**, 2199–2204.
- COLE, M., LINDEQUE, P., HALSBAND, C. & GALLOWAY, T. S. (2011). Microplastics as contaminants in the marine environment: a review. *Marine Pollution Bulletin* **62**, 2588–2597.
- CONAN, P., PHILIP, L., ORTEGA-RETUERTA, E., ODOBEL, C., DURAN, C., PANDIN, C., GIRAUD, C., MEISTERTZHEIM, A.-L., BARBE, V., TER HALL, A., PUJO-PAY, M. & GHIGLIONE, J.-F. (2022). Evidence of coupled autotrophy and heterotrophy on plastic biofilms and its influence on surrounding seawater. *Environmental Pollution* **315**, 120463.
- \*CORNEJO-D'OTTONE, M., MOLINA, V., PAVEZ, J. & SILVA, N. (2020). Greenhouse gas cycling by the plastisphere: the sleeper issue of plastic pollution. *Chemosphere* **246**, 125709.
- CROSS, W. F., WALLACE, J. B. & ROSEMOND, A. D. (2007). Nutrient enrichment reduces constraints on material flows in a detritus-based food web. *Ecology* **88**, 2563–2575.
- \*CUNHA, C., FARIA, M., NOGUEIRA, N., FERREIRA, A. & CORDEIRO, N. (2019a). Marine vs freshwater microalgal expolymer as biosolutions to microplastics pollution. *Environmental Pollution* **249**, 372–380.
- \*CUNHA, C., LOPES, J., PAULO, J., FARIA, M., KAUFMANN, M., NOGUEIRA, N., FERREIRA, A. & CORDEIRO, N. (2020). The effect of microplastics pollution in microalgal biomass production: a biochemical study. *Water Research* **186**, 116370.
- \*CUNHA, C., PAULO, J., FARIA, M., KAUFMANN, M. & CORDEIRO, N. (2019b). Ecotoxicological and biochemical effects of environmental concentrations of the plastic-bond pollutant dibutyl phthalate on *Senedesmus* sp. *Aquatic Toxicology* **215**, 105281.
- DAJBROWSKA, A. (2021). A roadmap for a plastisphere. *Marine Pollution Bulletin* **167**, 112322.
- \*DAVARPANA, E. & GUILHERMINO, L. (2015). Single and combined effects of microplastics and copper on the population growth of the marine microalgae *Tetraselmis chuii*. *Estuarine, Coastal and Shelf Science* **167**, 269–275.
- \*DAVARPANA, E. & GUILHERMINO, L. (2019). Are gold nanoparticles and microplastics mixtures more toxic to the marine microalgae *Tetraselmis chuii* than the substances individually? *Ecotoxicology and Environmental Safety* **181**, 60–68.
- \*DAVIDOV, K., IANKEVICH-KOUNIO, E., YAKOVENKO, I., KOUCHEROV, Y., RUBIN-BLUM, M. & OREN, M. (2020). Identification of plastic-associated species in the Mediterranean Sea using DNA metabarcoding with nanopore MinION. *Scientific Reports* **10**, 17533.
- \*DE OLIVEIRA, T. T. S., ANDREU, I., MACHADO, M. C., VIMBELA, G., TRIPATHI, A. & BOSE, A. (2020). Interaction of cyanobacteria with nanometer and micron sized polystyrene particles in marine and fresh water. *Langmuir* **36**, 3963–3969.
- \*DEDMAN, C. J., CHRISTIE-OLEZA, J. A., FERNÁNDEZ-JUÁREZ, V. & ECHEVESTE, P. (2022). Cell size matters: nano- and micro-plastics preferentially drive declines of large marine phytoplankton due to co-aggregation. *Journal of Hazardous Materials* **424**, 127488.
- \*DELACUVELLERIE, A., GÉRON, A., GOBERT, S. & WATTIEZ, R. (2022). New insights into the functioning and structure of the PE and PP plastispheres from the Mediterranean Sea. *Environmental Pollution* **295**, 118678.
- \*DEMIR-YILMAZ, I., YAKOVENKO, N., ROUX, C. & GUIRAUD, P. (2022). The role of microplastics in microalgae cells aggregation. A study at the molecular scale using atomic force microscopy. *Science of the Total Environment* **832**, 155036.
- \*DI PIPPO, F., VENEZIA, C., SIGHICELLI, M., PIETRELLI, L., DI VITO, S., NUGLIO, S. & ROSSETTI, S. (2020). Microplastic-associated biofilms in lentic Italian ecosystems. *Water Research* **187**, 116429.
- \*DONG, J., LI, L., LIU, Q., YANG, M., GAO, Z., QIAN, P., GAO, K. & DENG, X. (2022). Interactive effects of polymethyl methacrylate (PMMA) microplastics and salinity variation on a marine diatom *Phaeodactylum tricornutum*. *Chemosphere* **289**, 133240.
- DU, Y., LIU, X., DONG, X. & YIN, Z. (2022). A review on marine plastisphere: biodiversity, formation, and role in degradation. *Computational and Structural Biotechnology Journal* **20**, 975–988.
- \*DUDEK, K. L., CRUZ, B. N., POLIDORO, B. & NEUER, S. (2020). Microbial colonization of microplastics in the Caribbean Sea. *Limnology and Oceanography Letters* **5**, 5–17.
- \*DUSSUD, C., HUDEC, C., GEORGE, M., FABRE, P., HIGGS, P., BRUZAUD, S., DELORT, A.-M., EYHERAGUIBEL, B., MEISTERTZHEIM, A.-L., JACQUIN, J., CHENG, J., CALLAC, N., ODOBEL, C., RABOUILLE, S. & GHIGLIONE, J.-F. (2018a). Colonization of non-biodegradable and biodegradable plastics by marine microorganisms. *Frontiers in Microbiology* **9**, 1571.
- DUSSUD, C., MEISTERTZHEIM, A. L., CONAN, P., PUJO-PAY, M., GEORGE, M., FABRE, P., COUDANE, J., HIGGS, P., ELINEAU, A., PEDROTTI, M. L., GORSKY, G. & GHIGLIONE, J. F. (2018b). Evidence of niche partitioning among bacteria living on plastics, organic particles and surrounding seawaters. *Environmental Pollution* **236**, 807–816.
- EICH, A., WEBER, M. & LOTT, C. (2021). Disintegration half-life of biodegradable plastic films on different marine beach sediments. *PeerJ* **9**, e11981.
- \*ELERSEK, T., NOTERSBERG, T., KOVAČIČ, A., HEATH, E. & FILIPIČ, M. (2021). The effects of bisphenol A, F and their mixture on algal and cyanobacterial growth: from additivity to antagonism. *Environmental Science and Pollution Research* **28**, 3445–3454.
- \*FABRA, M., WILLIAMS, L., WATTS, J. E. M., HALE, M. S., COUCEIRO, F. & PRESTON, J. (2021). The plastic Trojan horse: biofilms increase microplastic uptake in marine filter feeders impacting microbial transfer and organism health. *Science of the Total Environment* **797**, 149217.
- \*FALCAO, V. G. O., CARNEIRO, D. D. C., PEREIRA, S. A., DA SILVA, M. R. D., CANDÉ, A. A. & DA CUNHA LIMA, S. T. (2020). Analyzing the toxicity of bisphenol-A to microalgae for ecotoxicological applications. *Environmental Monitoring and Assessment* **192**, 8.
- \*FAN, Y., LIU, T., QIAN, X., DENG, L., RAO, W., ZHANG, Q., ZHENG, J. & GAO, X. (2022). Metabolic impacts of polystyrene microplastics on the freshwater microalga *Microcystis aeruginosa*. *Science of the Total Environment* **836**, 155655.
- \*FENG, L.-J., SUN, X.-D., ZHU, F.-P., FENG, Y., DUAN, J.-L., XIAO, F., LI, X.-Y., SHI, Y., WANG, Q., SUN, J.-W., LIU, X.-Y., LIU, J.-Q., ZHOU, L.-L., WANG, S.-G., DING, Z., ET AL. (2020). Nanoplastics promote microcystin synthesis and release from cyanobacterial *Microcystis aeruginosa*. *Environmental Science & Technology* **54**, 3386–3394.
- FERNÁNDEZ-JUÁREZ, V., LÓPEZ-ALFARJA, X., FRANK-COMAS, A., ECHEVESTE, P., BENNASAR-FIGUERAS, A., RAMIS-MUNAR, G., GOMILA, R. M. & AGAWIN, N. S. R. (2021). “The good, the bad and the double-sword” effects of microplastics and their organic additives in marine bacteria. *Frontiers in Microbiology* **11**, 581118.
- FOLEY, C. J., FEINER, Z. S., MALINICH, T. D. & HÖÖK, T. O. (2018). A meta-analysis of the effects of exposure to microplastics on fish and aquatic invertebrates. *Science of the Total Environment* **631–632**, 550–559.

- FRIAS, J. P. G. L. & NASH, R. (2019). Microplastics: finding a consensus on the definition. *Marine Pollution Bulletin* **138**, 145–147.
- FU, D., ZHANG, Q., FAN, Z., QI, H., WANG, Z. & PENG, L. (2019). Aged microplastics polyvinyl chloride interact with copper and cause oxidative stress towards microalgae *Chlorella vulgaris*. *Aquatic Toxicology* **216**, 105319.
- \*FULFER, V. M. & MENDEN-DEUER, S. (2021). Heterotrophic dinoflagellate growth and grazing rates reduced by microplastic ingestion. *Frontiers in Marine Science* **8**, 716349.
- \*GAMBARDELLA, C., MORGANA, S., BRAMINI, M., ROTINI, A., MANFRA, L., MIGLIORE, L., PIAZZA, V., GARAVENTA, F. & FAIMALI, M. (2018). Ecotoxicological effects of polystyrene microbeads in a battery of marine organisms belonging to different trophic levels. *Marine Environmental Research* **141**, 313–321.
- \*GAMBARDELLA, C., PIAZZA, V., ALBENTOSA, M., BEBIANNO, M. J., CARDOSO, C., FAIMALI, M., GARAVENTA, F., GARRIDO, S., GONZÁLEZ, S., PÉREZ, S., SENDRA, M. & BEIRAS, R. (2019). Microplastics do not affect standard ecotoxicological endpoints in marine unicellular organisms. *Marine Pollution Bulletin* **143**, 140–143.
- GAO, G., XIN, Z., PENG, J., GAO, K. & BEARDALL, J. (2021). Current understanding and challenges for aquatic primary producers in a world with rising micro- and nano-plastic levels. *Journal of Hazardous Materials* **406**, 124685.
- \*GAO, L., FAN, H., SU, Y. & XIE, Y. (2023). Impacts of microplastic-petroleum pollution on nutrient uptake, growth, and antioxidative activity of *Chlorella vulgaris*. *Aquatic Toxicology* **255**, 106395.
- \*GAO, Z., WANG, S., ZHANG, Y. & LIU, F. (2022). Single and combined toxicity of polystyrene nanoplastics and copper on *Platymonas helgolandica* var. *tsingtaensis*: perspectives from growth inhibition, chlorophyll content and oxidative stress. *Science of the Total Environment* **829**, 154571.
- \*GARRIDO, S., LINARES, M., CAMPILLO, J. A. & ALBENTOSA, M. (2019). Effect of microplastics on the toxicity of chlorpyrifos to the microalgae *Isochrysis galbana*, clone t-ISO. *Ecotoxicology and Environmental Safety* **173**, 103–109.
- GE, J., YANG, Q., FANG, Z., LIU, S., ZHU, Y., YAO, J., MA, Z., GONÇALVES, R. J. & GUAN, W. (2022). Microplastics impacts in seven flagellate microalgae: role of size and cell wall. *Environmental Research* **206**, 112598.
- GERDES, Z., OGONOWSKI, M., NYBOM, I., EK, C., ADOLFSSON-ERICI, M., BARTH, A. & GOROKHOVA, E. (2019). Microplastic-mediated transport of PCBs? A depuration study with *Daphnia magna*. *PLoS One* **14**, e0205378.
- \*GHAFFAR, I., JAVID, A., HUSSAIN, A. & MEHMOOD, S. (2022). Uptake of Cu<sup>2+</sup> by unicellular microalgae *Chlorella vulgaris* from synthetic wastewaters is attenuated by polystyrene microspheres. *Chemosphere* **290**, 133333.
- \*GIRI, S. & MUKHERJEE, A. (2021). Ageing with algal EPS reduces the toxic effects of polystyrene nanoplastics in freshwater microalgae *Scenedesmus obliquus*. *Journal of Environmental Chemical Engineering* **9**, 105978.
- \*GOMES, T., ALMEIDA, A. C. & GEORGANTZOPOULOU, A. (2020). Characterization of cell responses in *Rhodomonas baltica* exposed to PMMA nanoplastics. *Science of the Total Environment* **726**, 138547.
- \*GONZÁLEZ-FERNÁNDEZ, C., LE GRAND, F., BIDEAU, A., HUVET, A., PAUL-PONT, I. & SOUDANT, P. (2020). Nanoplastics exposure modulate lipid and pigment compositions in diatoms. *Environmental Pollution* **262**, 114274.
- \*GONZÁLEZ-FERNÁNDEZ, C., TOULLEC, J., LAMBERT, C., LE GOÏC, N., SEOANE, M., MORICEAU, B., HUVET, A., BERCHEL, M., VINCENT, D., COURCOT, L., SOUDANT, P. & PAUL-PONT, I. (2019). Do transparent exopolymeric particles (TEP) affect the toxicity of nanoplastics on *Chaetoceros neogracile*? *Environmental Pollution* **250**, 873–882.
- \*GONZÁLEZ-PLEITER, M., PEDROUZO-RODRÍGUEZ, A., VERDÚ, I., LEGANÉS, F., MARCO, E., ROSAL, R. & FERNÁNDEZ-PIÑAS, F. (2021). Microplastics as vectors of the antibiotics azithromycin and clarithromycin: effects towards freshwater microalgae. *Chemosphere* **268**, 128824.
- \*GOPALAKRISHNAN, K. & KASHIAN, D. R. (2022). Extracellular polymeric substances in green algae facilitate microplastic deposition. *Chemosphere* **286**, 131814.
- GREEN, D. S. (2016). Effects of microplastics on European flat oysters, *Ostrea edulis* and their associated benthic communities. *Environmental Pollution* **216**, 95–103.
- \*GREEN, D. S., BOOTS, B., O'CONNOR, N. E. & THOMPSON, R. (2017). Microplastics affect the ecological functioning of an important biogenic habitat. *Environmental Science & Technology* **51**, 68–77.
- \*GRUBIŠIĆ, M., SANTEK, M. I. & SANTEK, B. (2019). Potential of microalgae for the production of different biotechnological products. *Chemical & Biochemical Engineering Quarterly* **33**, 161–181.
- \*GUERRINI, F., LOBELLE, D., MARI, L., CASAGRANDE, R. & VAN SEBILLE, E. (2023). Modeling carbon export mediated by biofouled microplastics in the Mediterranean Sea. *Limnology and Oceanography* **68**, 1078–1090.
- \*GUNAALAN, K., ALMEDA, R., LORENZ, C., VIANELLO, A., IORDACHESCU, L., PAPACHARALAMPOS, K., ROHDE KIER, C. M., VOLLERTSEN, J. & NIELSEN, T. G. (2023). Abundance and distribution of microplastics in surface waters of the Kattegat/Skagerrak (Denmark). *Environmental Pollution* **318**, 120853.
- \*GUO, Y., MA, W., LI, J., LIU, W., QI, P., YE, Y., GUO, B., ZHANG, J. & QU, C. (2020). Effects of microplastics on growth, phenanthrene stress, and lipid accumulation in a diatom, *Phaeodactylum tricornutum*. *Environmental Pollution* **257**, 113628.
- \*GUO, Y., O'BRIEN, A. M., LINS, T. F., SHAHMOHAMADLOO, R. S., ALMIRALL, X. O., ROCHMAN, C. M. & SINTON, D. (2021). Effects of hydrogen peroxide on cyanobacterium *Microcystis aeruginosa* in the presence of nanoplastics. *ACS ES&T Water* **1**, 1596–1607.
- \*GUSCHINA, I. A., HAYES, A. J. & ORMEROD, S. J. (2020). Polystyrene microplastics decrease accumulation of essential fatty acids in common freshwater algae. *Environmental Pollution* **263**, 114425.
- \*HADIYANTO, H., HARIS, A., MUHAMMAD, F., AFIATI, N. & KHOIRONI, A. (2021a). Interaction between styrofoam and microalgae *Spirulina platensis* in brackish water system. *Toxics* **9**, 43.
- \*HADIYANTO, H., KHOIRONI, A., DIANRATRI, I., HUDA, K., SUHERMAN, S. & MUHAMMAD, F. (2022a). Biodegradation of oxidized high-density polyethylene and oxo-degradable plastic using microalgae *Dunaliella salina*. *Environmental Pollutants and Bioavailability* **34**, 469–481.
- \*HADIYANTO, H., KHOIRONI, A., DIANRATRI, I., SUHERMAN, S., MUHAMMAD, F. & VAIDYANATHAN, S. (2021b). Interactions between polyethylene and polypropylene microplastics and *Spirulina* sp. microalgae in aquatic systems. *Heliyon* **7**, e07676.
- \*HADIYANTO, H., MUSLIHUDDIN, M., KHOIRONI, A., PRATIWI, W. Z., FADLILAH, M. N., MUHAMMAD, F., AFIATI, N. & DIANRATRI, I. (2022b). The effect of salinity on the interaction between microplastic polyethylene terephthalate (PET) and microalgae *Spirulina* sp. *Environmental Science and Pollution Research* **29**, 7877–7887.
- HARRISON, J. P. (2018). Microplastic-associated biofilms: a comparison of freshwater and marine environments. In *Freshwater Microplastics*. Springer, Cham.
- HARTMANN, N. B., HÜFFER, T., THOMPSON, R. C., HASSELLÖV, M., VERSCHOOR, A., DAUGAARD, A. E., RIST, S., KARLSSON, T., BRENNHOLT, N., COLE, M., HERRLING, M. P., HESS, M. C., IVLEVA, N. P., LUSHER, A. L. & WAGNER, M. (2019). Are we speaking the same language? Recommendations for a definition and categorization framework for plastic debris. *Environmental Science & Technology* **53**, 1039–1047.
- \*HAZEEM, L. J., YESILAY, G., BOUOUDINA, M., PERNA, S., CETIN, D., SULUDERE, Z., BARRAS, A. & BOUKHERROUB, R. (2020). Investigation of the toxic effects of different polystyrene micro- and nanoplastics on microalgae *Chlorella vulgaris* by analysis of cell viability, pigment content, oxidative stress and ultrastructural changes. *Marine Pollution Bulletin* **156**, 111278.
- HITCHCOCK, J. N. (2022). Microplastics can alter phytoplankton community composition. *Science of the Total Environment* **819**, 153074.
- HOELLEIN, T., ROJAS, M., PINK, A., GASIOR, J. & KELLY, J. (2014). Anthropogenic litter in urban freshwater ecosystems: distribution and microbial interactions. *PLoS One* **9**(6), e98485.
- HOELLEIN, T. J., SHOGREN, A. J., TANK, J. L., RISTECA, P. & KELLY, J. J. (2019). Microplastic deposition velocity in streams follows patterns for naturally occurring allochthonous particles. *Scientific Reports* **9**, 3740.
- \*HOLZER, M., MITRANO, D. M., CARLES, L. & WAGNER, B. (2022). Important ecological processes are affected by the accumulation and trophic transfer of nanoplastics in a freshwater periphyton-grazer food chain. *Environmental Science. Nano* **9**, 2990–3003.
- HOPE, J. A., COCO, G., LADEWIG, S. & THRUSH, S. F. (2021). The distribution and ecological effects of microplastics in an estuarine ecosystem. *Environmental Pollution* **288**, 117731.
- \*HOPE, J. A., COCO, G. & THRUSH, S. F. (2020). Effects of polyester microfibers on microphytobenthos and sediment-dwelling infauna. *Environmental Science & Technology* **54**, 7970–7982.
- \*HOU, H., WANG, S., JI, B., ZHANG, Y., PI, K. & SHI, Y. (2022). Adaptation responses of microalgal-bacterial granular sludge to polystyrene microplastic particles in municipal wastewater. *Environmental Science and Pollution Research* **29**, 59965–59973.
- \*HOU, X., MU, L., HU, X. & GUO, S. (2023). Warming and microplastic pollution shape the carbon and nitrogen cycles of algae. *Journal of Hazardous Materials* **447**, 130775.
- HU, D., SHEN, M., ZHANG, Y., LI, H. & ZENG, G. (2019). Microplastics and nanoplastics: would they affect global biodiversity change? *Environmental Science and Pollution Research* **26**, 19997–20002.
- \*HUANG, H., LIU, P., SHI, Y., WU, X. & GAO, S. (2022). Remarkable characteristics and distinct community of biofilms on the photoaged polyethylene films in riverine microcosms. *Environmental Pollution* **292**, 118485.
- HUTCHINS, D. A. & FU, F. (2017). Microorganisms and ocean global change. *Nature Microbiology* **2**, 17058.
- JACQUIN, J., CHENG, J., ODOBEL, C., PANDIN, C., CONAN, P., PUJO-PAY, M., BARBE, V., MEISTERHEIM, A.-L. & GHIGLIONE, J.-F. (2019). Microbial ecotoxicology of marine plastic debris: a review on colonization and biodegradation by the “plastisphere”. *Frontiers in Microbiology* **10**, 865.



- JAMBECK, J. R., GEYER, R., WILCOX, C., SIEGLER, T. R., PERRYMAN, M., ANDRADY, A., NARAYAN, R. & LAW, K. L. (2015). Plastic waste inputs from land into the ocean. *Science* **347**, 768–771.
- \*JIAO, Y., ZHU, Y., CHEN, M., WAN, L., ZHAO, Y., GAO, J., LIAO, M. & TIAN, X. (2022). The humic acid-like substances released from *Microcystis aeruginosa* contribute to defending against smaller-sized microplastics. *Chemosphere* **303**, 135034.
- JIAZHU, Z., LINGWEI, K., YAN, Z., QINGMING, L., SHAOJIE, H., YAFANG, J., ZENGLING, M. & WANCHUN, G. (2022). Antagonistic and synergistic effects of warming and microplastics on microalgae: case study of the red tide species *Prorocentrum donghaiense*. *Environmental Pollution* **307**, 119515.
- \*JIN, Z., DU, L., CHENG, Q., JIANG, Y., HUI, C., XU, L., ZHAO, Y. & JIANG, H. (2022). Physiological and transcriptional responses of *Dictyosphaerium* sp. under co-exposure of a typical microplastic and nonylphenol. *Environmental Research* **204**, 112287.
- \*KERFAHI, D., HARVEY, B. P., YANG, Y. & KIM, H. (2022). Whole community and functional gene changes of biofilms on marine plastic debris in response to ocean acidification. *Microbial Ecology* **85**, 1202–1214.
- KETTNER, M. T., OBERBECKMANN, S., LABRENZ, M. & GROSSART, H.-P. (2019). The eukaryotic life on microplastics in brackish ecosystems. *Frontiers in Microbiology* **10**, 538.
- KETTNER, M. T., ROJAS-JIMENEZ, K., OBERBECKMANN, S., LABRENZ, M. & GROSSART, H.-P. (2017). Microplastics alter composition of fungal communities in aquatic ecosystems. *Environmental Microbiology* **19**, 4447–4459.
- \*KHALID, N., AQEEL, M., NOMAN, A., HASHEM, M., MOSTAFA, Y. S., ALHAITHLOUL, H. A. S. & ALGHANEM, S. M. (2021). Linking effects of microplastics to ecological impacts in marine environments. *Chemosphere* **264**, 128541.
- KHOIRONI, A., ANGGORO, S. & SUDARNO, S. (2019). Evaluation of the interaction among *Microalgae Spirulina* sp., plastics polyethylene terephthalate and polypropylene in freshwater environment. *Journal of Ecological Engineering* **20**, 161–173.
- \*KIKI, C., QIU, Y., WANG, Q., IFON, B. E., QIN, D., CHABI, K., YU, C.-P., ZHU, Y.-G. & SUN, Q. (2022). Induced aging, structural change, and adsorption behavior modifications of microplastics by microalgae. *Environment International* **166**, 107382.
- \*KIM, L., CUI, R., KWAK, J. I. & AN, Y.-J. (2022). Trophic transfer of nanoplastics through a microalgae–crustacean–small yellow croaker food chain: inhibition of digestive enzyme activity in fish. *Journal of Hazardous Materials* **440**, 129715.
- KIRSTEIN, I. V., WICHELS, A., GULLANS, E., KROHNE, G. & GERDTS, G. (2019). The plastisphere – uncovering tightly attached plastic “specific” microorganisms. *PLoS One* **14**, e0215859.
- KOELMANS, A. A., BAKIR, A., BURTON, G. A. & JANSSEN, C. R. (2016). Microplastic as a vector for chemicals in the aquatic environment: critical review and model-supported reinterpretation of empirical studies. *Environmental Science & Technology* **50**, 3315–3326.
- \*KÖGEL, T., BJØRØY, Ø., TOTO, B., BIENFAIT, A. M. & SANDEN, M. (2020). Micro- and nanoplastic toxicity on aquatic life: determining factors. *Science of the Total Environment* **709**, 136050.
- KONG, X. & KOELMANS, A. A. (2019). Modeling decreased resilience of shallow lake ecosystems toward eutrophication due to microplastic ingestion across the food web. *Environmental Science & Technology* **53**, 13822–13831.
- KOOI, M., VAN NES, E. H., SCHEFFER, M. & KOELMANS, A. A. (2017). Ups and downs in the ocean: effects of biofouling on vertical transport of microplastics. *Environmental Science & Technology* **51**, 7963–7971.
- \*KRUGLOVA, A., MUÑOZ PALAZÓN, B., GONZÁLEZ-MARTÍNEZ, A. & MIKOLA, A. (2022). The dangerous transporters: a study of microplastic-associated bacteria passing through municipal wastewater treatment. *Environmental Pollution* **314**, 120316.
- LAGARDE, F., OLIVIER, O., ZANELLA, M., DANIEL, P., HIARD, S. & CARUSO, A. (2016). Microplastic interactions with freshwater microalgae: hetero-aggregation and changes in plastic density appear strongly dependent on polymer type. *Environmental Pollution* **215**, 331–339.
- \*LANG, X., NI, J. & HE, Z. (2022). Effects of polystyrene microplastic on the growth and volatile halocarbons release of microalgae *Phaeodactylum tricornutum*. *Marine Pollution Bulletin* **174**, 113197.
- LARUE, C., SARRET, G., CASTILLO-MICHEL, H. & PRADAS DEL REAL, A. E. (2021). A critical review on the impacts of nanoplastics and microplastics on aquatic and terrestrial photosynthetic organisms. *Small* **17**, 2005834.
- LAURITANO, C., RIZZO, C., LO GIUDICE, A. & SAGGIOMO, M. (2020). Physiological and molecular responses to main environmental stressors of microalgae and bacteria in polar marine environments. *Microorganisms* **8**, 1957.
- \*LEAL FILHO, W., HUNT, J. & KOVALEVA, M. (2021). Garbage patches and their environmental implications in a plastisphere. *Journal of Marine Science and Engineering* **9**, 1289.
- \*LEAR, G., KINGSBURY, J. M., FRANCHINI, S., GAMBARINI, V., MADAY, S. D. M., WALLBANK, J. A., WEAVER, L. & PANTOS, O. (2021). Plastics and the microbiome: impacts and solutions. *Environmental Microbiomes* **16**, 2.
- LETTE, I. D. P., MENEGOTTO, A., LANA, P. D. C. & JÚNIOR, L. L. M. (2022). A new look at the potential role of marine plastic debris as a global vector of toxic benthic algae. *Science of the Total Environment* **838**, 156262.
- \*LI, J., MAO, S., YE, Y., LÜ, J., JING, F., GUO, Y., LIU, H., WANG, P., MA, W., QI, P., ZHENG, J. & QU, C. (2021a). Micro-polyethylene particles reduce the toxicity of nano zinc oxide in marine microalgae by adsorption. *Environmental Pollution* **290**, 118042.
- \*LI, K., JIA, W., XU, L., ZHANG, M. & HUANG, Y. (2023a). The plastisphere of biodegradable and conventional microplastics from residues exhibit distinct microbial structure, network and function in plastic-mulching farmland. *Journal of Hazardous Materials* **442**, 130011.
- \*LI, R.-R., WANG, B.-L., NAN, F.-R., LV, J.-P., LIU, X.-D., LIU, Q., FENG, J. & XIE, S.-L. (2023b). Effects of polystyrene nanoplastics on the physiological and biochemical characteristics of microalga *Scenedesmus quadricauda*. *Environmental Pollution* **319**, 120987.
- LI, S., WANG, P., ZHANG, C., ZHOU, X., YIN, Z., HU, T., HU, D., LIU, C. & ZHU, L. (2020). Influence of polystyrene microplastics on the growth, photosynthetic efficiency and aggregation of freshwater microalgae *Chlamydomonas reinhardtii*. *Science of the Total Environment* **714**, 136767.
- \*LI, X., QIU, H., ZHANG, P., SONG, L., ROMERO-FREIRE, A. & HE, E. (2023c). Role of heteroaggregation and internalization in the toxicity of differently sized and charged plastic nanoparticles to freshwater microalgae. *Environmental Pollution* **316**, 120517.
- \*LI, Y., LIU, X., SHINDE, S., WANG, J. & ZHANG, P. (2021b). Impacts of micro- and nanoplastics on photosynthesis activities of photoautotrophs: a mini-review. *Frontiers in Microbiology* **12**, 773226.
- \*LIAO, Y., JIANG, X., XIAO, Y. & LI, M. (2020). Exposure of microalgae *Euglena gracilis* to polystyrene microbeads and cadmium: perspective from the physiological and transcriptional responses. *Aquatic Toxicology* **228**, 105650.
- \*LIBORIUSSEN, L. & JEPPESEN, E. (2006). Structure, biomass, production and depth distribution of periphyton on artificial substratum in shallow lakes with contrasting nutrient concentrations. *Freshwater Biology* **51**, 95–109.
- \*LIBRALATO, G., GALDIERO, E., FALANGA, A., CAROTENUTO, R., DE ALTERIIS, E. & GUIDA, M. (2017). Toxicity effects of functionalized quantum dots, gold and polystyrene nanoparticles on target aquatic biological models: a review. *Molecules* **22**, 1439.
- \*LIN, W., SU, F., LIN, M., JIN, M., LI, Y., DING, K., CHEN, Q., QIAN, Q. & SUN, X. (2020). Effect of microplastics PAN polymer and/or Cu<sup>2+</sup> pollution on the growth of *Chlorella pyrenoidosa*. *Environmental Pollution* **265**, 114985.
- \*LIN, X., LUO, L., MAO, Z., WANG, H., CHU, S., WANG, H. & LUO, S. (2022). Effect of microplastics on the removal of nitrogen and phosphorus from synthetic piggery digestate by microalgae. *Polymers* **14**, 4349.
- \*LISHA, V. S., KOTHALE, R. S., SIDHARTH, S. & KANDASUBRAMANIAN, B. (2022). A critical review on employing algae as a feed for polycarbohydrate synthesis. *Carbohydrate Polymer Technologies and Applications* **4**, 100242.
- \*LIU, C., QIU, J., TANG, Z., HU, H., MENG, F. & LI, A. (2021a). Effects of polystyrene microplastics on growth and toxin production of *Alexandrium pacificum*. *Toxins* **13**, 293.
- LIU, C., QIU, J., TANG, Z., HU, H., MENG, F. & LI, A. (2021b). Effects of polystyrene microplastics on growth and toxin production of *Alexandrium pacificum*. *Toxins* **13**, 293.
- \*LIU, F.-F., WANG, S.-C., ZHU, Z.-L. & LIU, G.-Z. (2021c). Current progress on marine microplastics pollution research: a review on pollution occurrence, detection, and environmental effects. *Water* **13**, 1713.
- LIU, G., JIANG, R., YOU, J., MUIR, D. C. G. & ZENG, E. Y. (2020). Microplastic impacts on microalgae growth: effects of size and humic acid. *Environmental Science & Technology* **54**, 1782–1789.
- \*LIU, Q., TANG, X., LUYING, L., MENGCHEN, L., LI, J., KANG, C. & ZHAO, Y. (2022a). The effects of two sized polystyrene nanoplastics on the growth, physiological functions, and toxin production of *Alexandrium tamarense*. *Chemosphere* **291**, 132943.
- \*LIU, Q., WU, H., CHEN, J., GUO, B., ZHAO, X., LIN, H., LI, W., ZHAO, X., LV, S. & HUANG, C. (2022b). Adsorption mechanism of trace heavy metals on microplastics and simulating their effect on microalgae in river. *Environmental Research* **214**, 113777.
- LONG, M., PAUL-PONT, I., HÉGARET, H., MORICEAU, B., LAMBERT, C., HUVET, A. & SOUDANT, P. (2017). Interactions between polystyrene microplastics and marine phytoplankton lead to species-specific hetero-aggregation. *Environmental Pollution* **228**, 454–463.
- \*LU, Y., HUANG, R., WANG, J., WANG, L. & ZHANG, W. (2022). Effects of polyester microfibers on the growth and toxicity production of bloom-forming cyanobacterium *Microcystis aeruginosa*. *Water* **14**, 2422.
- \*LUO, H., LI, Y., ZHAO, Y., XIANG, Y., HE, D. & PAN, X. (2020). Effects of accelerated aging on characteristics, leaching, and toxicity of commercial lead chromate pigmented microplastics. *Environmental Pollution* **257**, 113475.
- \*LUO, H., XIANG, Y., HE, D., LI, Y., ZHAO, Y., WANG, S. & PAN, X. (2019). Leaching behavior of fluorescent additives from microplastics and the toxicity of leachate to *Chlorella vulgaris*. *Science of the Total Environment* **678**, 1–9.

- MALINOWSKI, C. R., SEARLE, C. L., SCHABER, J. & HÖÖK, T. O. (2023). Microplastics impact simple aquatic food web dynamics through reduced zooplankton feeding and potentially releasing algae from consumer control. *The Science of the Total Environment* **904**, 166691.
- \*MALLIK, A., XAVIER, K. A. M., NAIDU, B. C. & NAYAK, B. B. (2021). Ecotoxicological and physiological risks of microplastics on fish and their possible mitigation measures. *Science of the Total Environment* **779**, 146433.
- MAO, Y., AI, H., CHEN, Y., ZHANG, Z., ZENG, P., KANG, L., LI, W., GU, W., HE, Q. & LI, H. (2018). Phytoplankton response to polystyrene microplastics: perspective from an entire growth period. *Chemosphere* **208**, 59–68.
- \*MARSAY, K. S., AMBROSINO, A. C., KOUCHEROV, Y., DAVIDOV, K., FIGUEIREDO, N., YAKOVENKO, I., ITZAHRI, S., MARTINS, M., SOBRAL, P. & OREN, M. (2023). The geographical and seasonal effects on the composition of marine microplastic and its microbial communities: the case study of Israel and Portugal. *Frontiers in Microbiology* **14**, 1089926.
- \*MARSAY, K. S., KOUCHEROV, Y., DAVIDOV, K., IANKELEVICH-KOUNIO, E., ITZAHRI, S., SALMON-DIVON, M. & OREN, M. (2022). High-resolution screening for marine prokaryotes and eukaryotes with selective preference for polyethylene and polyethylene terephthalate surfaces. *Frontiers in Microbiology* **13**, 845144.
- MASÓ, M., FORTUÑO, J. M., DE JUAN, S. & DEMESTRE, M. (2016). Microfouling communities from pelagic and benthic marine plastic debris sampled across Mediterranean coastal waters. *Scientia Marina* **80**, 117–127.
- MASÓ, M., GARCÉS, E., PAGÈS, F. & CAMP, J. (2003). Drifting plastic debris as a potential vector for dispersing Harmful Algal Bloom (HAB) species. *Scientia Marina* **67**, 107–111.
- MATTSSON, K., JOHNSON, E. V., MALMENDAL, A., LINSE, S., HANSSON, L.-A. & CEDERVALL, T. (2017). Brain damage and behavioural disorders in fish induced by plastic nanoparticles delivered through the food chain. *Scientific Reports* **7**, 11452.
- \*MIAO, L., WANG, C., ADYEL, T. M., WU, J., LIU, Z., YOU, G., MENG, M., QU, H., HUANG, L., YU, Y. & HOU, J. (2020). Microbial carbon metabolic functions of biofilms on plastic debris influenced by the substrate types and environmental factors. *Environment International* **143**, 106007.
- MIAO, L., YU, Y., ADYEL, T. M., WANG, C., LIU, Z., LIU, S., HUANG, L., YOU, G., MENG, M., QU, H. & HOU, J. (2021). Distinct microbial metabolic activities of biofilms colonizing microplastics in three freshwater ecosystems. *Journal of Hazardous Materials* **403**, 123577.
- \*MICHLEK-KOZMA, D. N., NEU, T. R. & GABEL, F. (2022). Environmental conditions affect the food quality of plastic associated biofilms for the benthic grazer *Physa fontinalis*. *Science of the Total Environment* **816**, 151663.
- \*MILOLOZA, M., BULE, K., PREVARIĆ, V., CVETNIĆ, M., UKIĆ, S., BOLANČA, T. & KUČIĆ GRGIĆ, D. (2022). Assessment of the influence of size and concentration on the ecotoxicity of microplastics to microalgae *Scenedesmus* sp., bacterium *Pseudomonas putida* and yeast *Saccharomyces cerevisiae*. *Polymers* **14**, 1246.
- \*MILOLOZA, M., BULE, K., UKIĆ, S., CVETNIĆ, M., BOLANČA, T., KUŠIĆ, H., BULATOVIĆ, V. O. & GRGIĆ, D. K. (2021). Ecotoxicological determination of microplastic toxicity on algae *Chlorella* sp.: response surface modeling approach. *Water, Air, & Soil Pollution* **232**, 327.
- \*MIZUKAMI-MURATA, S., SUZUKI, Y., SAKURAI, K. & YAMASHITA, H. (2021). Freshwater alga *Raphidocelis subcapitata* undergoes metabolic changes in response to electrostatic adhesion by micrometer-sized nylon 6 particles. *Environmental Science and Pollution Research* **28**, 66901–66913.
- \*M'RABET, C., KÉFI-DALY YAHIA, O., CHOMÉRAT, N., ZENTZ, F., BILIEN, G. & PRINGAULT, O. (2021). Transient effect of bisphenol A (BPA) and di-(2-ethylhexyl) phthalate (DEHP) on the cosmopolitan marine diatom *Chaetoceros decipiens-lorenzianus*. *Environmental Pollution* **285**, 117362.
- \*M'RABET, C., PRINGAULT, O., ZMERLI-TRIKI, H., BEN GHARBI, H., COUET, D. & KÉFI-DALY YAHIA, O. (2018). Impact of two plastic-derived chemicals, the Bisphenol A and the di-2-ethylhexyl phthalate, exposure on the marine toxic dinoflagellate *Alexandrium pacificum*. *Marine Pollution Bulletin* **126**, 241–249.
- \*NAM, S.-H., LEE, J. & AN, Y.-J. (2022). Towards understanding the impact of plastics on freshwater and marine microalgae: a review of the mechanisms and toxicity endpoints. *Journal of Hazardous Materials* **423**, 127174.
- NASSER, F. & LYNCH, I. (2016). Secreted protein eco-corona mediates uptake and impacts of polystyrene nanoparticles on *Daphnia magna*. *Journal of Proteomics* **137**, 45–51.
- \*NATARAJAN, L., JENIFER, M. A., PEIJNENBURG, W. J. G. M. & MUKHERJEE, A. (2023). Algal extracellular polymeric substances (algal-EPS) for mitigating the combined toxic effects of polystyrene nanoplastics and nano-TiO<sub>2</sub> in *Chlorella* sp. *Nanotoxicology* **17**, 143–156.
- \*NATARAJAN, L., OMER, S., JETLY, N., JENIFER, M. A., CHANDRASEKARAN, N., SURAIKUMAR, G. K. & MUKHERJEE, A. (2020). Eco-corona formation lessens the toxic effects of polystyrene nanoplastics towards marine microalgae *Chlorella* sp. *Environmental Research* **188**, 109842.
- NAVA, V., CHANDRA, S., AHERNE, J., ALFONSO, M. B., ANTÃO-GERALDES, A. M., ATTERMEYER, K., BAO, R., BARTRONS, M., BERGER, S. A., BIERNACZYK, M., BISSEN, R., BROOKES, J. D., BROWN, D., CAÑEDO-ARGÜELLES, M., CANLE, M., ET AL. (2023). Plastic debris in lakes and reservoirs. *Nature* **619**, 317–322.
- NAVA, V. & LEONI, B. (2021). A critical review of interactions between microplastics, microalgae and aquatic ecosystem function. *Water Research* **188**, 116476.
- NAVA, V., LEONI, B., ARIENZO, M. M., HOGAN, Z. S., GANDOLFI, I., TATANGELO, V., CARLSON, E., CHEA, S., SOUM, S., KOZLOSKI, R. & CHANDRA, S. (2024). Plastic pollution affects ecosystem processes including community structure and functional traits in large rivers. *Water Research* **259**, 121849.
- NAVA, V., MATIAS, M. G., CASTILLO-ESCRIVÁ, A., MESSYASZ, B. & LEONI, B. (2022). Microalgae colonization of different microplastic polymers in experimental mesocosms across an environmental gradient. *Global Change Biology* **28**, 1402–1413.
- \*NI, Z., TAN, L., WANG, J., CHEN, Y., ZHANG, N., MENG, F. & WANG, J. (2023). Toxic effects of pristine and aged polystyrene and their leachate on marine microalgae *Skeletonema costatum*. *Science of the Total Environment* **857**, 159614.
- \*NIGAM, H., JAIN, R., MALIK, A. & SINGH, V. (2022). Effect of different polystyrene nano-plastic concentrations on *Chlorella pyrenoidosa*. *Algal Research* **67**, 102835.
- \*NIKOLOPOULOU, I., PIPERAGKAS, O., MOSCHOS, S. & KARAYANNI, H. (2023). Bacteria release from microplastics into new aquatic environments. *Diversity* **15**, 115.
- \*NIU, Z., VANDEGEHUCHTE, M. B., CATARINO, A. I. & EVERAERT, G. (2021). Environmentally relevant concentrations and sizes of microplastic do not impede marine diatom growth. *Journal of Hazardous Materials* **409**, 124460.
- NOLTE, T. M., HARTMANN, N. B., KLEIJN, J. M., GARNÆS, J., VAN DE MEENT, D., JAN HENDRIKS, A. & BAUN, A. (2017). The toxicity of plastic nanoparticles to green algae as influenced by surface modification, medium hardness and cellular adsorption. *Aquatic Toxicology* **183**, 11–20.
- OBERBECKMANN, S., KREIKEMEYER, B. & LABRENZ, M. (2018). Environmental factors support the formation of specific bacterial assemblages on microplastics. *Frontiers in Microbiology* **8**, 2709.
- \*OBERBECKMANN, S. & LABRENZ, M. (2020). Marine microbial assemblages on microplastics: diversity, adaptation, and role in degradation. *Annual Review of Marine Science* **12**, 209–232.
- OBERBECKMANN, S., LOEDER, M. G. J., GERDTS, G. & OSBORN, A. M. (2014). Spatial and seasonal variation in diversity and structure of microbial biofilms on marine plastics in Northern European waters. *FEMS Microbiology Ecology* **90**, 478–492.
- OBERBECKMANN, S., OSBORN, A. M. & DUHAIME, M. B. (2016). Microbes on a bottle: substrate, season and geography influence community composition of microbes colonizing marine plastic debris. *PLoS One* **11**, e0159289.
- O'CONNOR, J. D., LALLY, H. T., KOELMANS, A. A., MAHON, A. M., O'CONNOR, I., NASH, R., O'SULLIVAN, J. J., BRUEN, M., HEEREY, L. & MURPHY, S. (2022). Modelling the transfer and accumulation of microplastics in a riverine freshwater food web. *Environmental Advances* **8**, 100192.
- ODOBEL, C., DUSSUD, C., PHILIP, L., DERIPPE, G., LAUTERS, M., EYHERAGUIBEL, B., BURGAUD, G., TER HALLE, A., MEISTERTZHEIM, A.-L., BRUZAUD, S., BARBE, V. & GHIGLIONE, J.-F. (2021). Bacterial abundance, diversity and activity during long-term colonization of non-biodegradable and biodegradable plastics in seawater. *Frontiers in Microbiology* **12**, 734782.
- OKKE, E. S., EZEORBA, T. P. C., CHEN, Y., MAO, G., FENG, W. & WU, X. (2022). Ecotoxicological and health implications of microplastic-associated biofilms: a recent review and prospect for turning the hazards into benefits. *Environmental Science and Pollution Research* **29**, 70611–70634.
- \*PAGE, T. S., ALMEDA, R., KOSKI, M., BOURNACA, E. & NIELSEN, T. G. (2022). Toxicity of tyre wear particle leachates to marine phytoplankton. *Aquatic Toxicology* **252**, 106299.
- PAN, Y., LONG, Y., HUI, J., XIAO, W., YIN, J., LI, Y., LIU, D., TIAN, Q. & CHEN, L. (2022). Microplastics can affect the trophic cascade strength and stability of plankton ecosystems via behavior-mediated indirect interactions. *Journal of Hazardous Materials* **430**, 128415.
- PARSAI, T., DALVI, V., MARTINS, M. & FIGUEIREDO, N. (2022). Implication of microplastic toxicity on functioning of microalgae in aquatic system. *Environmental Pollution* **308**, 119626.
- \*PENG, C., WANG, J., LIU, X. & WANG, L. (2022). Differences in the plastispheres of biodegradable and non-biodegradable plastics: a mini review. *Frontiers in Microbiology* **13**, 849147.
- PENG, G., PU, Z., CHEN, F., XU, H., CAO, X., CHUN CHEN, C., WANG, J., LIAO, Y., ZHU, X. & PAN, K. (2023). Metal leaching from plastics in the marine environment: an ignored role of biofilm. *Environment International* **177**, 107988.
- \*PICCARDO, M., PROVENZA, F., ANSELMIS, S. & RENZI, M. (2022). Ecotoxicological assessment of “glitter” leachates in aquatic ecosystems: an integrated approach. *Toxics* **10**, 677.
- POLHILL, L., DE BRUIJN, R., AMARAL-ZETTLER, L., PRAETORIUS, A. & VAN WEZEL, A. (2022). *Daphnia magna*'s favorite snack: biofouled plastics. *Environmental Toxicology and Chemistry* **41**, 1977–1981.
- PRATA, J. C., DA COSTA, J. P., LOPES, I., DUARTE, A. C. & ROCHA-SANTOS, T. (2019). Effects of microplastics on microalgae populations: a critical review. *Science of the Total Environment* **665**, 400–405.

- PRATA, J. C., LAVORANTE, B. R. B. O., B.S.M. MONTENEGRO, M. D. C. & GUILHERMINO, L. (2018). Influence of microplastics on the toxicity of the pharmaceuticals procainamide and doxycycline on the marine microalgae *Tetraselmis chuii*. *Aquatic Toxicology* **197**, 143–152.
- PRIYA, A. K., JALIL, A. A., DUTTA, K., RAJENDRAN, S., VASSEGHIAN, Y., KARIMI-MALEH, H. & SOTO-MOSCOSO, M. (2022). Algal degradation of microplastic from the environment: mechanism, challenges, and future prospects. *Algal Research* **67**, 102848.
- \*RAJU, P., SANTHANAM, P., PANDIAN, S. S., DIVYA, M., ARUNKRISHNAN, A., DEVI, K. N., ANANTH, S., ROOPAVATHY, J. & PERUMAL, P. (2022). Impact of polystyrene microplastics on major marine primary (phytoplankton) and secondary producers (copepod). *Archives of Microbiology* **204**, 84.
- RANI-BORGES, B., MOSCHINI-CARLOS, V. & POMPÊO, M. (2021). Microplastics and freshwater microalgae: what do we know so far? *Aquatic Ecology* **55**, 363–377.
- \*REICHEL, S. & GOROKHOVA, E. (2020). Micro- and nanoplastic exposure effects in microalgae: a meta-analysis of standard growth inhibition tests. *Frontiers in Environmental Science* **8**, 131.
- \*REISSER, J., SHAW, J., HALLEGRAEFF, G., PROIETTI, M., BARNES, D. K. A., THUMS, M., WILCOX, C., HARDESTY, B. D. & PATTIARATCHI, C. (2014). Millimeter-sized marine plastics: a new pelagic habitat for microorganisms and invertebrates. *PLoS One* **9**, e100289.
- \*RIPKEN, C., KHALTURIN, K. & SHOGUCHI, E. (2020). Response of coral reef dinoflagellates to nanoplastics under experimental conditions suggests downregulation of cellular metabolism. *Microorganisms* **8**, 1759.
- ROCHMAN, C. M. (2020). The story of plastic pollution: from the distant ocean gyres to the global policy stage. *Oceanography* **33**, 60–70.
- ROCHMAN, C. M. & HOELLEIN, T. (2020). The global odyssey of plastic pollution. *Science* **368**, 1184–1185.
- ROSENKRANZ, P., CHAUDHRY, Q., STONE, V. & FERNANDES, T. F. (2009). A comparison of nanoparticle and fine particle uptake by *Daphnia magna*. *Environmental Toxicology and Chemistry* **28**, 2142–2149.
- \*ROWENCZYK, L., LEFLAIVE, J., CLERGEAUD, F., MINET, A., FERRIOL, J., GAUTHIER, L., GIGAULT, J., MOUCHET, F., ORY, D., PINELLI, E., ALBIGNAC, M., ROUX, C., MINGOTAUD, A. F., SILVESTRE, J., TEN-HAGE, L., ET AL. (2021). Heteroaggregates of polystyrene nanospheres and organic matter: preparation, characterization and evaluation of their toxicity to algae in environmentally relevant conditions. *Nanomaterials* **11**, 482.
- RUMMEL, C. D., JAHNKE, A., GOROKHOVA, E., KÜHNEL, D. & SCHMITT-JANSEN, M. (2017). Impacts of biofilm formation on the fate and potential effects of microplastic in the aquatic environment. *Environmental Science & Technology Letters* **4**, 258–267.
- SCHERER, C., WEBER, A., LAMBERT, S. & WAGNER, M. (2018). Interactions of microplastics with freshwater water. In *Freshwater Microplastics* (eds M. WAGNER and S. LAMBERT), pp. 153–180. Springer International Publishing, Cham.
- \*SCHIAVO, S., OLIVIERO, M., CHIAVARINI, S., DUMONTET, S. & MANZO, S. (2021). Polyethylene, polystyrene, and polypropylene leachate impact upon marine microalgae *Dunaliella tertiolecta*. *Journal of Toxicology and Environmental Health, Part A* **84**, 249–260.
- SCHNEIDER, S. C. & PETRIN, Z. (2017). Effects of flow regime on benthic algae and macroinvertebrates - a comparison between regulated and unregulated rivers. *Science of the Total Environment* **579**, 1059–1072.
- \*SENDRA, M., STAFFIERI, E., YESTE, M. P., MORENO-GARRIDO, I., GATICA, J. M., CORSI, I. & BLASCO, J. (2019). Are the primary characteristics of polystyrene nanoplastics responsible for toxicity and ad/absorption in the marine diatom *Phaeodactylum tricornutum*? *Environmental Pollution* **249**, 610–619.
- SENOUSY, H. H., KHAIRY, H. M., EL-SAYED, H. S., SALLAM, E. R., EL-SHEIKH, M. A. & EL-SHOBARY, M. E. (2023). Interactive adverse effects of low-density polyethylene microplastics on marine microalgae *Chaetoceros calcitrans*. *Chemosphere* **311**, 137182.
- SEOANE, M., GONZÁLEZ-FERNÁNDEZ, C., SOUDANT, P., HUVET, A., ESPERANZA, M., CID, Á. & PAUL-PONT, I. (2019). Polystyrene microbeads modulate the energy metabolism of the marine diatom *Chaetoceros neogracile*. *Environmental Pollution* **251**, 363–371.
- \*SHEELA, A. M., MANIMEKALAI, B. & DHINAGARAN, G. (2022). Review on the distribution of microplastics in the oceans and its impacts: need for modeling-based approach to investigate the transport and risk of microplastic pollution. *Environmental Engineering Research* **27**, 210243.
- SHEN, C., HUANG, L., XIE, G., WANG, Y., MA, Z., YAO, Y. & YANG, H. (2021). Effects of plastic debris on the biofilm bacterial communities in lake water. *Water* **13**, 1465.
- \*SHEN, M., YE, S., ZENG, G., ZHANG, Y., XING, L., TANG, W., WEN, X. & LIU, S. (2020). Can microplastics pose a threat to ocean carbon sequestration? *Marine Pollution Bulletin* **150**, 110712.
- SHERIDAN, E. A., FONVIELLE, J. A., COTTINGHAM, S., ZHANG, Y., DITTMAR, T., ALDRIDGE, D. C. & TANENTZAP, A. J. (2022). Plastic pollution fosters more microbial growth in lakes than natural organic matter. *Nature Communications* **13**, 4175.
- \*SHIU, R.-F., VAZQUEZ, C. I., CHIANG, C.-Y., CHIU, M.-H., CHEN, C.-S., NI, C.-W., GONG, G.-C., QUIGG, A., SANTSCI, P. H. & CHIN, W.-C. (2020). Nano- and microplastics trigger secretion of protein-rich extracellular polymeric substances from phytoplankton. *Science of the Total Environment* **748**, 141469.
- SJOLLEMA, S. B., REDONDO-HASSELERHARM, P., LESLIE, H. A., KRAAK, M. H. S. & VETHAAK, A. D. (2016). Do plastic particles affect microalgal photosynthesis and growth? *Aquatic Toxicology* **170**, 259–261.
- \*SNIGIROVA, A. O., UZUN, O. Y., BONDARENKO, O. S., KAPSHYNA, I. A., SYNEGUB, I. A., PORTIANKO, V. V., KUDRENKO, S. A., RYBALKO, O. A., VOROBYOVA, L. V. & VYNOGRADOV, O. K. (2022). Biofouling growth on plastic substrates: experimental studies in the Black Sea. *Biosystems Diversity* **30**, 397–405.
- SOLANO, C., ECHEVERZ, M. & LASA, I. (2014). Biofilm dispersion and quorum sensing. *Current Opinion in Microbiology* **18**, 96–104.
- SONG, C., LIU, Z., WANG, C., LI, S. & KITAMURA, Y. (2020). Different interaction performance between microplastics and microalgae: the bio-elimination potential of *Chlorella* sp. L38 and *Phaeodactylum tricornutum* MASCC-0025. *Science of the Total Environment* **723**, 138146.
- \*SONG, W., FU, C., FANG, Y., WANG, Z., LI, J., ZHANG, X., BHATT, K., LIU, L., WANG, N., LIU, F. & ZHU, S. (2023a). Single and combined toxicity assessment of primary or UV-aged microplastics and adsorbed organic pollutants on microalga *Chlorella pyrenoidosa*. *Environmental Pollution* **318**, 120925.
- \*SONG, Y., ZHANG, B., SI, M., CHEN, Z., GENG, J., LIANG, F., XI, M., LIU, X. & WANG, R. (2023b). Roles of extracellular polymeric substances on *Microcystis aeruginosa* exposed to different sizes of polystyrene microplastics. *Chemosphere* **312**, 137225.
- SOSA, A. & CHEN, F. (2022). An *in situ* study to understand community structure of estuarine microbes on the plastisphere. *Microorganisms* **10**, 1543.
- \*SRIDHARAN, S., KUMAR, M., BOLAN, N. S., SINGH, L., KUMAR, S., KUMAR, R. & YOU, S. (2021). Are microplastics destabilizing the global network of terrestrial and aquatic ecosystem services? *Environmental Research* **198**, 111243.
- SU, X., YANG, L., YANG, K., TANG, Y., WEN, T., WANG, Y., RILLIG, M. C., ROHE, L., PAN, J., LI, H. & ZHU, Y. (2022a). Estuarine plastisphere as an overlooked source of N<sub>2</sub>O production. *Nature Communications* **13**, 3884.
- \*SU, Y., CHENG, Z., HOU, Y., LIN, S., GAO, L., WANG, Z., BAO, R. & PENG, L. (2022b). Biodegradable and conventional microplastics posed similar toxicity to marine algae *Chlorella vulgaris*. *Aquatic Toxicology* **244**, 106097.
- \*SU, Y., QI, H., HOU, Y., GAO, M., LI, J., CAI, M., ZHU, X., CHEN, M., GE, C., FU, D., WANG, Z. & PENG, L. (2022c). Combined effects of microplastics and benzo[a]pyrene on the marine diatom *Chaetoceros muelleri*. *Frontiers in Marine Science* **8**, 779321.
- SUN, A. & WANG, W.-X. (2023). Human exposure to microplastics and its associated health risks. *Environment & Health* **1**, 139–149.
- \*SUN, S., HU, X., KANG, W. & YAO, M. (2023). Combined effects of microplastics and warming enhance algal carbon and nitrogen storage. *Water Research* **233**, 119815.
- \*TARAFDAR, A., LIM, J.-Y. & KWON, J.-H. (2022). UV stabilizers can foster early development of biofilms on freshwater microplastics. *Environmental Pollution* **315**, 120444.
- TETU, S. G., SARKER, I., SCHRAMMEYER, V., PICKFORD, R., ELBOURNE, L. D. H., MOORE, L. R. & PAULSEN, I. T. (2019). Plastic leachates impair growth and oxygen production in *Prochlorococcus*, the ocean's most abundant photosynthetic bacteria. *Communications Biology* **2**, 184.
- \*TIWARI, N., BANSAL, M., SANTHIYA, D. & SHARMA, J. G. (2022). Insights into microbial diversity on plastisphere by multi-omics. *Archives of Microbiology* **204**, 216.
- \*TODD, P. A., ONG, X. & CHOU, L. M. (2010). Impacts of pollution on marine life in Southeast Asia. *Biodiversity and Conservation* **19**, 1063–1082.
- \*TONG, C. Y. & DEREK, C. J. C. (2021). The role of substrates towards marine diatom *Cylindrotheca fusiformis* adhesion and biofilm development. *Journal of Applied Phycology* **33**, 2845–2862.
- \*TUNALI, M., UZOEFUNA, E. N., TUNALI, M. M. & YENIGUN, O. (2020). Effect of microplastics and microplastic-metal combinations on growth and chlorophyll a concentration of *Chlorella vulgaris*. *Science of the Total Environment* **743**, 140479.
- VADEBONCOEUR, Y., KALFF, J., CHRISTOFFERSEN, K. & JEPPESEN, E. (2006). Substratum as a driver of variation in periphyton chlorophyll and productivity in lakes. *Journal of the North American Benthological Society* **25**, 379–392.
- VADEBONCOEUR, Y. & STEINMAN, A. D. (2002). Periphyton function in lake ecosystems. *The Scientific World Journal* **2**, 1449–1468.
- \*VENÂNCIO, C., FERREIRA, I., MARTINS, M. A., SOARES, A. M. V. M., LOPES, I. & OLIVEIRA, M. (2019). The effects of nanoplastics on marine plankton: a case study with polymethylmethacrylate. *Ecotoxicology and Environmental Safety* **184**, 109632.
- \*VERDÚ, I., GONZÁLEZ-PLEITER, M., LEGANÉS, F., ROSAL, R. & FERNÁNDEZ-PIÑAS, F. (2021). Microplastics can act as vector of the biocide triclosan exerting damage to freshwater microalgae. *Chemosphere* **266**, 129193.
- \*VIDAL-VERDÚ, À., LATORRE-PÉREZ, A., MOLINA-MENOR, E., BAIXERAS, J., PERETÓ, J. & PORCAR, M. (2022). Living in a bottle: bacteria from sediment-associated Mediterranean waste and potential growth on polyethylene terephthalate. *Microbiology Open* **11**, e1259.

- VINCENT, A. E. S., CHAUDHARY, A., KELLY, J. J. & HOELLEIN, T. J. (2022). Biofilm assemblage and activity on plastic in urban streams at a continental scale: site characteristics are more important than substrate type. *Science of the Total Environment* **835**, 155398.
- \*WAN, J.-K., CHU, W.-L., KOK, Y.-Y. & LEE, C.-S. (2021). Influence of polystyrene microplastic and nanoplastic on copper toxicity in two freshwater microalgae. *Environmental Science and Pollution Research* **28**, 33649–33668.
- \*WANG, C., JIANG, L., HUANG, W., WANG, C. & HE, M. (2022a). Light availability modulates the responses of the microalgae *Desmodesmus* sp. to micron-sized polyvinyl chloride microplastics. *Aquatic Toxicology* **249**, 106234.
- WANG, K., LIN, H., WANG, S., DONG, X., SUN, L., ZHOU, Q., CHEN, Y., SU, B., PAN, Z., CHEN, B. & GAO, Y. (2022b). Species diversity and community structure of microalgae living on microplastics in Liyuan Bay, China. *Marine Pollution Bulletin* **180**, 113809.
- \*WANG, L., TONG, J., LI, Y., ZHU, J., ZHANG, W., NIU, L. & ZHANG, H. (2021a). Bacterial and fungal assemblages and functions associated with biofilms differ between diverse types of plastic debris in a freshwater system. *Environmental Research* **196**, 110371.
- \*WANG, Q., LIU, W., ZEB, A., LIAN, Y., SHI, R., LI, J. & ZHENG, Z. (2023a). Toxicity effects of polystyrene nanoplastics and arsenite on *Microcystis aeruginosa*. *Science of the Total Environment* **874**, 162496.
- \*WANG, S., GAO, Z., LIU, F., CHEN, S. & LIU, G. (2021b). Effects of polystyrene and triphenyl phosphate on growth, photosynthesis and oxidative stress of *Chaetoceros muelleri*. *Science of the Total Environment* **797**, 149180.
- \*WANG, S., LIU, G. & LIU, F. (2023b). Physiological and metabolic toxicity of polystyrene microplastics to *Dunaliella salina*. *Environmental Pollution* **316**, 120544.
- WANG, S., LIU, M., WANG, J., HUANG, J. & WANG, J. (2020a). Polystyrene nanoplastics cause growth inhibition, morphological damage and physiological disturbance in the marine microalga *Platymonas helgolandica*. *Marine Pollution Bulletin* **158**, 111403.
- \*WANG, S., SHI, Y., WANG, H., LI, Z. & ZHAO, M. (2023c). Succession of bacteria attached to microplastics after transferring from a mariculture area to a seagrass meadow. *Bulletin of Environmental Contamination and Toxicology* **110**, 69.
- WANG, S., WANG, Y., LIANG, Y., CAO, W., SUN, C., JU, P. & ZHENG, L. (2020b). The interactions between microplastic polyvinyl chloride and marine diatoms: physiological, morphological, and growth effects. *Ecotoxicology and Environmental Safety* **203**, 111000.
- \*WANG, S.-C., LIU, F.-F., HUANG, T.-Y., FAN, J.-L., GAO, Z.-Y. & LIU, G.-Z. (2021c). Effects of nanoplastics on the dinoflagellate *Amphidinium carterae* Hulbert from the perspectives of algal growth, oxidative stress and hemolysin production. *Nanomaterials* **11**, 2471.
- WANG, W., GAO, H., JIN, S., LI, R. & NA, G. (2019). The ecotoxicological effects of microplastics on aquatic food web, from primary producer to human: a review. *Ecotoxicology and Environmental Safety* **173**, 110–117.
- WANG, Y. & CHEN, X. (2023). Aggregation behavior of polyethylene microplastics in the nearshore environment: the role of particle size, environmental condition and turbulent flow. *Science of the Total Environment* **901**, 165941.
- WEN, B., LIU, J.-H., ZHANG, Y., ZHANG, H.-R., GAO, J.-Z. & CHEN, Z.-Z. (2020). Community structure and functional diversity of the plastisphere in aquaculture waters: does plastic color matter? *Science of the Total Environment* **740**, 140082.
- WOODWARD, G., PERKINS, D. M. & BROWN, L. E. (2010). Climate change and freshwater ecosystems: impacts across multiple levels of organization. *Philosophical Transactions of the Royal Society B: Biological Sciences* **365**, 2093–2106.
- \*WRIGHT, R. J., BOSCH, R., LANGILLE, M. G. I., GIBSON, M. I. & CHRISTIE-OLEZA, J. A. (2021). A multi-OMIC characterisation of biodegradation and microbial community succession within the PET plastisphere. *Microbiome* **9**, 141.
- WRIGHT, R. J., ERNI-CASSOLA, G., ZADJELOVIC, V., LATVA, M. & CHRISTIE-OLEZA, J. A. (2020). Marine plastic debris: a new surface for microbial colonization. *Environmental Science & Technology* **54**, 11657–11672.
- \*WU, D., WANG, T., WANG, J., JIANG, L., YIN, Y. & GUO, H. (2021). Size-dependent toxic effects of polystyrene microplastic exposure on *Microcystis aeruginosa* growth and microcystin production. *Science of the Total Environment* **761**, 143265.
- \*WU, Y., GUO, P., ZHANG, X., ZHANG, Y., XIE, S. & DENG, J. (2019). Effect of microplastics exposure on the photosynthesis system of freshwater algae. *Journal of Hazardous Materials* **374**, 219–227.
- \*XIA, Y., ZHANG, X., ZHANG, M., CHEN, L., TANG, X., SUN, Y. & LI, X. (2022). Plastic materials and water sources actively select and shape wastewater plastispheres over time. *Frontiers of Environmental Science & Engineering* **16**, 145.
- XIANBIAO, J., BAOHONG, C., KANG, W., CONGHUI, P., YAHUI, G. & HUI, L. (2023). A new microalgae community—epimicroplastic microalgae (EMP-MA). *Algal Research* **71**, 103059.
- XIAO, Y., JIANG, X., LIAO, Y., ZHAO, W., ZHAO, P. & LI, M. (2020). Adverse physiological and molecular level effects of polystyrene microplastics on freshwater microalgae. *Chemosphere* **255**, 126914.
- XIN, X., CHEN, B., PÉQUIN, B., SONG, P., YANG, M., SONG, X. & ZHANG, B. (2022). Binary toxicity of polystyrene nanoplastics and polybrominated diphenyl ethers to Arctic cyanobacteria under ambient and future climates. *Water Research* **226**, 119188.
- \*XIN, X., CHEN, B., YANG, M., GAO, S., WANG, H., GU, W., LI, X. & ZHANG, B. (2023). A critical review on the interaction of polymer particles and co-existing contaminants: adsorption mechanism, exposure factors, effects on plankton species. *Journal of Hazardous Materials* **445**, 130463.
- \*XU, H., LI, L., WANG, Y., QIU, K., CHEN, S., ZENG, J., LIU, R., YANG, Q. & HUANG, W. (2023). Differential physiological response of marine and freshwater microalgae to polystyrene microplastics. *Journal of Hazardous Materials* **448**, 130814.
- \*XU, X., WANG, S., GAO, F., LI, J., ZHENG, L., SUN, C., HE, C., WANG, Z. & QU, L. (2019). Marine microplastic-associated bacterial community succession in response to geography, exposure time, and plastic type in China's coastal seawaters. *Marine Pollution Bulletin* **145**, 278–286.
- YAN, Z., XU, L., ZHANG, W., YANG, G., ZHAO, Z., WANG, Y. & LI, X. (2021). Comparative toxic effects of microplastics and nanoplastics on *Chlamydomonas reinhardtii*: growth inhibition, oxidative stress, and cell morphology. *Journal of Water Process Engineering* **43**, 102291.
- \*YANG, W., GAO, P., LI, H., HUANG, J., ZHANG, Y., DING, H. & ZHANG, W. (2021a). Mechanism of the inhibition and detoxification effects of the interaction between nanoplastics and microalgae *Chlorella pyrenoidosa*. *Science of the Total Environment* **783**, 146919.
- \*YANG, W., GAO, P., MA, G., HUANG, J., WU, Y., WAN, L., DING, H. & ZHANG, W. (2021b). Transcriptome analysis of the toxic mechanism of nanoplastics on growth, photosynthesis and oxidative stress of microalga *Chlorella pyrenoidosa* during chronic exposure. *Environmental Pollution* **284**, 117413.
- \*YANG, W., GAO, P., NIE, Y., HUANG, J., WU, Y., WAN, L., DING, H. & ZHANG, W. (2021c). Comparison of the effects of continuous and accumulative exposure to nanoplastics on microalga *Chlorella pyrenoidosa* during chronic toxicity. *Science of the Total Environment* **788**, 147934.
- \*YANG, W., GAO, X., WU, Y., WAN, L., TAN, L., YUAN, S., DING, H. & ZHANG, W. (2020a). The combined toxicity influence of microplastics and nonylphenol on microalga *Chlorella pyrenoidosa*. *Ecotoxicology and Environmental Safety* **195**, 110484.
- \*YANG, Y., GUO, Y., O'BRIEN, A. M., LINS, T. F., ROCHMAN, C. M. & SINTON, D. (2020b). Biological responses to climate change and nanoplastics are altered in concert: full-factor screening reveals effects of multiple stressors on primary producers. *Environmental Science & Technology* **54**, 2401–2410.
- \*YE, S., RAO, M., XIAO, W., ZHOU, J. & LI, M. (2023). The relative size of microalgal cells and microplastics determines the toxicity of microplastics to microalgae. *Process Safety and Environmental Protection* **169**, 860–868.
- \*YE, T., YANG, A., WANG, Y., SONG, N., WANG, P. & XU, H. (2022). Changes of the physicochemical properties of extracellular polymeric substances (EPS) from *Microcystis aeruginosa* in response to microplastics. *Environmental Pollution* **315**, 120354.
- \*YOKOTA, K. & MEHLROSE, M. (2020). Lake phytoplankton assemblage altered by irregularly shaped PLA body wash microplastics but not by PS calibration beads. *Water* **12**, 2650.
- YOKOTA, K., WATERFIELD, H., HASTINGS, C., DAVIDSON, E., KWIETNIEWSKI, E. & WELLS, B. (2017). Finding the missing piece of the aquatic plastic pollution puzzle: interaction between primary producers and microplastics. *Limnology and Oceanography Letters* **2**, 91–104.
- YU, Y., MIAO, L., ADYEL, T. M., WALDSCHLÄGER, K., WU, J. & HOU, J. (2023). Aquatic plastisphere: interactions between plastics and biofilms. *Environmental Pollution* **322**, 121196.
- YUANYUAN, S., ZHIRUO, C., YIPENG, H., SHENGYOU, L., LIU, G., ZEZHENG, W., RUIQI, B. & LICHENG, P. (2022). Biodegradable and conventional microplastics posed similar toxicity to marine algae *Chlorella vulgaris*. *Aquatic Toxicology* **244**, 106097.
- ZETTLER, E. R., MINCER, T. J. & AMARAL-ZETTLER, L. A. (2013). Life in the “plastisphere”: microbial communities on plastic marine debris. *Environmental Science & Technology* **47**, 7137–7146.
- ZHANG, B., TANG, X., LIU, Q., LI, L., ZHAO, Y. & ZHAO, Y. (2022a). Different effecting mechanisms of two sized polystyrene microplastics on microalgal oxidative stress and photosynthetic responses. *Ecotoxicology and Environmental Safety* **244**, 114072.
- \*ZHANG, C., CHEN, X., WANG, J. & TAN, L. (2017). Toxic effects of microplastic on marine microalgae *Skeletonema costatum*: interactions between microplastic and algae. *Environmental Pollution* **220**, 1282–1288.
- \*ZHANG, J., KONG, L., ZHAO, Y., LIN, Q., HUANG, S., JIN, Y., MA, Z. & GUAN, W. (2022b). Antagonistic and synergistic effects of warming and microplastics on microalgae: case study of the red tide species *Prorocentrum donghaiense*. *Environmental Pollution* **307**, 119515.
- \*ZHAO, T., TAN, L., HUANG, W. & WANG, J. (2019). The interactions between micro polyvinyl chloride (mPVC) and marine dinoflagellate *Karenia mikimotoi*: the inhibition of growth, chlorophyll and photosynthetic efficiency. *Environmental Pollution* **247**, 883–889.



- \*ZHAO, T., TAN, L., ZHU, X., HUANG, W. & WANG, J. (2020). Size-dependent oxidative stress effect of nano/micro-scaled polystyrene on *Karenia mikimotoi*. *Marine Pollution Bulletin* **154**, 111074.
- ZHAO, Y., XIONG, X., WU, C., XIA, Y., LI, J. & WU, Y. (2018). Influence of light and temperature on the development and denitrification potential of periphytic biofilms. *Science of the Total Environment* **613–614**, 1430–1437.
- \*ZHENG, X., LIU, X., ZHANG, L. & WANG, Z. (2022). Toxicity mechanism of Nylon microplastics on *Microcystis aeruginosa* through three pathways: photosynthesis, oxidative stress and energy metabolism. *Journal of Hazardous Materials* **426**, 128094.
- ZHENG, X., ZHANG, W., YUAN, Y., LI, Y., LIU, X., WANG, X. & FAN, Z. (2021). Growth inhibition, toxin production and oxidative stress caused by three microplastics in *Microcystis aeruginosa*. *Ecotoxicology and Environmental Safety* **208**, 111575.
- ZHOU, J., GAO, L., LIN, Y., PAN, B. & LI, M. (2021). Micrometer scale polystyrene plastics of varying concentrations and particle sizes inhibit growth and upregulate microcystin-related gene expression in *Microcystis aeruginosa*. *Journal of Hazardous Materials* **420**, 126591.
- ZHU, H., FU, S., ZOU, H., SU, Y. & ZHANG, Y. (2021). Effects of nanoplastics on microalgae and their trophic transfer along the food chain: recent advances and perspectives. *Environmental Science: Processes & Impacts* **23**, 1873–1883.

## X. SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of the article.

**Appendix S1.** Methodology for the literature search.

**Table S1.** List of references included in the review of plastic–phytoplankton interactions.

(Received 24 January 2024; revised 23 October 2024; accepted 30 October 2024; published online 14 November 2024)