



Review

Human health risk associated with the management of phosphorus in freshwaters using lanthanum and aluminium



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HIGHLIGHTS

- Geo-engineering materials containing La and Al used to manage P in lakes.
- Potential impact of the use of these compounds on human health is of interest.
- La and Al uptake, kinetics and toxicity profile differ within the humans and organisms.
- Monitoring of La and Al is recommended to avoid acute and chronic exposure.

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ABSTRACT

The use of geo-engineering materials to manage phosphorus in lakes has increased in recent years with aluminium and lanthanum based materials being most commonly applied. Hence the potential impact of the use of these compounds on human health is receiving growing interest. This review seeks to understand, evaluate and compare potential unintended consequences on human health and ecotoxicological risks associated with the use of lanthanum- and aluminium-based materials to modify chemical and ecological conditions in water bodies. In addition to their therapeutic use for the reduction of intestinal phosphate absorption in patients with impaired renal function, the phosphate binding capacity of aluminium and lanthanum also led to the development of materials used for water treatment. Although lanthanum and aluminium share physicochemical similarities and have many common applications, their uptake and kinetics within the human body and living organisms importantly differ from each other which is reflected in a different toxicity profile. Whilst a causal role in the development of neurological pathologies, skeletal lesions, hematopoietic disorders and respiratory effects has unequivocally been demonstrated with increased exposure to aluminium, studies until now have failed to find such a clear association after exposure to lanthanum although caution is warranted. Our review indicates that lanthanum and aluminium have a distinctly different profile with respect to their potential effects on human health. Regular monitoring of both aluminium and lanthanum concentrations in lanthanum-/aluminium-treated water by the responsible authorities is recommended to avoid acute accidental or chronic low level accumulation.

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1. Introduction

Aluminium is a ubiquitous substance encountered both naturally (as the third most abundant element) and intentionally (used in water treatment, foods, pharmaceuticals, and vaccines); it is also present in ambient and occupational airborne particulates. Existing data underscore the importance of the physicochemical characteristics of aluminium in relation to its uptake, accumulation, and systemic bioavailability (Van Landeghem et al., 1997; Willhite et al., 2014). Aluminium has been shown to have the potential to be a toxicant to the central nervous, skeletal and hematopoietic systems. This is most prevalent through exposure to aluminium-contaminated dialysis and intravenous fluids and oral consumption of large amounts of aluminium-containing antacids and phosphate binders, especially in patients with impaired renal function. Although caricatural aluminium overload, reflected by blood and tissue levels being up to 1000 times higher compared to those currently observed has now disappeared, data in the literature suggest that low level exposure to aluminium via drinking water in individuals with normal renal function may be a contributing factor in the development of Alzheimer's disease and related disorders (Yokel and McNamara, 2001; Bondy, 2016). With regard to drinking water exposure, an important question is whether the aluminium is derived from natural sources for instance from ingestion of clay minerals (geophagia) or as a consequence of water treatment methods. Water treatment using

aluminium sulphate, i.e. alum, generally increases the percentage of dissolved, low molecular weight, (poly) aluminium species that are chemically reactive and possibly more readily absorbed, especially when used in lakes with low to moderate alkalinity (Cooke et al., 1993; Stevenson and Vance, 1989; Yokel and McNamara, 2001).

In comparison to aluminium, the ubiquity of lanthanum in the environment as well as the element's industrial applications and use in daily life is significantly less. As compared to aluminium there is less evidence for lanthanum toxicity through environmental and medical exposure which is mainly due to differences in gastrointestinal absorption, uptake kinetics, tissue accumulation and routes of elimination. Importantly, however, that with exception of its therapeutic use of lanthanum in uremic patients, much less experimental and epidemiological studies have been performed so far that have evaluated the potential toxicity of long-term low level environmental exposure. With the relatively recent introduction of the lanthanum-modified bentonite (LMB), commercially known as Phoslock (Douglas, 2002; Douglas et al., 2004a,b, 2008; Robb et al., 2003) for use in phosphorus management in polluted freshwaters, an assessment of the potential effects of lanthanum on human health deserves increasing attention.

The use of geo-engineering materials to control phosphorus in lakes has increased in recent years with aluminium and lanthanum based materials being most commonly applied (Copetti et al., 2016; Huser et al., 2016). Given that these materials are used to achieve

improvements in chemical and ecological conditions in water bodies it is understandable that efforts to forecast potential unintended consequences have focused on *ecotoxicological* risks associated with aluminium (Reitzel et al., 2013) and lanthanum (Spears et al., 2013). However, as these materials are frequently used to control harmful algal blooms in recreational water bodies (Lürling and van Oosterhout, 2013; Lurling and Tolman, 2010; Meis et al., 2012), direct contact with treated waters by humans is unavoidable. For example, LMB was used as a preventative measure in both Strathclyde Loch and The Serpentine during the Commonwealth Games (Glasgow, 2014), and Olympic Games (London, 2012), respectively, to reduce the risk of human health effects associated with cyanobacteria during open water swimming events. In addition, the use of geo-engineering materials in drinking water reservoirs has received attention given the potential for mass human exposure through drinking water supplies (Perkins and Underwood, 2001; Schintu et al., 2000). Assessments of the risk posed by human consumption of fish in treated waters have also been conducted (Landman and Ling, 2006; Landman et al., 2007). In recognition of these concerns, the use of LMB has been assessed in the context of human and environmental health protection and associated legislative mechanisms (NICNAS, 2014).

To aid water managers in the selection and appropriate use of geo-engineering materials it is important to comprehensively assess the potential for heightened human health risks across a wide range of water body types. To address this, we review here the occurrence, metabolism, routes of exposure, and potential toxicity associated with water bodies treated with lanthanum and aluminium, two of the most commonly used materials for phosphorus control (Lürling et al., 2016). We draw on evidence of concentrations of chemical species reported for treated water bodies and provide recommendations for use of materials in the context of dose and effect scenarios. The assessment approach used here has relevance for the comparative assessment of other materials proposed for use in water bodies.

2. General aspects

2.1. Occurrence and exposure of lanthanum

Lanthanum, a member of the element group called rare earth elements (REE) or lanthanides, is relatively common in the earth's crust. Its abundance may be as high as 18 parts per million (Redling, 2006), making it nearly as common as copper or zinc. Lanthanum is widely dispersed throughout the earth's crust, most commonly occurring in REE minerals such as monazite and bastnasite. These minerals generally contain all of the other REE, often in variable abundance. Reports on environmental pollution by lanthanum are scarce and mainly originate from particular regions in China (Zhao et al., 2013). The element, lanthanum, is increasingly used in industrial applications with some of its compounds being used in lamps, color televisions, cigarette lighters, optical fibers and hybrid engines (Behets, 2005; Das et al., 1988). According to a recent survey, the annual production of lanthanum was 12,500 tonnes worldwide (Haque et al., 2014). Plants generally do not accumulate lanthanum, although in some instances, accumulation of REE has been described for tea, cucumber, maize and pine leaves (Xu et al., 2003). Mosses and lichen generally contain the highest lanthanum concentrations (up to 100 ppm) (Behets, 2005; Das et al., 1988; Xu et al., 2012).

2.2. Occurrence and exposure of aluminium

Aluminium (Al) is the most abundant metallic element within the lithosphere, occurring at about 8% by weight (so over 4000

times more enriched relative to lanthanum) and the third most abundant element preceding iron (4.7%) but less abundant than oxygen and silicon. Aluminium exists primarily associated with silicates and oxides in minerals of low solubility, explaining the low (generally <1 mg/L) dissolved aluminium concentrations detected in rivers, lakes and sea water (Gensemer and Playle, 1999). Nevertheless, reports in the early 1980s pointed towards acidification of lakes as a result of acid rain thereby enhancing aluminium solubility below pH 6 and hence toxicity for fish and other biota including birds living in the immediate surroundings (Van Landeghem, 1997).

Today the daily ingested dose via drinking water is estimated to be around 160 µg aluminium/day (Willhite et al., 2014). Thanks to governmental efforts to reduce the release of sulphur dioxide (SO₂) into the atmosphere emissions in the US and Europe dropped 40% and 70% respectively since the 1990s whilst according to the Pacific Research Institute, acid rain levels have dropped 65% since 1976 (http://en.wikipedia.org/wiki/Acid_rain). Human activities have also changed exposure of living organisms to aluminium in other ways since it is widely used in transportation, packaging, construction, water treatment, a wide range of household items (Frumkin et al., 2008). In 2014 the global annual aluminium production had reached 54 million tons. It has been reported that exposure to aluminium may also occur through aluminium leaching from ceramic products (Bolte et al., 2011), migration from glass bottles (Fekete et al., 2012), smoking (Exley et al., 2006), in some antacids, and prescription phosphate binders. Hence, it is not surprising that depending on location, weather conditions, and type and level of industrial activity in the area, daily exposures may range from 0.005 to 0.18 µg/m³ in clean ambient air to 0.4–8.0 µg/m³ in urban and industrial areas. Hence exposure rates may be as little as 0.03 µg/kg/day (assuming a 70 kg body weight) in clean air to 233 µg/kg/day in polluted air to as high as 3500–5200 mg aluminium/day (i.e. 50 mg/kg/day–75 mg/kg/day) as a result of aluminium-based antacid consumption (D'Haese, 1988; Krewski et al., 2007; <https://www.atsdr.cdc.gov/phs/phs.asp?id=1076&tid=34>). To appreciate health consequences of aluminium exposure it is important to consider that the speciation is more influential to health outcomes than exposure *per se* (Willhite et al., 2014), which is also applicable to other elements, including lanthanum.

2.3. Aqueous chemistry of lanthanum

Lanthanum (La), electron configuration (Xe) 5d¹ 6s², atomic mass 138.91, is the most electropositive (cationic) element of the REE, is uniformly trivalent and its binding is generally ionic. It is a so-called hard electron acceptor with a strong preference for oxygen-containing anions. Therefore, the most common biological ligands with which it can form strong complexes are carboxyl and phosphate groups. Carbonates, phosphates and oxalates formed with lanthanum are essentially insoluble, while the chloride and sulphate complexes are soluble (Cetiner et al., 2005; Cetiner and Xiong, 2008). In aqueous solutions without any other oxyanions present, chemical modelling indicates that the majority of lanthanum occurs as a free La³⁺ cation until pH8 (Fig. 1). Above this pH a series of La–OH complexes coexist with the insoluble La(OH)₃ complex predominating between approximately pH9 and pH12. In lake restoration, lanthanum is used to intercept phosphate released from sediments and to reduce water column phosphate. Lanthanum and phosphate bind to rhabdophane (LaPO₄), a mineral with an extreme low solubility (K_{sp} 10^{-24.7} to 10^{-25.7} mol² l⁻²) (Johannesson and Lyons, 1994; Liu and Byrne, 1997). The lanthanum-phosphate bond is only affected by conditions where pH is <4 or >12. The phosphate binding capacity of lanthanum is not affected by altered redox conditions such as those in anoxic

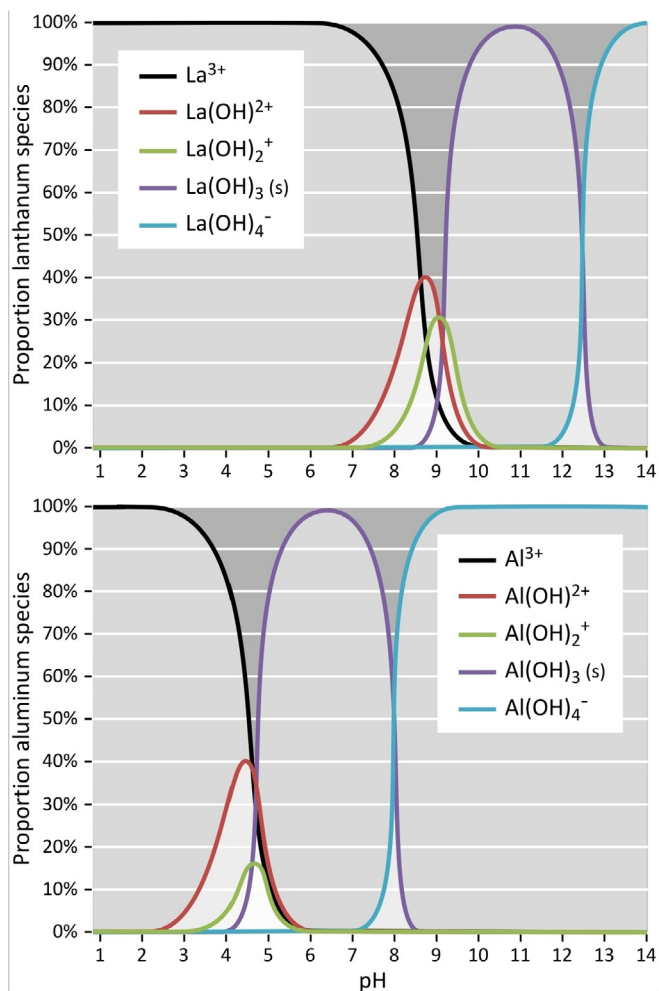


Fig. 1. Effect of pH on the aqueous chemistry of lanthanum and aluminium. The speciation of lanthanum and aluminium was evaluated by chemical equilibrium modelling using the program CHEAQS Pro (release P2013.1; Verweij, 2013) in the pH range 1–14 with 1 μM La or Al and 1 mM NaCl. The five most prominent species are presented.

waters (Ross et al., 2008). The formation of rhabdophane is not only predicted by chemical equilibrium modelling, but also has been found in sediments of 10 treated lakes across Europe (Dithmer et al., 2016). Given that rhabdophane is extremely stable and will not “separate”, the equilibrium is de facto a precipitation reaction. Consequently, any phosphate bound to lanthanum can be viewed as permanently removed from the biogeochemical cycling. An example of this is the persistence of LaPO_4 minerals in the weathering cycle that may span millions of years.

2.4. Aqueous chemistry of aluminium

Aluminium (Al), electron configuration (Ne) $3s^2 3p^1$, molecular mass 26.98, exists exclusively in the trivalent oxidation state. It is amphoteric, combining with both acids and bases to form, respectively, aluminium salts and aluminates. As it combines a relatively small ionic radius (0.54 \AA) with a high charge the free Al^{3+} concentration in aqueous solutions is very low due to the formation of aluminium hydroxide complexes. The chemical nature of aluminium in water is essentially the chemistry of $\text{Al}(\text{OH})_3$ which has an amphoteric character and a tendency to form complex ions and to polymerize. Evidence has been provided by chemical

modelling that in solutions with a pH below 5, aluminium exists predominantly as $\text{Al}(\text{H}_2\text{O})_6^{3+}$, with rising pH an insoluble $\text{Al}(\text{OH})_3$ complex forms at circumneutral pH, which re-dissolves at higher pH as the $\text{Al}(\text{OH})_4^-$ (aluminate) complex (Fig. 1) (Anderson and Berkowitz, 2010; Gensemer and Playle, 1999; Reitzel et al., 2013). Importantly, the speciation of aluminium is remarkably similar to that of lanthanum, albeit with the majority of similar Al–OH species offset by 4–5 pH units lower (Fig. 1). In lake restoration, aluminium is used as a coagulant to settle particulate matter, to complex with water column and sediment released phosphate (Cooke et al., 1993). The chemistry of aluminium is complex, as hydrolysis of aluminium is pH and temperature dependent, and amorphous aluminium hydroxide, bayerite, gibbsite with attached phosphates may form. These forms may either loose some of the attached phosphates or loose binding capacity because (i) $\text{Al}(\text{OH})_3$ begins to crystallize after forming (Berkowitz et al., 2006; de Vicente et al., 2008), (ii) are stable over a smaller pH range compared to LaPO_4 , but are just like lanthanum in that they are redox insensitive. Under high phosphate conditions minerals such as variscite ($\text{AlPO}_4 \cdot 2\text{H}_2\text{O}$) and kingite ($\text{Al}_3(\text{PO}_4)_2(\text{OH})_3 \cdot 9\text{H}_2\text{O}$) may be formed.

3. Metabolism

3.1. Inhalation of lanthanum

Lanthanum in combination with other REE may accumulate in the lungs after inhalation, mainly in occupational settings. Taking into account the limited pathological potential of REE for pulmonary lesions in combination with modern occupational exposure practices which efficiently restrict the respiratory intake of particles at work sites, health impairment is not readily expected (Redling, 2006; Richter, 2003). Electron microscopic evidence for cerium and lanthanum particles in the lung was provided in a single patient with an occupational history of REE exposure and affected by dendriform pulmonary ossification and pneumoconiosis (Yoon et al., 2005).

3.2. Inhalation of aluminium

Aluminium workers can encounter a mixture of aluminium fumes and inhalable (aerodynamic diameter $<100 \mu\text{m}$), thoracic ($<28 \mu\text{m}$), and respirable ($<10 \mu\text{m}$) aluminium particles in the occupational environment (Willhite et al., 2014). A fraction of the aluminium present in dust remains indefinitely in the lungs after inhalation, thus without entering systemic blood circulation. Hence, unlike other tissue stores of aluminium, concentrations in the lung increase with age (Han et al., 2004).

3.3. Ingestion and gastrointestinal absorption of lanthanum

Lanthanum, as all lanthanide elements, forms soluble chlorides and nitrates, but their phosphates and carbonates are generally insoluble and therefore have a low potential for systemic absorption. Studies have shown that oral doses of lanthanum are only minimally absorbed from the gut. When given as lanthanum carbonate to rats, the oral bioavailability was 0.0007% (Damment and Pennick, 2007) whilst in humans the average systemic bioavailability across different studies was $0.00089 \pm 0.00084\%$ ($n = 25$) with the highest bioavailability in any subject being 0.00294%. No clear differences in bioavailability were seen between healthy volunteers and dialysis patients, i.e. the target population for phosphate binding treatment with lanthanum carbonate (Damment and Pennick, 2008; Pennick et al., 2006).

3.4. Ingestion and gastrointestinal absorption of aluminium

Daily intake of aluminium from food is considered small. Recent studies suggest that it is in the range of 2–5 mg/day (Crisponi et al., 2013). The gastrointestinal absorption of aluminium is profoundly affected by speciation (e.g. aluminium citrate versus aluminium hydroxide) and reported fractional gastrointestinal absorption varies between 0.001% and 27% (Drüeke, 2002). The main reasons for this lack of agreement are related to analytical detection, contamination and differences in experimental protocols. With the introduction of accelerator mass spectrometry (AMS) and the possibility of using the ^{26}Al radioisotope more reliable data were generated, which however, still varied between 0.04% and 1.0% (Jouhannau et al., 1993). A fractional gastrointestinal absorption of $\pm 0.2\%$ is now generally accepted (Shirley and Lote, 2005). It is believed that intestinal absorption of aluminium includes both (i) paracellular pathways along enterocytes and through tight junctions by passive processes and, (ii) transcellular pathways through enterocytes involving both active and passive processes. Other factors that have been reported to alter intestinal aluminium absorption include calcium, iron status, parathyroid hormone, vitamin D, and the uremic state (Van Landeghem et al., 1997).

3.5. Lanthanum in blood/plasma

Background concentrations in chronic renal failure patients not treated with lanthanum carbonate revealed concentrations of $< 0.05\text{--}0.90\ \mu\text{g/L}$ in plasma whilst in subjects with normal renal function values consistently are below $0.05\ \mu\text{g/L}$ (Pennick et al., 2006). *In vitro* binding studies demonstrated that lanthanum is extensively bound ($> 99.7\%$) to plasma proteins (Damment and Pennick, 2007). In dialysis patients ($N=93$) treated with lanthanum carbonate on a daily basis over six years, plasma lanthanum concentrations $> 2.0\ \mu\text{g/L}$ were recorded in only 15 out of a total of 574 analyses with no evidence of safety concerns or increased frequency of adverse events (Hutchison et al., 2008).

3.6. Aluminium in serum/plasma

Serum aluminium concentrations in healthy non-exposed subjects as measured using appropriate contamination free techniques are below $2.0\ \mu\text{g/L}$ and should not exceed $10\ \mu\text{g/L}$ (Guidelines for aluminum toxicity are mainly based on the toxic effects seen in patients with impaired renal function. In these patients a serum aluminum level (D'Haese and De Broe, 2007):

$< 30\ \mu\text{g/L}$: aluminum-related bone disease is unlikely but possible particularly when patients are iron-overloaded.

$30\text{--}60\ \mu\text{g/L}$: aluminum-related bone disease is quite possible, especially if serum parathyroid hormone (PTH) levels are low or low-normal.

$> 60\ \mu\text{g/L}$: aluminum-related bone disease is probable, but not invariably present, especially if serum PTH levels are high, iron-transferrin saturation is low.

$> 100\ \mu\text{g/L}$: aluminum-related bone disease is most probable unless patients are iron deficient. Neurologic disorders should be checked for by taking the patients' electroencephalogram.

Whilst in the past values up to $> 500\ \mu\text{g/L}$ were seen in uremic patients treated with aluminium-containing phosphate binders either in combination with aluminium-contaminated dialysis fluids or not, nowadays with the introduction of aluminium-free medication and high performance water treatment systems concentrations above $10\ \mu\text{g/L}$ are rarely seen. The majority (80–90%) of aluminium in serum is bound to transferrin which can accommodate two aluminium ions, the first at the C-lobe and the second at the N-lobe (Mujika et al., 2012), and is considered the most

important, if not the sole, carrier protein of the element in plasma (Van Landeghem et al., 1994, 1998) with the remainder fraction bound to low molecular mass compounds. Hence, it is not surprising that the protein bound fraction is influenced by the iron status; i.e. iron-transferrin saturation, as both elements compete for binding to transferrin (Van Landeghem et al., 1997). This implies that when iron-transferrin saturation is high there is less binding of aluminium to transferrin, hence, more non-protein bound aluminium, i.e. free aluminium in circulation which thus becomes available (i) for being deposited in the calcified bone compartment thereby impairing bone mineralization, (ii) to pass the blood brain barrier by which it may induce deleterious neurological effects. On the other hand when iron transferring saturation is low, more aluminium will bind to transferrin which however may be taken up by the parathyroid gland through transferrin-mediated endocytosis which in turn may lead to a decreased PTH secretion/synthesis, hence hypoparathyroidism ensuing in the so-called adynamic or low turn-over bone disease (D'Haese, 1988; Smans et al., 2000; Van Landeghem et al., 1997 & 1998a).

Once bound to transferrin, studies from Mujika et al. (2012) revealed that conformational changes under conditions where Tyr188 is protonated permit aluminium release from the protein.

3.7. Tissue distribution of lanthanum

Data on the tissue distribution in humans, with the exception of lanthanum concentrations measured in the framework of clinical studies evaluating the therapeutic use of lanthanum carbonate, are scarce. Studies in animals with normal renal function and chronic renal failure that were environmentally exposed revealed that lanthanum concentrations in various tissues did not exceed $0.08\ \mu\text{g/g}$ wet weight and this was not dependent on renal function. In rats treated with lanthanum carbonate at an oral dose of $2000\ \text{mg/kg/day}$ over a 12 week exposure, a substantial increase was seen, particularly in bone, liver and kidney with liver lanthanum concentrations in uremic rats being 2- to 3-fold higher than those seen in non-uremic rats (Slatopolsky et al., 2005). This may be ascribed to a disruption of the intercellular junctions in the intestinal epithelium inherent to chronic renal failure. A carefully controlled time-course rat study, however, showed that steady-state concentrations of about $3\ \mu\text{g/g}$ wet weight were achieved within 6–12 weeks of treatment, indicating hepatic lanthanum uptake and elimination to be in equilibrium (Bervoets et al., 2009). Lanthanum concentrations in bone biopsy samples of patients being treated with pharmacological doses of lanthanum carbonate during 12 months revealed concentrations of $1.8\ \mu\text{g/g}$ wet weight (D'Haese et al., 2003) whilst in patients treated for 4–5 years the average measured bone lanthanum concentration was $5\ \mu\text{g/g}$ wet weight (unpublished data).

3.8. Tissue distribution of aluminium

In adults with normal renal function the total aluminium burden is estimated to be 30 mg with the highest concentrations found in the lungs, skeleton and skeletal muscles. Chronic accumulation occurs in patients with end-stage renal disease because the major elimination route; i.e. the kidney, does not function. In these patients a somewhat different distribution pattern is seen particularly in those taking aluminium-containing medication or being treated with aluminium-contaminated dialysis fluids. In these patients highest concentrations were observed in the liver, bone (up to $200\ \mu\text{g/g}$ wet weight), spleen and parathyroid glands (D'Haese, 1988; D'Haese et al., 1999; D'Haese and De Broe, 1999). Tissue distribution/elimination further depends on the element's concentration and speciation (van Ginkel et al., 1993). Data from

various studies indeed point toward a preferential transport of the circulating aluminium-transferrin complex to tissues expressing transferrin receptors such as the liver and the spleen. However, aluminium bound to citrate (or low molecular mass components in general) will in the presence of an intact renal function rapidly be excreted whereas in the absence of a renal function low molecular mass Al compounds will by preference be deposited at the bone mineralization surface, an area with no transferrin receptors (Van Landeghem et al., 1998).

3.9. Elimination of lanthanum

The kidneys are responsible for eliminating only a very small fraction of systemic lanthanum which also explains the lack of appreciable plasma lanthanum concentrations over time in uremic patients. Following intravenous administration of lanthanum chloride to rats, biliary excretion was the predominant route of elimination, with 85.6% of recovered lanthanum collected from bile over a period of 5 days. Experimental studies presented evidence for lanthanum to be transported and eliminated by the liver via a transcellular, endosomal-lysosomal-biliary canicular transport route (Bervoets et al., 2009).

3.10. Elimination of aluminium

Although only a small fraction of ingested aluminium is absorbed, and thus enters the blood compartment, it is vital that absorbed aluminium is quickly removed from the body because aluminium accumulation is a risk factor in a number of disorders (see below). A small amount of aluminium is excreted in the bile, but the major route of aluminium elimination is via the kidney. Hence it is not surprising that most healthy adults can tolerate large repeated daily oral aluminium exposure (up to 3500–7200 mg/day from e.g. antacids or buffered aspirin) without any adverse effect but that other people; i.e. preterm infants, young children and in particular patients with impaired renal function are at serious risk of aluminium accumulation/toxicity at even much lower daily doses (Willhite et al., 2014). *In vitro* determinations using artificial membranes indicated that $\pm 10\%$ of the total amount of circulating aluminium is filtered at normal plasma concentrations which is similar to the unbound aluminium fraction. However, when plasma aluminium is raised experimentally, its filterability falls, unless the excess aluminium is complexed with citrate whereby the aluminium citrate complex appears to be freely filtered. Information on tubular reabsorption of aluminium at normal plasma concentrations is inconsistent. Filtered aluminium appears to be at least partially reabsorbed, although the reabsorptive mechanisms remain speculative. A consensus is emerging that elevated plasma aluminium concentrations result in a fall in fractional aluminium reabsorption, and a recent micropuncture study indicates that under these circumstances the only significant site of aluminium reabsorption is the loop of Henle (Shirley and Lote, 2005).

4. General toxicity

4.1. Lanthanum

Although lanthanum has no known biological role, with the REE including lanthanum generally considered to be of low toxicity, and depending on its chemical form, the acute oral dose of lanthanum as assessed in rats varies from 3400 mg/kg body weight (lanthanum-ammonium nitrate) to > 10,000 mg/kg body weight (as lanthanum oxide) (Redling, 2006). Evaluation of potential genotoxicity using a range of *in vitro* assays in the presence and absence of post-mitochondrial fraction (S9) and *in vivo* in three

independent tests for mutagenicity and clastogenicity indicated that lanthanum is not genotoxic and that lanthanum carbonate is unlikely to present a latent hazard in therapeutic use (Damment et al., 2005). A single experimental study in mice reported nephrotoxic effects associated with oxidative stress through exposure to lanthanides. The most severe damage was induced by epigastric exposure to cerium chloride followed by neodymium chloride whilst only minor damage was seen with lanthanum chloride (Zhao et al., 2013). Results from another experimental study in mice suggested that these lanthanides enter hepatocytes and mainly accumulate in the nuclei and induce oxidative damage in hepatic nuclei and mitochondria (Huang et al., 2011). To what extent these observations are clinically relevant needs to be determined. Hormetic concentration-related trends, implying stimulatory or protective effects at low levels, then adverse effects at higher concentrations have been reported for lanthanum in various models including seedlings, bovine vascular smooth muscle cells and murine preosteoblast cells (Pagano et al., 2015).

4.2. Aluminium

Despite the ubiquity of aluminium in the environment and its presence in living organisms, albeit in small concentrations (ppb-ppm range), no biological function has so far been attributed. For this reason aluminium is considered to be a nonessential metal. Aluminium has long been considered inert for living organisms and as such was not regarded as a toxic element until the 1960s. At that time only a few reports dealt with the toxic effects of aluminium in humans and animals, which, however, did not receive much attention (Van Landeghem et al., 1998). In the 1970s, the element was linked to particular disease states noticed in patients with end-stage renal failure particularly those treated by dialysis (Alfrey et al., 1972, 1976; Berlyne et al., 1970). Although aluminium toxicity is mainly a matter of concern in dialysis patients and the element is adequately removed by the kidneys, occupationally exposed workers (e.g., welders) are also at risk for the deleterious effects of aluminium. In the latter population growing evidence is being provided for the element to cause pulmonary lesions (Kongerud and Søyseth, 2014; Raghu et al., 2014) as well as neurological disorders (Sińczuk-Walczak et al., 2003). Because of its persistence in the environment and the frequency of exposure of the general population, intensive research was conducted during the last decades to unravel the mechanism(s) underlying the element's potential health effects. Owing to its physicochemical characteristics aluminium has been reported to perturb iron homeostasis, disrupt biological membranes, enhance reactive oxygen species, and damage DNA (Exley, 2004, 2006b; Kumar et al., 2009; Mailloux et al., 2011; Zatta et al., 2002). Exposure of neurons and astrocytes to aluminium is known to activate apoptotic cascades, provoke cell cycle arrest, and interfere with cell signaling pathways (Drago et al., 2008; Lemire et al., 2009). Hence it is not surprising that during the last decade much attention has been paid to the potential neurotoxic effects of aluminium in humans and numerous groups assessed the aluminium content in the brain of subjects with various neurological disorders, in particular those with Alzheimer disease as well as non-affected individuals. Most of these data have been summarized by Exley and House (2011) reporting a normal range between 0.1 and 4.5 $\mu\text{g/g}$ dry weight with the higher values (>2 $\mu\text{g/g}$ dry weight) in brains of non-demented elderly, Alzheimer patients (up to 11.5 $\mu\text{g/g}$ dry weight), dialysis encephalopathy (up to 14.1 $\mu\text{g/g}$ dry weight – see below also), congophilic amyloid angiopathy (up to 23.0 $\mu\text{g/g}$ dry weight) and other encephalopathies (up to 47.4 $\mu\text{g/g}$ dry weight). Despite these numerous data the question as to whether in the general population, aluminium exposure is

either the cause, a potential contributor to the onset, progression and aggressiveness, or increased concentrations in the brain are the consequence of the neurological condition itself, with exception to dialysis encephalopathy, has been a matter of debate for many years (Kawahara and Kato-Negishi, 2011; Martyn et al., 1989; Willhite et al., 2014).

4.3. Therapeutic use

4.3.1. Lanthanum

Because of its low solubility, lanthanum carbonate was preferred to lanthanum chloride for further investigation on its therapeutic use as an intestinal phosphate binder. In the acidic environment of the stomach and upper small intestine, lanthanum dissociates sufficiently to become available for phosphate binding. *In vitro* more than 97% of phosphate was removed by a two-fold molar excess of lanthanum carbonate (Autissier et al., 2007). *In vivo* in a rat model with chronic renal failure, lanthanum carbonate was as effective as aluminium hydroxide and more effective than calcium carbonate or sevelamer (a polymeric amine that binds phosphate) at binding dietary phosphate at equivalent doses (Damment, 2011). In contrast to concurrent phosphate binding agents intestinal phosphate binding of lanthanum carbonate does not depend on variations in intestinal pH (Autissier et al., 2007). As biliary excretion is the major route of elimination of lanthanum and gastrointestinal absorption of the element is minimal, its therapeutic use in individuals with a compromised renal function does not expose them to an increased risk of systemic accumulation as compared to subjects with normal renal function. Long-term experimental studies in which rats with either chronic renal failure or normal renal function were administered lanthanum carbonate by oral gavage on a daily base at doses up to 2000 mg/kg (corresponding to a daily dose of 150 g/day in humans) did not show significant direct adverse effects on bone (Behets et al., 2005a, 2005b; Bervoets et al., 2006). In contrast to aluminium (see below), in bone, lanthanum could be localized at sites of active as well as non-active bone remodeling/mineralization with no association between histological deposition sites and the typical bone pathologies observed in renal failure (Behets et al., 2005c). As mentioned above, in the liver the localization of the element is lysosomal whilst lanthanum treatment during 20 weeks at a daily 1000 mg/kg/day dose was not accompanied by an increased concentration of liver enzymes (Yang et al., 2006; Bervoets et al., 2009). Following gavage (863 mg/kg/day during several weeks) or intravenous (a route enabling >300-fold higher plasma lanthanum concentrations) administration (0.03 mg/kg/day) of lanthanum, median brain concentrations remained near the lower limit of quantification (2.4 ng/g). This together with data from ultrastructural studies thus provide strong evidence that lanthanum does not cross the blood-brain barrier (Damment et al., 2009). In patients, lanthanum carbonate monotherapy was effective and well tolerated for up to 6 years with no evidence of safety concerns or increased frequency of adverse events (Hutchison et al., 2008). A 2-year follow up study in hemodialysis patients indicated that lanthanum carbonate as a phosphate binder did not adversely affect cognitive function compared with standard therapy (Altmann et al., 2007) whilst a bone-biopsy based study in dialysis patients receiving a median daily dose of 1250 mg elemental lanthanum/day showed an evolution towards normal bone histology and absence of aluminium-like effects (see below) on bone after 1-year treatment (D'Haese et al., 2003).

4.3.2. Aluminium

Being widely used in the past, aluminium hydroxide has proven to be a highly effective phosphate binder. Its substantial

gastrointestinal absorption and renal route of elimination however posed its target population for therapeutic use to an increased risk of accumulation. In the past aluminium-phosphate binder treatment, in particular when used in combination with aluminium-contaminated dialysis fluids (see below) led to the development of severe side-effects, mainly in the bone and brain. In bone, aluminium accumulates at the osteoid calcification front, a critical site of bone mineralization which at high exposure leads to the so-called aluminium-induced osteomalacia, a disease manifested by recurrent fractures and resistance to vitamin D therapy (Goodman, 1985; Verbueken et al., 1984). This type of bone disease is characterized by an increased amount of osteoid due to a defective mineralization. Another type of aluminium-related bone disease is adynamic bone characterized by a dramatically reduced bone turnover, and absence of osteoblasts, osteoclasts and osteoid (Goodman, 1985). Aluminium is unquestionably neurotoxic in patients treated by dialysis. The so-called dialysis encephalopathy syndrome, the result of acute intoxication of aluminium caused by the use of an aluminium-containing dialysate, was a common occurrence prior to 1980 (Rob et al., 2001; Ward et al., 1978). Although with the introduction of modern techniques of water purification, acute intoxication can now be avoided, occurrences of aluminium intoxication may still occur (D'Haese and De Broe, 1996; Simoes et al., 1994), and chronic moderately elevated concentrations may still be seen in dialysis centers in particular regions (Hou et al., 2010). The neurologic symptoms may be precipitated by concomitant ingestion of aluminium-containing phosphorus binders and citrate (D'Haese and De Broe, 2007). Onset or exacerbation of neurological disorders has been observed during deferoxamine therapy, presumably because of redistribution of mobilized deferoxamine-bound aluminium into the brain (Barata et al., 1996). Main symptoms are speech disturbances, tremor, epilepsy and an altered electroencephalopathic pattern while the serum aluminium concentrations usually exceed 100 µg/L (Van Ginkel, 2001). The disease progresses and ends mostly with the death of the patient within one year of initial symptoms. Although with the replacement of aluminium-based phosphate binders and adequate monitoring of dialysis fluids the major clinical manifestations have now disappeared, aluminium has also been implicated in more subtle diseases, such as microcytic hypochromic anemia, resistance to erythropoietin treatment and suppression of parathyroid hormone secretion. With regard to the latter, an increased effect of aluminium has been reported in the presence of a relative iron deficiency (Smans et al., 2000; Van Landeghem et al., 1997). Comparing different tissues of aluminium-intoxicated uremic patients, however, the relation between aluminium overload and toxicity is not straightforward. Whilst bone aluminium concentrations in aluminium-related bone disease, are distinctly elevated, comparable or even higher liver aluminium concentrations are seen without any apparent toxicity in humans. On the contrary, aluminium toxicity has been demonstrated in the brain at concentrations below 3 µg/g wet weight. This discrepancy points to the importance of the ultrastructural/subcellular localization of the element which determines its potential interference with physiological processes (Verbueken et al., 1984).

5. Potential exposure routes in water treatment

5.1. Lanthanum exposure and toxicity

Lanthanum is used as the active component of LMB, consisting of a bentonite carrier which holds the lanthanum cations within the clay interlayer where they retain their ability to bind with other ions such as phosphate and thus can be used to remove phosphorus from water bodies and reduce the incidence of algal blooms (Robb

et al., 2003). Some evidence indicates that 'free' (uncomplexed) lanthanum is toxic to some aquatic organisms (Reitzel et al., 2013a; Herrmann et al., 2016). Free metal ions are assumed to be responsible for detrimental effects, as in the widely used free ion activity model (FIAM) (Brown and Markich, 2000). Speciation modelling is needed for getting such insight in complexation and in which species are present, but it might not just be the aqua ion that is mobile or bioavailable. FIAM predictions are not always confirmed for aluminum (Gensemer and Playle, 1999). Lanthanum is highly reactive, easily giving up the $5d^1$ and $6s^2$ electrons and the free ion activity commonly will be very low in waters suffering from eutrophication, but not in low alkalinity water (see Fig. 4 in Spears et al., 2013). The biotic-ligand model assumes complexation of metals with reactive ligands on/in organisms, for instance by forming surface complexes at a metal transport sites on membranes (Niogi and Wood, 2004). Lanthanum, however, will not easily persist as free La^{3+} in serum, in cytoplasm or in natural surface water, which makes us a bit reluctant in referring to FIAM/BLM. We fully agree that speciation modelling is essential and this is also commonly applied in LMB research (e.g. Lürding et al., 2016; Lürding et al., 2016; Spears et al., 2013) and when aluminum is used.

Data from experimental studies where lanthanum was given as lanthanum chloride in drinking water to rats and mice suggested neurobehavioral impairment, although a clear dose-response relationship is often lacking (Briner et al., 2000; Damment et al., 2007a; Feng et al., 2006, 2006a; He et al., 2008; Yang et al., 2013; Zarros et al., 2013). Given (i) the maximal amounts at which lanthanum is leached into the water following application of LMB to surface waters either as filterable lanthanum (nominally <0.2 or $0.45 \mu\text{m}$), or total lanthanum or predicted 'free' (uncomplexed) ionic lanthanum (0.026 mg/L to 2.30 mg/L ; 0.002 mg/L to 0.14 mg/L and $<0.0004 \text{ mg/L}$ to 0.12 mg/L respectively) (Spears et al., 2013), (ii) the duration and magnitude of exposure to lanthanum-treated water, (iii) the ability of 'free' lanthanum to directly bind phosphate and other oxyanions in the intestine resulting in a low bioavailability and, (iv) the absence of significant toxic effects when used therapeutically during years at doses up to 3000 to 5000 times higher than those seen in lanthanum-treated water, one may reasonably accept that exposure to lanthanum via the drinking water or leisure activities will pose no increased risk for toxicity in humans even in patients with impaired renal function as biliary excretion is the major route of elimination of lanthanum. Nevertheless, when using lanthanum treated water to prepare dialysis fluids, even in dialysis centers equipped with the highest standards of water purification (carbon filtration, reverse osmosis, ultrafiltration, deionized water systems, ultraviolet TOC reduction/disinfection), co-measurement of lanthanum during regular monitoring of the in-house treated water and the dialysis fluid may be indicated as with dialysis treatment the gastrointestinal barrier is circumvented and a direct transfer of the element towards the blood compartment may occur, in particular for highly protein bound elements such as lanthanum.

It should be noted that lanthanum-modified bentonite is mostly used to counteract toxic cyanobacterial blooms, and that such blooms may pose potentially even more severe risks to bathers and definitely to patients receiving renal dialysis treatment as the 'Caruaru Syndrome' unambiguously elucidated with dozens of casualties (Azevedo et al., 2002; Jochimsen et al., 1998).

5.2. Aluminium exposure and toxicity

Varying concentrations of aluminium are present naturally in groundwater and surface water, including those used as sources of drinking water. The concentration of aluminium in surface water varies, ranging from 0.012 to 2.25 mg/L in North American rivers

(Jones and Bennett, 1986). Furthermore, aluminium (aluminium sulphate and polyaluminium chloride) has been used for more than three decades to inactivate phosphate from migrating from lake bed sediments to the overlying waters (Berkowitz et al., 2006; Cooke et al., 1993a; Lewandowski et al., 2003; Reitzel et al., 2005, 2013; Rydin and Welch, 1998; Rydin et al., 2000; Welch and Cooke, 1999; Welch et al., 1988).

Aluminium can liberate from alum due to changes of pH and the presence of low alkalinity water (Aziz et al., 2007; Paul et al., 2008). Water treatment has been reported to increase the percentage of dissolved, low molecular weight, chemically reactive and possibly more readily absorbed aluminium species (LaZerte et al., 1997). The toxicity of aluminium to fish has been well documented, in particular when the pH decreases to below 6 (Gensemer and Playle, 1999).

The hypothesis that aluminium exposure via drinking water is etiologically related to Alzheimer's disease has led to much debate. The possibility of such a relation was suggested by the presence of aluminium in senile plaques and neurofibrillary degeneration, two histologic lesions that are characteristic of the disease (Edwardson et al., 1992). Several studies have found that intake of aluminium (Praticò et al., 2002; El-Rahman, 2003) increases expression of amyloid protein in rodent tissues, a step that may be critical to the development of Alzheimer's disease. Ecotoxicological studies have suggested that concentrations of aluminium in drinking water of 0.1 – 0.2 mg/L may increase the risk of Alzheimer's disease, with relative risks or odds ratios ranging from 1.35 to 2.67 (Gauthier et al., 2000; Martyn et al., 1989; Rondeau et al., 2000).

With regard to the individual exposure via drinking water, reports have shown a high daily intake of aluminium ($>0.1 \text{ mg/day}$) to be significantly associated with an increased risk of dementia. Conversely, a concomitant intake of aluminium with an increase of 10 mg/day in silica intake via drinking water was associated with a reduced risk of dementia (Rondeau et al., 2009).

Until the early 1980s aluminium in the dialysate appeared to be the major source of the metal in chronic renal failure patients who developed aluminium toxicity (Wills and Savory, 1985). As at that time adequate water purification systems were not available in all dialysis units, the aluminium concentration of the dialysate depended primarily on the aluminium concentration of the water with which it was prepared; whether further enriched with aluminium-contaminated chemicals or not in the concentrates which are added to the water to prepare the final dialysis fluids (D'Haese et al., 1990). With the introduction of modern water purification systems in the dialysis centers the incidence of caricatural aluminium intoxication has now disappeared. Nevertheless, as the concentration gradient between the dialysate aluminium and the non-protein bound aluminium fraction ($<20\%$) in the serum compartment is the driving force for aluminium transfer during hemodialysis, chronic accumulation in a patient with a serum aluminium concentration of e.g. $10 \mu\text{g/L}$ theoretically may still occur in the presence of a dialysate aluminium accumulation as low as $3 \mu\text{g/L}$. Moreover, accidental intoxications cannot be excluded (Berend et al., 2004; D'Haese and De Broe, 1999a).

In Curaçao, in order to protect a water distribution pipe supplying water to a dialysis center from corrosion, the pipe was internally lined with a cement mortar. Because of the aggressiveness of the distilled water, calcium and aluminium leached from the cement mortar into the water used to prepare dialysate causing a possible hard water syndrome and definite acute aluminium encephalopathy resulting in the death of 10 patients (Berend et al., 2001). In the South of Portugal, the low rainfall in the early 1990s resulted in a subsequent decrease in the available water sources resulting in high concentrations of suspended particles which in

turn necessitated the addition of alum as a flocculating/coagulating agent. The passage of this contaminated water through the water purification installation of a hemodialysis center resulted in the obstruction of the cartridge filters and malfunction of the reverse osmosis membranes. Finally insufficiently treated water was sent via dialysis to the patients. This led to acute aluminium intoxication, manifested by the epidemic appearance of encephalopathy, microcytic anemia and death of 18 patients (Barata et al., 1996; Simoes et al., 1994).

6. Importance of chemical speciation and effects on toxicity

6.1. Lanthanum

Substantial information exists on the aqueous chemistry and speciation of the REE/lanthanides. Geochemical modelling with the chemical equilibrium model MINEQL + indicated that dissolved lanthanides (Ln) are complexed mainly to carbonates and dissolved organic matter. In the aqueous phase, the relative abundance of the free ion, LnCO_3^{2+} , and humic complexes decreases from lanthanum to lutetium, whereas the relative abundance of $\text{Ln}(\text{CO}_3)_2^+$ increases (Moermond et al., 2001). As is the case for any element, the toxicity of lanthanum primarily depends on the inorganic salt (anion) with which it occurs, with LD_{50} values for oral doses in rats and mice varying between 2354 and > 10,000 mg/kg body weight for lanthanum chloride (LaCl_3) and lanthanum oxide (La_2O_3) respectively (Shimomura et al., 1980; Cochran et al., 1950). With regard to the use of LMB, concern has been raised regarding the potential for release of filterable lanthanum in lakes and surface waters and the potential unintended ecological implications of this release (Lürling and Tolman, 2010; Spears et al., 2013). The speciation of filterable lanthanum ions is also important when considering the ecotoxicological impact and of all filterable lanthanum species (i.e. La^{3+} , $\text{La}(\text{OH})_2^+$, and $\text{La}(\text{OH})_2^+$). The La^{3+} cation carries the greatest risk of biological effects (Das et al., 1988). In humans, once absorbed, lanthanum circulates >99.7% protein bound in plasma (Damment and Pennick, 2007, 2008). High protein binding, to a certain extent, may explain (i) the low toxicity profile of lanthanum, as it is unlikely for the lanthanum-protein complex to cross the blood-brain barrier, incorporate in the calcified bone matrix or interfere with the various ionized calcium-regulated cell biological functions, and (ii) the almost unique biliary elimination of the element after transferrin-mediated endocytosis by the hepatocyte (Bervoets et al., 2009).

6.2. Aluminium

In the environment as well as in the human body aluminium occurs in various chemical species which have different physical, chemical and biological properties (Harris et al., 1996; Van Landeghem et al., 1998; Yokel and McNamara, 2001). The chemical speciation of aluminium in drinking water is of particular interest, as the form of aluminium regulates its solubility, bioavailability and toxicity. Absorption from the gut depends largely on the presence of complexing ligands, particularly carboxylic acids, with which the metal can form absorbable neutral aluminium species.

One factor determining the form of aluminium in water is pH. In raw water with low concentrations of dissolved organic compounds such as humic and fulvic acids, the dependence of dissolved aluminium concentration on pH resembles a parabola with a sharp solubility minimum at around pH 6.5 (Driscoll and Letterman, 1995, Fig. 1). The solubility of aluminium increases at lower pH owing to the formation of $\text{Al}(\text{OH})_2^+$, $\text{Al}(\text{OH})_2^+$ and $\text{Al}(\text{H}_2\text{O})_6^{3+}$ - often abbreviated as Al^{3+} and sometimes referred to in the literature as 'free'

aluminium. The solid $\text{Al}(\text{OH})_3$ is the predominant species between pH 5.2 and 8.8, whereas the soluble $\text{Al}(\text{OH})_4^-$ predominates above pH 9 (Martell and Motekaitis, 1989, Fig. 1).

The form in which aluminium is present in drinking water is also dependent on whether the water is fluoridated, as fluoride has a strong affinity for aluminium, particularly under acidic conditions (Nieboer et al., 1995). When alum is added to raw water for treatment, the form of aluminium changes along a number of pathways, depending on the quantity of alum added, the temperature, the pH, the types and concentrations of dissolved materials as well as the types and surface area of particulate matter present (Driscoll and Letterman, 1988).

Concomitant intake of aluminium hydroxide with citrate has been demonstrated to increase gastrointestinal absorption of the element which was reported to occur in the proximal bowel via the paracellular pathway due to the ability of citrate to open the epithelial tight junctions (Froment et al., 1989). On the other hand, dissolved silicon has been regarded as an important factor in limiting the absorption of dietary aluminium (Edwardson et al., 1993; Parry et al., 1998). Once absorbed in the serum compartment, aluminium strongly binds to proteins mainly transferrin, the remaining ultrafiltrable fraction to circulate as either bound to phosphate or citrate. Within the serum compartment it has been demonstrated that aluminium may compete with iron for transferrin binding, which to a certain extent also determines the tissue deposition and toxicity of aluminium (Van Landeghem et al., 1997, 1998a; Smans et al., 2000). In contrast to the serum compartment in the brain aluminium occurs as a non-protein bound, low molecular mass, probably silicate compound. This latter finding is supported by the high molar ratio of both citrate/transferrin and silica/transferrin in cerebrospinal fluid. The occurrence of 'free' aluminium might also explain the element's high toxicity at very low concentrations and gives rise to a hypothesis to explain discrepancies in the neurotoxic effects of aluminium in dialysis dementia and Alzheimer's disease (Van Landeghem et al., 1997a).

7. Exposure pathways – practical considerations

Application of either lanthanum- or aluminium-containing compounds to natural waters to reduce the concentration of dissolved phosphorus will result in a range of potential exposure pathways to in-situ and transient biota and humans over a range of temporal (including acute and chronic exposure) and spatial scales. We consider the main exposure routes for humans to lanthanum or aluminium in treated waterbodies to be through drinking treated water, consuming biota (e.g. crustaceans, fish and plants), dermal exposure via water or sediment and through the inadvertent consumption of sediments particularly in young children. We describe the likelihood of human health effects associated with realistic exposure rates below for lanthanum and aluminium.

7.1. Water

During application of LMB the maximum reported total and filterable lanthanum concentrations in the surface water of 16 treated lakes were up to 2.3 mg/L and 0.4 mg/L, respectively (Spears et al., 2013). Smeltzer et al. (1999) reported dissolved aluminium concentrations in Lake Morey (USA) of up to 0.2 mg/L. Reitzel et al. (2013) determined experimentally that dissolved aluminium concentrations may reach 0.85 mg/L following an application, as a result of diffusion from bed sediments back to the water column following settlement. Wauer and Teien (2010) reported maximum concentrations of reactive aluminium of 2.0 mg/L in field observations. Given that lanthanum as lanthanum carbonate when used therapeutically (Fosrenol[®], Shire Pharmaceuticals) is administered

at doses up to 1500 mg per day (~900 mg lanthanum/day) without toxicity to the patients after up to 10 years of treatment (Hutchison et al., 2016), consumers would have to consume at least 390 L of the surface water per day to attain a similar, nominally safe dose. Using the highest application dose of 333 mg/L LMB (Spears et al., 2013) a consumption of 54 L is needed to attain a dose of 900 mg lanthanum. It is likely that in the absence of episodic resuspension, the highest risk of exposure will occur within the following few days to weeks.

7.2. Sediment

Following settling of the applied phosphorus removal agent, the bed sediments are the location of treated waterbodies where lanthanum concentrations are highest following an application. Consumption of these sediments, although unlikely, may be considered, for example, playing children may express some geophagia, whereas pica disorder may result in considerable consumption of soil material (Rose et al., 2000). Soil pica is referred to as eating 500 mg to more than 50 g of soil per day (Callahan, 2003). Using the data presented in Table 2 in Spears et al. (2013) an average dose of 348 g LMB/m² (range 6–667) is derived. With an assumed maximum 5% weight of lanthanum in LMB this makes an average of 17.4 g lanthanum/m² (range 0.3–33.3) or 1.74 mg lanthanum/cm². Using the data presented by Reitzel et al. (2013) applied aluminium concentrations in bed sediments may reach 54 g aluminium/m², although Wauer and Teien (2010) report sediment aluminium concentrations of up to 200 g aluminium/m². Assuming a specific density of 1 g/cm³ and a thickness of 1 cm, a person would need to consume 860 g of sediment to reach a nominally safe dose of 1500 mg lanthanum, or 450 g using the highest lanthanum application dosing (3.33 mg lanthanum/cm²).

Based on the Tolerable Weekly Intake (TWI) of 1 mg aluminium per kg body-weight as proposed by the European Food and Safety Authority 2008 (https://www.efsa.europa.eu/sites/default/files/corporate_publications/files/ar08en%2C0.pdf), a 60 kg person would need to eat 3–11 g of sediment following sediment aluminium content of Reitzel et al. (2013a) and Wauer and Teien (2010), respectively, to reach this TWI.

7.3. Crustaceans

Accumulation of lanthanum in the crustacean zooplankton *Daphnia magna* has been observed (Yang et al., 1999), yet these small animals are not directly consumed by humans. Nonetheless, they may provide a food chain vector of transmission via fish that may predate heavily on zooplankton. The most obvious human exposure route is via bottom dwelling crustaceans, such as crayfish that may be exposed for prolonged periods to relatively high concentrations of LMB. In Lake Rauwbraken (The Netherlands) the lanthanum concentration in the flesh of crayfish (*Orconectes limosus*) increased from 0.12 ± 0.05 µg lanthanum/g dry-weight before application to 89 ± 42 µg lanthanum/g dry weight in animals caught 4 months after application and 37 ± 13 µg lanthanum/g dry weight in animals collected 14 months after treatment. In practical terms, this means that a person would have to consume daily 10 kg of crayfish with 89 µg lanthanum/g and 24 kg of crayfish with 37 µg lanthanum/g to reach the recommended therapeutic prescription dose of lanthanum carbonate. In a controlled laboratory experiment, exposure of marbled crayfish (*Procambarus fallax* f. *virginialis*) to 67 g LMB/m² led to a maximum lanthanum concentration in the flesh of 13.6 µg lanthanum/g dry weight (van Oosterhout et al., 2014). Consequently, a daily consumption of 66 kg crayfish would equal the safe recommend therapeutic dose of Fosrenol®. It should be noted that the lanthanum concentration is expressed per unit

dry-weight that only constitutes approximately 17% of the live-weight and thus daily crayfish consumption would need to be almost 390 kg to attain the nominally safe adult medication dose.

Data on aluminium concentrations in crustacea are sparse in relation to whole lake applications. However, Elangovan et al. (1999) reported uptake of aluminium into in the aquatic isopod *Asellus aquaticus* to reach about 2.4 mg aluminium/g dry weight and that aluminium uptake into tissue of the animal can be explained through a significant linear regression relationship with the aluminium concentration in the water column.

7.4. Fish

Data on La concentrations in edible fish parts after LMB applications are rare. Some lanthanum accumulation in the liver and hepatopancreas of fish collected from Lake Okareka (New Zealand) has been reported, while lanthanum in the flesh of trout and koura remained below the level of detection (Landman et al., 2007). Lanthanum concentrations in the livers of eel caught two years after a whole lake treatment showed a 94 fold increase compared to pre-intervention liver concentrations and a 133 fold increase in eels caught five years later (Waajen et al., 2017). Elevated lanthanum concentrations were also found in the flesh of fish collected before and after a whole lake application in Lake De Kuil, The Netherlands (Waajen et al., 2016), and from LMB treated and control compartments constructed in urban ponds (Waajen et al., 2016a, 2017). In Lake De Kuil, prior to application, the mean lanthanum concentration in the flesh of the five most abundant fish species (bream, eel, perch, pike, tench) was 0.03 ± 0.02 µg lanthanum/g dry weight (Waajen et al., 2017). Two years after the LMB application at a dose of 593 g LMB/m² (Waajen et al., 2016), the lanthanum concentration in the muscle tissue of these five species had increased to an average of 0.10 ± 0.05 µg lanthanum/g dry weight and after 5 years the lanthanum had returned to pre-application concentrations (0.03 ± 0.01 µg lanthanum/g dry weight) in specimens of these five fish species. In two urban ponds in The Netherlands 300 m² (pond Dongen) and 400 m² sized compartments (pond Eindhoven) were constructed of which some were treated with 750 g LMB/m² and 1130 g LMB/m², respectively (Waajen et al., 2016a). The muscle tissue of fish collected after two years in the LMB treated compartments in pond Dongen contained on average 0.22 ± 0.34 µg lanthanum/g dry weight, while that of non-LMB exposed fish was on average 0.06 ± 0.09 µg lanthanum/g dry weight. In pond Eindhoven, muscle tissue in LMB exposed fish was 0.07 ± 0.06 µg lanthanum/g dry weight, while that of non-LMB exposed fish was 0.03 ± 0.02 µg lanthanum/g dry weight. Although lanthanum from LMB thus may accumulate in fish and raised lanthanum concentrations have been reported up to five years following treatment with the highest lanthanum concentration found in liver, no toxic effects were observed following LMB bentonite treatments (Waajen et al., 2017).

Taking the highest flesh lanthanum concentrations measured of all examined fish (0.81 µg lanthanum/g dry weight) a person would have to consume more than 1000 kg of fish per day to reach the nominally safe recommended therapeutic dose of lanthanum in adults.

Wauer and Teien (2010) reported concentrations of aluminium in the gills of perch (*Perca fluviatilis*) and ruffe (*Gymnocephalus cernuus*) up to about 100 µg aluminium/g dry weight and 401 µg aluminium/g dry weight respectively, although no similar observations were reported for roach (*Rutilus rutilus*), bream (*Abramis brama*), or silver carp (*Hypophthalmichthys molitrix*). Given the >100-fold higher aluminium concentrations in combination with a 10–100 higher gastrointestinal absorption as compared to lanthanum (see above) potential aluminium toxicity should be

Table 1
Maximum reported concentrations for lanthanum (La) and aluminium (Al) in various abiotic and biotic components of lakes following additions of LMB or aluminium containing salts or waste waters, where data from eutrophication control studies were not available in the peer reviewed literature.

Exposure route	Upper concentrations reported	Period of effect	Reference	Likelihood of exceeding exposure limits	Severity of impact	Risk rating
Water column						
	0.4 mg FLa/L	<12 months	Spears et al. (2013)	Low	Mod-High	Low
	0.2 mg FAl/L	>30 days	Smeltzer et al., 1999	Low	Mod-High	Low
Bed sediments						
	33.3 g La/m ²	Unknown	Spears et al. (2013)	Low	Mod-High	Low
	200 g Al/m ²	Unknown	Wauer and Teien (2010)	Low	Mod-High	Low
Crustacea						
<i>Orconectes limosus</i>	131 µg La/g DW	>14 months		Low	Mod-High	Low
<i>Asellus aquaticus</i>	2400 µg Al/g DW	<1 month	Elangovan et al. (1999)	Low	Mod-High	Low
Fish						
<i>Perca fluviatilis</i>	0.56 µg La/g DW	5 years	Waajen et al. (2017)	Low	Mod-High	Low
<i>Gymnocephalus cemus</i>	401 µg Al/g DW	Unknown	Wauer and Teien (2010)	Low	Mod-High	Low
Plants						
<i>Elodea nuttallii</i>	871 µg La/g DW	Unknown	Waajen et al. (2017)	Low	Mod-High	Low
<i>Lemna minor</i>	17 mg Al/g DW	Unknown	Goulet et al. (2005)	Low	Mod-High	Low

considered. In this context it is worth to be mentioned that given the fact that aluminium is eliminated via the kidney, the risk for toxic effects, even with this degree of exposure is rather limited in subjects with normal renal function. Nevertheless regular monitoring of aluminium in the drinking water and consumable fish by water and health authorities should be performed. In individuals with impaired renal function regularly consuming local fish or water, serum aluminium measurement is recommended, particularly in patients presenting with undefined bone and/or neurologic complaints or signs of anemia. Here the first line of therapy should consist in the withdrawal of these sources of aluminium exposure.

7.5. Plants

Aquatic plants may take up lanthanum and a lanthanum bioaccumulation factor of 138 for duckweed has been reported (Yang et al., 1999). The aquatic macrophyte *Elodea nuttallii* was harvested on three occasions from the different compartments in the above-mentioned urban waters (Waajen et al., 2016a). Samples consisting of the complete plants including the roots and shoots were analysed for lanthanum (Waajen et al., 2017). The mean La concentration of *E. nuttallii* from control compartments varied between 0.35 and 7.03 µg lanthanum/g dry weight at pond Dongen and between 0.14 and 13.53 µg lanthanum/g dry weight at pond Eindhoven, the stocking plant material at the start of the experiment contained on average 7.50 µg lanthanum/g dry weight. The mean lanthanum concentration of *E. nuttallii* from the LMB treated compartments reached up to 380 µg lanthanum/g at pond Dongen and 871 µg lanthanum/g at pond Eindhoven. Although consumption of macrophytes by humans is restricted to a few species (e.g., lotus, water chestnut, water caltrop, water spinach, watercress) and *E. nuttallii* is normally not one of the consumed aquatic plants, a person still would need to ingest 2.4 kg per day of dry plant equivalent from Dongen and 1 kg of dried plant equivalent per day from pond Eindhoven to attain the nominally safe lanthanum dose recommended for therapeutic purposes. Use of harvested LMB exposed plant material in fish feed (Hasan and Chakrabarti, 2009) or cattle fodder (Banerjee and Matai, 1990; Goopy and Murray, 2003), however, warrants caution as no information on the magnitude of possible La transfer and bioaccumulation exists.

Data on macrophyte uptake of aluminium following a whole lake treatment do not appear to be available in the peer reviewed literature. However, we draw here on data published by Goulet et al. (2005) on a series of phytoremediation mesocosm

experiments (at water concentrations of up to 1 mg total dissolved aluminium/L) in which aluminium uptake across 4 different macrophyte species (i.e. *Potamogeton epihydrus*, *Nuphar variegatum*, *Lemna minor* and *Typha latifolia*) ranged between <0.01 and 17.2 mg aluminium/g dry weight for root tissues; 0.34 and 1.44 mg aluminium/g dry weight for stem tissues and 0.18 and 6.25 mg aluminium/g dry weight for leaf tissues (see Table 1).

8. General conclusions

- Because of their inherently strong phosphate binding capacity over a wide range, and mostly overlapping set of physicochemical conditions, application of aluminium- and lanthanum-based compounds has proven to be efficacious for water treatment and therapeutic phosphate control in uremic patients.
- Kinetics as well as mechanisms underlying the possible toxic effects of aluminium and lanthanum substantially differ from each other
- Although aluminium and lanthanum have physicochemical similarities their aqueous chemistry differs with hydrolysis of lanthanum occurring at substantially higher pH than that of aluminium
- The speciation, concentration and exposure pathways to living organisms of lanthanum and aluminium is strongly dependent on pH and salinity
- The extent of lanthanum leached into LMB-treated water reported so far, do not exceed concentrations that might be considered harmful, however, considerable care in manufacture and quality control needs to be exercised to minimize risk to receiving aquatic environments.
- Regular monitoring of both aluminium and lanthanum concentrations in lanthanum-/aluminium-treated water by the responsible authorities is recommended to avoid accidental acute or chronic lower level exposure

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References

- Alfrey, A.C., Mishell, J.M., Burks, J., 1972. Syndrome of dyspraxia and multifocal seizures associated with chronic hemodialysis. *Trans ASAIO* 18, 257–261.
- Alfrey, A.C., LeGendre, G.R., Kaehny, W.D., 1976. The dialysis encephalopathy syndrome. Possible Al intoxication. *N. Engl. J. Med.* 294, 184–188.
- Altmann, P., Barnett, M.E., Finn, W.F., SPD405-307 Lanthanum Carbonate Study Group, 2007. Cognitive function in Stage 5 chronic kidney disease patients on hemodialysis: no adverse effects of lanthanum carbonate compared with standard phosphate-binder therapy. *Kidney Int.* 71 (3), 252–259.
- Anderson, M.A., Berkowitz, J., 2010. Aluminium polymers formed following alum treatment of lake water. *Chemosphere* 81 (7), 832–836.
- Autissier, V., Damment, S.J., Henderson, R.A., 2007. Relative in vitro efficacy of the phosphate binders lanthanum carbonate and sevelamer hydrochloride. *J. Pharmacol. Sci.* 96 (10), 2818–2822.
- Azevedo, S.M., Carmichael, W.W., Jochimsen, E.M., Rinehart, K.L., Lau, S., Shaw, G.R., Eaglesham, G.K., 2002. Human intoxication by microcystins during renal dialysis treatment in Caruaru-Brazil. *Toxicology* 181–182, 441–446.
- Aziz, H.A., Alias, S., Assari, F., Adlan, M.N., 2007. The use of alum, ferric chloride and ferrous sulphate as coagulants in removing suspended solids, colour and COD from semi-aerobic landfill leachate at controlled pH. *Waste Manag. Res.* 25 (6), 556–565.
- Banerjee, A., Matai, S., 1990. Composition of Indian Aquatic Plants in Relation to Utilization as Animal Forage. *J. Aqua. Plant Manag.* 28, 69–73.
- Barata, J.D., D'Haese, P.C., Pires, C., Lamberts, L.V., Simões, J., De Broe, M.E., 1996. Low-dose (5 mg/kg) desferrioxamine treatment in acutely aluminium-intoxicated haemodialysis patients using two drug administration schedules. *Nephrol. Dial. Transplant.* 11 (1), 125–132.
- Behets, G., 2005. Lanthanum Carbonate and Bone. PhD-Thesis. Universities of Antwerp, (Belgium)/Leiden (The Netherlands). ISBN-13 978-90-9019934-4, 10 90-9019934-9.
- Behets, G.J., Verberckmoes, S.C., Oste, L., Bervoets, A.R., Salomé, M., Cox, A.G., Denton, J., De Broe, M.E., D'Haese, P.C., 2005a. Localization of lanthanum in bone of chronic renal failure rats after oral dosing with lanthanum carbonate. *Kidney Int.* 67 (5), 1830–1836.
- Behets, G.J., Gritters, M., Dams, G., De Broe, M.E., D'Haese, P.C., 2005b. Effects of efficient phosphate binding on bone in chronic renal failure rats. *Ren. Fail.* 27 (4), 475–484.
- Behets, G.J., Verberckmoes, S.C., Oste, L., Bervoets, A.R., Salomé, M., Cox, A.G., Denton, J., De Broe, M.E., D'Haese, P.C., 2005c. Localization of lanthanum in bone of chronic renal failure rats after oral dosing with lanthanum carbonate. *Kidney Int.* 67 (5), 1830–1836.
- Berend, K., van der Voet, G.B., Boer, W.H., 2001. Acute aluminium encephalopathy in a dialysis center caused by a cement mortar water distribution pipe. *Kidney Int.* 59, 746–753.
- Berend, K., Knoops, G.J., De Wolff, F.A., 2004. Prosecution after an outbreak of subacute aluminium intoxication in a hemodialysis center. *Leg Med. (Tokyo)* 6 (1), 1–10.
- Berkowitz, J., Anderson, M.A., Amrhein, C., 2006. Influence of aging on phosphorus sorption to alum floc in lake water. *Water Res.* 40 (5), 911–916.
- Berlyne, G.M., Ben Ari, J., Pest, D., Weinberger, J., Stern, M., Levine, R., Gilmore, G.R., 1970. Hyperaluminemia from Al resins in renal failure. *Lancet* 671, 494–496.
- Bervoets, A.R., Oste, L., Behets, G.J., Dams, G., Blust, R., Marynissen, R., Geryl, H., De Broe, M.E., D'Haese, P.C., 2006. Development and reversibility of impaired mineralization associated with lanthanum carbonate treatment in chronic renal failure rats. *Bone* 38 (6), 803–810.
- Bervoets, A.R., Behets, G.J., Schryvers, D., Roels, F., Yang, Z., Verberckmoes, S.C., Damment, S.J., Dauwe, S., Mubiana, V.K., Blust, R., De Broe, M.E., D'Haese, P.C., 2009. Hepatocellular transport and gastrointestinal absorption of lanthanum in chronic renal failure. *Kidney Int.* 75 (4), 389–9.
- Bolle, F., Brian, W., Petit, D., Boutakhrif, K., Feraille, G., Van Loc, J., 2011. Tea brewed in traditional metallic teapots as a significant source of lead, nickel and other chemical elements. *Food Addit. Contam. Part A Chem Anal Control Expo Risk Assess* 28 (9), 1287–1293.
- Bondy, S.C., 2016. Low levels of aluminium can lead to behavioral and morphological changes associated with Alzheimer's disease and age-related neurodegeneration. *Neurotoxicology* 52, 222–229.
- Briner, W., Rycek, R.F., Moellenberndt, A., Dannull, 2000. Neurodevelopmental effects of lanthanum in mice. *Neurotoxicol. Teratol.* 22 (4), 573–581.
- Brown, P.L., Markich, S.J., 2000. Evaluation of the free ion activity model of metal-organism interaction: extension of the conceptual model. *Aquat. Toxicol.* 51 (2), 177–194.
- Callahan, G.N., 2003. Eating dirt. *Emerg. Infect. Dis.* 9 (8), 1016–1021.
- Cetiner, Z.S., Wood, S.A., Gammons, C.H., 2005. The aqueous geochemistry of the rare earth elements. Part XIV. The solubility of rare earth element phosphates from 23 to 150. *Chem. Geol.* 217, 147–169.
- Cetiner, Z.S., Xiong, Y., 2008. Chemical controls on the solubility, speciation and mobility of lanthanum at near surface conditions: a geochemical modeling study. *Appl. Geochem.* 23, 2301–2315.
- Cochran, K.W., Doull, J., Mazur, M., Dublois, K.P., 1950. Acute toxicity of zirconium, columbium, strontium, lanthanum, cesium, tantalum and yttrium. *Arch. Ind. Hyg. Occup. Med.* 1 (6), 637–645.
- Cooke, G.D., Welch, E.B., Peterson, S.A., Newroth, P.R., 1993. In: Restoration and Management of Lakes and Reservoirs, second ed. CRC Lewis Publishers, p. 548.
- Cooke, G.D., Welch, E.B., Martin, A.B., Fulmer, D.G., Hyde, J.B., Schriever, G.D., 1993a. Effectiveness of Al, Ca, and Fe salts for control of internal phosphorus loading in shallow and deep lakes. *Hydrobiologia* 253, 323–335.
- Copetti, D., Finsterle, K., Marziali, L., Stefani, F., Tartari, G., Douglas, G., Reitzel, K., Spears, B.M., Winfield, I.J., Crosa, G., D'Haese, P., Yasseri, S., Lüring, M., 2016. Eutrophication management in surface waters using lanthanum modified bentonite: a review. *Water Res.* 97, 162–174.
- Crisponi, G., Fanni, D., Gerosa, C., Nemolato, S., Nurchi, V.M., Crespo-Alonso, M., Lachowicz, J.L., Faa, G., 2013. The meaning of aluminium exposure on human health and aluminium-related diseases. *Biomol. Concepts* 4 (1), 77–78.
- Damment, S.J., Beevers, C., Gatehouse, D.G., 2005. Evaluation of the potential genotoxicity of the phosphate binder lanthanum carbonate. *Mutagenesis* 20 (1), 29–37.
- Damment, S.J., Pennick, M., 2007. Systemic lanthanum is excreted in the bile of rats. *Toxicol. Lett.* 171 (1–2), 69–77.
- Damment, S.J., De Broe, M.E., D'Haese, P.C., Bramall, N., Cox, A.G., McLeod, C.W., 2007. Incredible effects of lanthanum? *Toxicol. Lett.* 168 (2), 186–189.
- Damment, S.J., Pennick, M., 2008. Clinical pharmacokinetics of the phosphate binder lanthanum carbonate. *Clin. Pharmacokinet.* 47 (9), 553–563.
- Damment, S.J., Cox, A.G., Secker, R., 2009. Dietary administration in rodent studies distorts the tissue deposition profile of lanthanum carbonate; brain deposition is a contamination artefact? *Toxicol. Lett.* 188 (3), 223–229.
- Damment, S.J., 2011. Pharmacology of the phosphate binder, lanthanum carbonate. *Ren. Fail.* 33 (2), 217–224.
- Das, T., Sharma, A., Talukder, G., 1988. Effects of lanthanum in cellular systems. A review. *Biol. Trace Elem. Res.* 18, 201–228.
- de Vincente, I., Huang, P., Andersen, F., Jensen, H., 2008. Phosphate adsorption by fresh and aged aluminum hydroxide: consequences for Lake Restoration. *Environ. Sci. Technol.* 42 (17), 6650–6655.
- D'Haese, P.C., 1988. Aluminium Accumulation in Patients with Chronic Renal Failure. Monitoring, Diagnosis and Therapy. PhD Thesis. Universities of Antwerp, (Belgium)/Amsterdam (The Netherlands).
- D'Haese, P.C., Lamberts, L.V., Lian, L.A., Boone, L.P., Van Waeleghem, J.P., De Broe, M.E., 1990. Contribution of parenteral and dialysate solutions to the aluminium accumulation in dialysis patients. *Blood Purif.* 8 (6), 359–362.
- D'Haese, P.C., De Broe, M.E., 1996. Adequacy of dialysis: trace elements in dialysis fluids. *Nephrol. Dial. Transplant.* 11 (Suppl. 2), 92–97.
- D'Haese, P.C., Couttenye, M.M., Lamberts, L.V., Elseviers, M.M., Goodman, W.G., Schrooten, I., Cabrera, W.E., De Broe, M.E., 1999. Aluminium, iron, lead, cadmium, copper, zinc, chromium, magnesium, strontium, and calcium content in bone of end-stage renal failure patients. *Clin. Chem. Sep.* 45 (9), 1548–1556.
- D'Haese, P.C., De Broe, M.E., 1999. Trace metals in chronic renal failure patients treated by dialysis: a review. *Trace Elem. Med.* 16, 163–174.
- D'Haese, P.C., Spasovski, G.B., Sikole, A., Hutchison, A., Freemont, T.J., Sulkova, S., Swanepoel, C., Pejanovic, S., Djukanovic, L., Balducci, A., Coen, G., Sulowicz, W., Ferreira, A., Torres, A., Curic, S., Popovic, M., Dimkovic, N., De Broe, M.E., 2003. A multicenter study on the effects of lanthanum carbonate (Fosrenol) and calcium carbonate on renal bone disease in dialysis patients. *Kidney Int.* 85 (Suppl. 1), 73–78.
- D'Haese, P.C., De Broe, M.E., 2007. Aluminium, lanthanum, and strontium. In: Daugirdas, J.T., Blake, P.G., Ing, T.S. (Eds.), *Handbook of Dialysis*, fourth ed. Lippincott Williams & Wilkins, pp. 714–726.
- Dithmer, L., Nielsen, U.G., Yasseri, S., Lundberg, D., Moore, A., Jensen, N.D., 2016. Responses in sediment phosphorus and lanthanum concentrations and composition across 10 lakes following applications of lanthanum modified bentonite. *Water Res.* 97, 101–110.
- Douglas, G.B., 2002. Remediation Material and Remediation Process for Sediments. US Patent, p. 6350383.
- Douglas, G.B., Robb, M.S., Coad, D.N., Ford, P.W., 2004a. A review of solid phase adsorbents for the removal of phosphorus from natural and waste waters. In: Valsami-Jones, E. (Ed.), *Phosphorus in Environmental Technology – Removal, Recovery, Application*. Chapter 13. IWA Publishing, pp. 291–320.
- Douglas, G.B., Robb, M.S., Ford, P.W., 2004b. Reassessment of the performance of mineral-based sediment capping materials to bind phosphorus: a comment on Akhurst et al. *Mar. Freshw. Res.* 59, 836–837.
- Drago, D., Cavaliere, A., Mascetra, N., Ciavardelli, D., di Ilio, C., Zatta, P., Sensi, S.L., 2008. Aluminium modulates effects of beta amyloid(1–42) on neuronal calcium homeostasis and mitochondria functioning and is altered in a triple transgenic mouse model of Alzheimer's disease. *Rejuvenation Res.* 11, 861–871.
- Driscoll, C.T., Letterman, R.D., 1988. Chemistry and fate of Al(III) in treated drinking water. *J. Environ. Eng.* 114 (1), 21–37.
- Driscoll, C.T., Letterman, R.D., 1995. Factors regulating residual aluminium

- concentrations in treated waters. *Environmetrics* 3, 287–309.
- Drüeke, T.B., 2002. Intestinal absorption of aluminium in renal failure. *Nephrol. Dial. Transplant.* 17 (Suppl. 2), 13–16.
- Edwardson, J.A., Candy, J.M., Ince, P.G., McArthur, F.K., Morris, C.M., Oakley, A.E., Taylor, G.A., Bjertness, E., 1992. Aluminium accumulation, beta-amyloid deposition and neurofibrillary changes in the central nervous system. *Ciba Found. Symp.* 169, 165–179.
- Edwardson, J.A., Moore, P.B., Ferrier, I.N., Lilley, J.S., Newton, G.W., Barker, J., Templar, J., Day, J.P., 1993. Effect of silicon on gastrointestinal absorption of aluminium. *Lancet* 342 (8865), 211–212.
- Elangovan, R., Ballance, S., White, K.N., McCrohan, C.R., Powell, J.J., 1999. Accumulation of aluminium by the freshwater crustacean *Asellus aquaticus* in neutral water. *Environ. Pollut.* 106 (3), 257–263.
- El-Rahman, S.S., 2003. Neuropathology of aluminium toxicity in rats (glutamate and GABA impairment). *Pharmacol. Res.* 47 (3), 189–194.
- Exley, C., 2004. The pro-oxidant activity of aluminium. *Free Radic. Biol. Med.* 36, 380–387.
- Exley, C., Begum, A., Woolley, M.P., Bloor, R.N., 2006. Aluminium in tobacco and cannabis and smoking-related disease. *Am. J. Med.* 119 (3), 276.e9–11.
- Exley, C., 2006. Aluminium and iron, but neither copper nor zinc, are key to the precipitation of beta-sheets of Aβ(42) in senile plaque cores in Alzheimer's disease. *J. Alzheimers Dis.* 10, 173–177.
- Exley, C., House, E.R., 2011. Aluminium in the human brain. *Montash Chem.* 142, 357–363.
- Fekete, V., Deconinck, E., Bolle, F., Van Loco, J., 2012. Modelling aluminium leaching into food from different foodware materials with multi-level factorial design of experiments. *Food Addit. Contam. Part A Chem Anal Control Expo Risk Assess* 29 (8), 1322–1333.
- Feng, L., Xiao, H., He, X., Li, Z., Li, F., Liu, N., Chai, Z., Zhao, Y., Zhang, Z., 2006. Long-term effects of lanthanum intake on the neurobehavioral development of the rat. *Neurotoxicol. Teratol.* 28 (1), 119–124.
- Feng, L., Xiao, H., He, X., Li, Z., Li, F., Liu, N., Zhao, Y., Huang, Y., Zhang, Z., Chai, Z., 2006a. Neurotoxicological consequence of long-term exposure to lanthanum. *Toxicol. Lett.* 165 (2), 112–120.
- Froment, D.P., Molitoris, B.A., Buddington, B., Miller, N., Alfrey, A.C., 1989. Site and mechanism of enhanced gastrointestinal absorption of aluminium by citrate. *Kidney Int.* 36 (6), 978–984.
- Frumkin, H., Gerberding, J.L., Keith, S., Jones, D., Rosemond, Z., Ingerman, L., Chapell, L., 2008. *Toxicological Profile for Aluminium*. U.S. Department of health and human services, Public Health Service Agency for Toxic Substances and Disease Registry. <http://www.atsdr.cdc.gov/toxprofiles/tp22.pdf>.
- Gauthier, E., Fortier, I., Courchesne, F., Pepin, P., Mortimer, J., Gauvreau, D., 2000. Aluminium forms in drinking water and risk of Alzheimer's disease. *Environ. Res.* 84 (3), 234–246.
- Gensemer, R.W., Playle, R.C., 1999. The bioavailability and toxicity of aluminium in aquatic environments. *Crit. Rev. Environ. Sci. Technol.* 29, 315–450.
- Goodman, W.G., 1985. Bone disease and aluminium: pathogenic considerations. *Am. J. Kidney Dis.* 6 (5), 330–335.
- Goopy, J.P., Murray, G.P., 2003. A review on the role of duckweed in nutrient reclamation and as a source of animal feed. *Asian Australas. J. Animal Sci.* 16 (2), 297–305.
- Goulet, R.R., Lalonde, J.D., Munger, C., Dupuis, S., Dumont-Frenette, G., Prémont, S., Campbell, P.G., 2005. Phytoremediation of effluents from aluminium smelters: a study of Al retention in mesocosms containing aquatic plants. *Water Res.* 39 (11), 2291–2300.
- Haque, N., Hughes, A., Lim, S., Vernon, C., 2014. Rare earth elements: overview of mining, mineralogy, uses, sustainability and environmental impact. *Resources* 3, 614–635.
- Hasan, M.R., Chakrabarty, R., 2009. *FAO Fisheries and Aquaculture Technical Paper*. No. 531. In: *Use of Algae and Aquatic Macrophytes as Feed in Small Scale Aquaculture - a Review*. FAO, Rome, p. 123.
- Han, S., Sakinci, U., Kose, S.K., Yazkan, R., 2004. The relationship between aluminium and spontaneous pneumothorax: treatment, prognosis, follow-up? *Interact. Cardiovasc. Thorac. Surg.* 3 (1), 79–82.
- Harris, W.R., Berthon, G., Day, J.P., Exley, C., Flaten, T.P., Forbes, W.F., Kiss, T., Orvig, C., Zatta, P.F., 1996. Speciation of aluminium in biological systems. *J. Toxicol. Environ. Health* 48 (6), 543–568.
- He, X., Zhang, Z., Zhang, H., Zhao, Y., Chai, Z., 2008. Neurotoxicological evaluation of long-term lanthanum chloride exposure in rats. *Toxicol. Sci.* 103 (2), 354–356.
- Herrmann, H., Nolde, J., Berger, S., Heise, S., 2016. Aquatic ecotoxicity of lanthanum – a review and an attempt to derive water and sediment quality criteria. *Ecotoxicol. Environ. Saf.* 124, 213–238.
- Hou, X.-H., Lamberts, L.V., Guan, G.-J., D'Haese, P.C., 2010. Increased exposure to aluminium: still a problem for chronic renal failure patients in East China? *Trace Elem. Electrolytes* 27 (1), 10–19.
- Huang, P., Li, J., Zhang, S., Chen, C., Han, Y., Liu, N., Xiao, Y., Wang, H., Zhang, M., Yu, Q., Liu, Y., Wang, W., 2011. Effects of lanthanum, cerium, and neodymium on the nuclei and mitochondria of hepatocytes: accumulation and oxidative damage. *Environ. Toxicol. Pharmacol.* 31 (1), 25–32.
- Huser, B.J., Futter, M., Lee, J.T., Perniel, M., 2016. In-lake measures for phosphorus control: the most feasible and cost-effective solution for long-term management of water quality in urban lakes. *Water Res.* 97, 142–152.
- Hutchison, A.J., Barnett, M.E., Krause, R., Kwan, J.T., Siami, G.A., SPD405-309 Lanthanum Study Group, 2008. Long-term efficacy and safety profile of lanthanum carbonate: results for up to 6 years of treatment. *Nephron Clin. Pract.* 110 (1), 15–23.
- Hutchison, A.J., Wilson, R.J., Garafola, S., Copley, J.B., 2016. Lanthanum carbonate: safety data after 10 years. *Nephrology (Carlton)* 21 (12), 987–999.
- Jochimsen, E.M., Carmichael, W.W., An, J.S., Cardo, D.M., Cookson, S.T., Holmes, C.E., Antunes, M.B., de Melo Filho, D.A., Lyra, T.M., Barreto, V.S., Azevedo, S.M., Jarvis, W.R., 1998. Liver failure and death after exposure to microcystins at a hemodialysis center in Brazil. *N. Engl. J. Med.* 338 (13), 873–878.
- Johannesson, K.H., Lyons, W.B., 1994. The rare earth element geochemistry of Mono Lake water and the importance of carbonate complexing. *Limnol. Oceanogr.* 39, 1141–1154.
- Jones, K.C., Bennett, B.G., 1986. Exposure of man to environmental aluminium – an exposure commitment assessment. *Sci. Total Environ.* 52, 65–82.
- Jouhannneau, P., Lacour, B., Raisbeck, G., Yiou, F., Banide, H., Brown, E., Drüeke, T., 1993. Gastrointestinal absorption of aluminium in rats using ²⁶Al and accelerator mass spectrometry. *Clin. Nephrol.* 40 (4), 244–248.
- Kawahara, M., Kato-Negishi, M., 2011. Link between aluminium and the pathogenesis of Alzheimer's disease: the integration of the aluminium and amyloid cascade hypotheses. *Int. J. Alzheimer's Dis.* 8, 276393.
- Kongerud, J., Søyseth, V., 2014. Respiratory disorders in aluminium smelter workers. *J. Occup. Environ. Med.* 56 (Suppl. 5), 60–70.
- Krewski, D., Yokel, R.A., Nieboer, E., Borchelt, D., Cohen, J., Harry, J., Kacew, S., Lindsay, J., Mahfouz, A.M., Rondeau, V., 2014. Human health risk assessment for aluminium, aluminium oxide, and aluminium hydroxide. *J. Toxicol. Environ. Health B Crit. Rev.* 10 (Suppl. 1), 1–269.
- Kumar, V., Bal, A., Gill, K.D., 2009. Aluminium-induced oxidative DNA damage recognition and cell-cycle disruption in different regions of rat brain. *Toxicology* 264, 137–144.
- Landman, M., Ling, N., 2006. Lake Okareka and Tikitapu Fish Health Monitoring. Landman, M., Brijis, J., Glover, C., Ling, N., 2007. Lake Okareka and Tikitapu Fish Health Monitoring. Scion report.
- LaZerte, B.D.G., van Loon, G., Anderson, B., 1997. Aluminium in water. In: Yokel, R.A., Golub, M.S. (Eds.), *Research Issues in Aluminium Toxicity*. Taylor & Francis, Washington DC, USA, pp. 17–45.
- Lemire, J., Mailloux, R., Puisseux-Dao, S., Appanna, V.D., 2009. Aluminium-induced defective mitochondrial metabolism perturbs cytoskeletal dynamics in human astrocytoma cells. *J. Neurosci. Res.* 87, 1474–1483.
- Lewandowski, I., Schausser, I., Hupfer, M., 2003. Long term effects of phosphorus precipitations with alum in hypereutrophic Lake Susser See (Germany). *Water Res.* 37 (13), 3194–3204.
- Liu, X., Byrne, R.H., 1997. Rare earth and yttrium phosphate solubilities in aqueous solution. *Geochem. Cosmochim. Acta* 61 (8), 1625–1633.
- Lürling, M., Tolman, Y., 2010. Effects of lanthanum and lanthanum-modified clay on growth, survival and reproduction of *Daphnia magna*. *Water Res.* 44 (1), 309–319.
- Lürling, M., van Oosterhout, 2013. Controlling eutrophication by combined bloom precipitation and sediment phosphorus inactivation. *Water Res.* 47, 6527–6537.
- Lürling, M., Mackay, E., Reitzel, K., Spears, B.M., 2016. Editorial - a critical perspective on geo-engineering for eutrophication management in lakes. *Water Res.* 97, 1–10.
- Mailloux, R.J., Lemire, J., Appanna, V.D., 2011. Hepatic response to aluminium toxicity: dyslipidemia and liver diseases. *Exp. Cell Res.* 317 (16), 2231–2238.
- Martell, A.E., Motekaitis, R.J., 1989. Coordination chemistry and speciation of Al(III) in aqueous solution. In: Lewis, T.E. (Ed.), *Environmental Chemistry and Toxicology of Aluminium*. Lewis Publishers, Chelsea, MI, pp. 3–17.
- Martyn, C.N., Barker, D.J., Osmond, C., Harris, E.C., Edwardson, J.A., Lacey, R.F., 1989. Geographical relation between Alzheimer's disease and aluminium in drinking water. *Lancet* 14 (1), 59–62.
- Meis, S., Spears, B.M., Maberly, S.C., O'Malley, M.B., Perkins, R.G., 2012. Sediment amendment with Phoslock (R) in Clatto Reservoir (Dundee, UK): investigating changes in sediment elemental composition and phosphorus fractionation. *J. Environ. Manag.* 185–193.
- Moermond, C.T., Tjink, J., van Wezel, A.P., Koelmans, A.A., 2001. Distribution, speciation, and bioavailability of lanthanides in the Rhine-Meuse estuary, The Netherlands. *Environ. Toxicol. Chem.* 20 (9), 1916–1926.
- Mujika, J.I., Escribano, B., Akhmatkaya, E., Ugalde, J.M., Lopez, X., 2012. Molecular dynamics simulations of iron- and aluminium-loaded serum transferrin: protonation of tyr188 is necessary for prompt metal release. *Biochemistry* 51, 7017–7027.
- National Industrial Chemicals Notification and Assessment Scheme (NICNAS), 2014. Phoslock. Existing Chemical Secondary Notification Assessment Report NA/8995. GPO Box 58, Sydney NSW 2001 AUSTRALIA, ISBN 978-0-9874434-1-0.
- Nieboer, E., Gibson, B.L., Oxman, A.D., Kramer, J.R., 1995. Health effects of aluminium: a critical review with emphasis on aluminium in drinking water. *Environ. Res.* 3, 29–81.
- Niogi, S., Wood, C.M., 2004. Biotic ligand model, a flexible tool for developing site-specific water quality guidelines for metals. *Environ. Sci. Technol.* 38 (23), 6177–6192.
- Pagano, G., Guida, M., Tommasi, F., Oral, R., 2015. Health effects and toxicity mechanisms of rare earth elements-Knowledge gaps and research prospects. *Ecotoxicol. Environ. Saf.* 115, 40–48.
- Parry, R., Plowman, D., Delves, H.T., Roberts, N.B., Birchall, J.D., Bellia, J.P., Davenport, A., Ahmad, R., Fahal, I., Altmann, P., 1998. Silicon and aluminium interactions in haemodialysis patients. *Nephrol. Dial. Transplant.* 13 (7), 1759–1762.
- Paul, W.J., Hamilton, D.P., Gibbs, M.M., 2008. Low-dose alum application trialled as a

- management tool for internal nutrient loads in Lake Okaro, New Zealand. *N. Z. J. Mar. Freshw. Res.* 42, 207–217.
- Pennick, M., Dennis, K., Damment, S.J., 2006. Absolute bioavailability and disposition of lanthanum in healthy human subjects administered lanthanum carbonate. *J. Clin. Pharmacol.* 46 (7), 738–744.
- Perkins, R.G., Underwood, G.J.C., 2001. The potential for phosphorus release across the sediment-water interface in a eutrophic reservoir dosed with ferric sulphate. *Water Res.* 35, 1399–1406.
- Praticò, D., Uryu, K., Sung, S., Tang, S., Trojanowski, J.Q., Lee, V.M., 2002. Aluminium modulates brain amyloidosis through oxidative stress in APP transgenic mice. *FASEB J.* 16 (9), 1138–1140.
- Raghu, G., Collins, B.F., Xia, D., Schmidt, R., Abraham, J.L., 2014. Pulmonary fibrosis associated with aluminium trihydrate (Corian) dust. *N. Engl. J. Med.* 370 (22), 2154–2156.
- Redling, K., 2006. Rare Earth Elements in Agriculture. With Emphasis on Animal Husbandry. PhD-Thesis. Ludwig-Maximilians-Universität München, Germany.
- Reitzel, K., Hansen, J., Andersen, F.O., Hansen, K.S., Jensen, H.S., 2005. Lake restoration by dosing aluminium relative to mobile phosphorus in the sediment. *Environ. Sci. Technol.* 39 (11), 4134–4140.
- Reitzel, K., Jensen, H.S., Egemose, S., 2013. pH dependent dissolution of sediment aluminium in six Danish lakes treated with aluminium. *Water Res.* 47, 1409–1420.
- Reitzel, K., Andersen, F.O., Egemose, S., Jensen, H.S., 2013a. Phosphate adsorption by lanthanum modified bentonite clay in fresh and brackish water. *Water Res.* 47 (8), 2787–2796.
- Richter, H., 2003. Hinweise zur Toxikologie von Seltenen Erden. In: XVI Tage der Seltenen Erden. Berlin, Germany, p. 18.
- Rob, P.M., Niederstadt, C., Reusche, E., 2001. Dementia in patients undergoing long-term dialysis: aetiology, differential diagnoses, epidemiology and management. *CNS Drugs* 15 (9), 691–699.
- Robb, M.B., Greenop, B., Goss, Z., Douglas, G., Adeney, J., 2003. Application of Phoslock™, an innovative binding clay, to two Western Australian waterways: preliminary findings. *Hydrobiologia* 494, 237–243.
- Rondeau, V., Commenges, D., Jacqmin-Gadda, H., Dartigues, J.F., 2000. Relation between aluminium concentrations in drinking water and Alzheimer's disease: an 8-year follow-up study. *Am. J. Epidemiol.* 152 (1), 59–66.
- Rondeau, V., Jacqmin-Gadda, H., Commenges, D., Helmer, C., Dartigues, J.F., 2009. Aluminium and silica in drinking water and the risk of Alzheimer's disease or cognitive decline: findings from 15-year follow-up of the PAQUID cohort. *Am. J. Epidemiol.* 169 (4), 489–496.
- Rose, E.A., Porcerelli, J.H., Neale, A.V., 2000. Pica: common but commonly missed. *J. Am. Board Fam. Pract.* 13 (5), 353–358 submitted for publication.
- Ross, G., Haghseresht, F., Cloete, T.M., 2008. The effect of pH and anoxia on the performance of Phoslock, a phosphorus binding clay. *Harmful Algae* 7, 545–550.
- Rydin, E., Welch, E.B., 1998. Aluminium dose required to inactivate phosphate in lake sediments. *Water Res.* 32 (10), 2969–2976.
- Rydin, E., Huser, B., Welch, E.B., 2000. Amount of phosphorus inactivated by alum treatments in Washington lakes. *Limnol. Oceanogr.* 45 (1), 226–230.
- Shimomura, H., Tsutsumi, S., Mizobuchi, J., Takuma, S., Nakagawa, K.I., Asai, Y., 1980. Experimental studies on acute toxicity of rare earth elements. *Shikwa Gakuho* 80 (9), 1303–1307 (Japanese).
- Schintu, M., Meloni, P., Contu, A., 2000. Aluminium fractions in drinking water from reservoirs. *Ecotoxicol. Environ. Saf.* 46, 29–33.
- Shirley, D.G., Lote, C.J., 2005. Renal handling of aluminium. *Nephron. Physiol.* 101 (4), 99–103.
- Simoes, J., Barata, J.D., D'Haese, P.C., De Broe, M.E., 1994. Cela n'arrive qu'aux autres (aluminium intoxication only happens in the other nephrologist's dialysis center). *Nephrol. Dial. Transplant.* 9 (1), 67–68.
- Sińczuk-Walczak, H., Szymczak, M., Raźniewska, G., Matczak, W., Szymczak, W., 2003. Effects of occupational exposure to aluminium on nervous system: clinical and electroencephalographic findings. *Int. J. Occup. Med. Environ. Health* 16 (4), 301–310.
- Slatopolsky, E., Liapis, H., Finch, J., 2005. Progressive accumulation of lanthanum in the liver of normal and uremic rats. *Kidney Int.* 68 (6), 2809–2813.
- Smans, K.A., D'Haese, P.C., Van Landeghem, G.F., Andries, L.J., Lamberts, L.V., Hendy, G.N., De Broe, M.E., 2000. Transferrin-mediated uptake of aluminium by human parathyroid cells results in reduced parathyroid hormone secretion. *Nephrol. Dial. Transplant.* 15 (9), 1328–1333.
- Smeltzer, E., Kirn, R.A., Fiske, S., 1999. Long-term water quality and biological effects of alum treatment of lake Morey, Vermont. *Lake Reserv. Manag.* 15 (3), 173–184.
- Spears, B.M., Lürling, M., Yasserli, S., Castro-Castellone, A.T., Gibbs, M., Meisa, S., McDonald, C., McIntosh, J., Sleep, D., van Oosterhout, F., 2013. Lake responses following lanthanum-modified bentonite clay (Phoslock®) application: an analysis of water column lanthanum data from 16 case study lakes. *Water Res.* 47 (15), 5930–5942.
- Stevenson, F., Vance, G., 1989. Naturally occurring aluminium-organic complexes. In: Sposito, G. (Ed.), *The Environmental Chemistry of Aluminium*. CRC Press, Boca Raton, FL, pp. 117–147.
- van Ginkel, M.F., van der Voet, G.B., D'Haese, P.C., De Broe, M.E., de Wolff, F.A., 1993. Effect of citric acid and maltol on the accumulation of aluminium in rat brain and bone. *J. Lab. Clin. Med.* 121 (3), 453–460.
- van Ginkel, M.F., 2001. Aluminium Metabolism and Neurotoxicity: a Role for Citrate. PhD thesis. University, Leiden The Netherlands.
- Van Landeghem, G.F., D'Haese, P.C., Lamberts, L.V., De Broe, M.E., 1994. Quantitative HPLC/ETAAS hybrid method with an on-line metal scavenger for studying the protein binding and speciation of aluminium and iron. *Anal. Chem.* 66 (2), 216–222.
- Van Landeghem, G.F., 1997. Aluminium Speciation in Biological Fluids. PhD Thesis. Universities of Antwerp, (Belgium)/Leiden (The Netherlands), ISBN 90-9010801-7.
- Van Landeghem, G.F., D'Haese, P.C., Lamberts, L.V., De Broe, M.E., 1997. Competition of iron and aluminium for transferrin: the molecular basis for aluminium deposition in iron-overloaded dialysis patients? *Exp. Nephrol.* 5 (3), 239–245.
- Van Landeghem, G.F., D'Haese, P.C., Lamberts, L.V., Barata, J.D., De Broe, M.E., 1997a. Aluminium speciation in cerebrospinal fluid of acutely aluminium-intoxicated dialysis patients before and after desferrioxamine treatment; a step in the understanding of the element's neurotoxicity. *Nephrol. Dial. Transplant.* 12 (8), 1692–1698.
- Van Landeghem, G.F., de Broe, M.E., D'Haese, P.C., 1998. Al and Si: their speciation, distribution, and toxicity. *Clin. Biochem.* 31 (5), 385–397.
- Van Landeghem, G.F., D'Haese, P.C., Lamberts, L.V., Djukanovic, L., Pejanovic, S., Goodman, W.G., De Broe, M.E., 1998a. Low serum aluminium values in dialysis patients with increased bone aluminium levels. *Clin. Nephrol.* 50 (2), 69–76.
- van Oosterhout, F., Goitom, E., Roessink, I., Lürling, M., 2014. Lanthanum from a modified clay used in eutrophication control is bioavailable to the marbled crayfish (*Procambarus fallax f. virginalis*). *PLoS One* 28, e102410, 9(7).
- Verbueken, A.H., Van de Vyver, F.L., Van Grieken, R.E., Paulus, G.J., Visser, W.J., D'Haese, P., De Broe, M.E., 1984. Ultrastructural localization of aluminium in patients with dialysis-associated osteomalacia. *Clin. Chem.* 30 (5), 763–768.
- Verweij, W., 2013. CHEAQS, a Program for Calculating Chemical Equilibria in Aquatic Systems, PRO Version 2013.1.
- Waajen, G., van Oosterhout, F., Douglas, G., Lürling, M., 2016. Management of eutrophication in Lake De Kuil (The Netherlands) using combined flocculant - lanthanum modified bentonite treatment. *Water Res.* 15 (97), 83–95.
- Waajen, G., van Oosterhout, F., Douglas, G., Lürling, M., 2016a. Geo-engineering experiments in two urban ponds to control eutrophication. *Water Res.* 15 (97), 69–82.
- Waajen, G., van Oosterhout, F., Lürling, M., 2017. Bio-accumulation of lanthanum from lanthanum modified bentonite treatments in lake restoration. *Environ. Pollut.* 230, 911–918.
- Ward, M.K., Feast, T.G., Ellis, H.A., Parkinson, I.S., Kerr, D.N., 1978. Osteomalacic dialysis osteodystrophy: evidence for a water-borne aetiological agent, probably aluminium. *Lancet* 22 (1), 841–845.
- Wauer, G., Teien, H.C., 2010. Risk of acute toxicity for fish during aluminium application to hardwater lakes. *Sci. Total Environ.* 408, 4020–4025.
- Welch, E.B., Cooke, G.D., 1999. Effectiveness and longevity of phosphorus inactivation with alum. *J. Lake Reserv. Manag.* 15, 5–27.
- Welch, E.B., De Gasperi, C.L., Spyridakis, D.E., Belnick, T.J., 1988. Internal phosphorus loading and alum effectiveness in shallow lakes. *Lake Reserv. Manag.* 4, 27–33.
- Willhite, C.C., Karyakina, N.A., Yokel, R.A., Yenugadhathi, N., Wisniewski, T.M., Arnold, I.M., Momoli, F., Krewski, D., 2014. Systematic review of potential health risks posed by pharmaceutical, occupational and consumer exposures to metallic and nanoscale aluminium, aluminium oxides, aluminium hydroxide and its soluble salts. *Crit. Rev. Toxicol.* 44 (Suppl. 4), 1–80.
- Wills, M.R., Savory, J., 1985. Water content of aluminium, dialysis dementia, and osteomalacia. *Environ. Health Perspect.* 63, 141–147.
- Xu, X., Zhu, W., Wang, Z., Witkamp, G.-J., 2003. Accumulation of rare earth elements in maize plants (*Zea mays* L.) after application of mixtures of rare earth elements and lanthanum. *Plant Soil* 252, 267–277.
- Xu, Q., Fu, Y., Min, H., Cai, S., Sha, S., Cheng, G., 2012. Laboratory assessment of uptake and toxicity of lanthanum (La) in the leaves of *Hydrocharis dubia* (Bl.) Backer. *Environ. Sci. Pollut. Res.* 19, 3950–3958.
- Yang, X., Yin, D., Sun, H., Wang, X., Dai, L., Chen, Y., Dai, L., Chen, Y., Cao, M., 1999. Distribution and bioavailability of rare earth elements in aquatic microcosm. *Chemosphere* 39 (14), 2443–2445.
- Yang, Z., Schryvers, D., Roels, F., D'Haese, P.C., De Broe, M.E., 2006. Demonstration of lanthanum in liver cells by energy-dispersive X-ray spectroscopy, electron energy loss spectroscopy and high-resolution transmission electron microscopy. *J. Microsc.* 223 (2), 133–139.
- Yang, J., Liu, Q., Wu, S., Xi, Q., Cai, Y., 2013. Effects of lanthanum chloride on glutamate level, intracellular calcium concentration and caspases expression in the rat hippocampus. *Biometals* 26 (1), 43–59.
- Yokel, R.A., McNamara, P.J., 2001. Aluminium toxicokinetics: an updated minireview. *Pharmacol. Toxicol.* 88 (4), 159–167.
- Yoon, H.K., Moon, H.S., Park, S.H., Song, J.S., Lim, Y., Kohyama, N., 2005. Dendritic pulmonary ossification in patient with rare earth pneumoconiosis. *Thorax* 60 (8), 701–703.
- Zarros, A., Byrne, A.M., Boomkamp, S.D., Tsakiris, S., Baillie, G.S., 2013. Lanthanum-induced neurotoxicity: solving the riddle of its involvement in cognitive impairment? *Arch. Toxicol.* 87 (11), 2031–2035.
- Zatta, T.K.P., Suwalsky, M., Berthon, G., 2002. Aluminium(III) as a promoter of cellular oxidation. *Coord. Chem. Rev.* 228, 271–284.
- Zhao, H., Hong, J., Yu, X., Zhao, X., Sheng, L., Ze, Y., Sang, X., Gui, S., Sun, Q., Wang, L., Hong, F., 2013. Oxidative stress in the kidney injury of mice following exposure to lanthanides trichloride. *Chemosphere* 93 (6), 875–884.